

**Evaluation of Partially Converted Lines from the
Sorghum Conversion Program to Determine
Combining Ability and Heterosis for Early Testing**

by

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Abstract

Hybrid breeding technology has been very successful in increasing yield of many cereal crops including sorghum (*Sorghum bicolor* (L.) Moench). The process of developing and evaluating the performance of hybrids is the most expensive and time-consuming activity. In sorghum, developing parental inbred lines and evaluating their potential hybrid performance is very expensive and time-consuming. Predicting hybrid performance in any way possible might help reduce the number of crosses needed and evaluated. The study was comprised of 40 lines, selected by the percentage of exotic genome recovered and the generation (BCF_{2:3} high, BCF_{2:3} low, F_{2:3} high, and F_{2:3} low), and four elite testers. The 160 F₁ hybrids and parents were evaluated in a randomized complete block design with two replications for combining ability and heterosis during the 2015-2016 cropping season. The results indicated that sufficient genetic variability was observed for all characters evaluated. Analysis of general combining ability (GCA) revealed that among the parents, lines RSC117-4 BCF_{2:3} high, RSC83-1 BCF_{2:3} low, RSC83-14 BCF_{2:3} high, and RSC112-19 BCF_{2:3} high, and the testers A.301 and A.319 were promising general combiners for increasing grain yield and most other important traits. Therefore, the parents can be used in sorghum breeding programs aimed toward developing high-yielding hybrids. A.301*RSC83-14 BCF_{2:3} high produced the third largest overall mean grain yield for hybrids, yielding slightly more than commercial check 301/41, with 67.3 days to anthesis and 127.5 cm total plant height. The hybrid had

significant midparent heterosis (MPH) with a positive specific combining ability (SCA) effect and GCA combining effect combination of high x high which provides complementary gene action. The hybrid had good combination for earliness, dwarfing genes, and large yield. A.319*RSC83-1 F_{2:3} high had the largest overall mean for grain yield, out yielding four of the best six commercial check hybrids, with 72.2 days to anthesis and 153.0 cm total plant height, with a positive MPH and SCA effect with high x low GCA effects combination indicating additive x dominance type of gene interaction. The cross could produce desirable transgressive segregates because of the additive genetic system in one general combiner of the parent and complementary epistatic effects in the other. The hybrid was taller and yielded more, with heavier panicle weights and 1000-kernel weight, and significantly greater MPH *per se* parents. Line x tester analysis revealed that the contribution of the lines to the total sum of squares was greater than testers in all studied traits except the number of days to anthesis. The variance due to GCA (σ^2_{gca}) was less than for SCA (σ^2_{sca}) for all traits except plant exsertion and concentration of protein, starch, fiber, and fat in grain, suggesting preponderance of non-additive gene action controlling the characters. Dominance variance (σ^2_D) was greater than additive variance (σ^2_A) for all traits except plant exsertion and concentration of protein, starch, fiber, and fat in the grain. The results are supported by the ratio of variance of general to specific combining ability ($\sigma^2_{gca} / \sigma^2_{sca}$) which was smaller than unity and by the degree of dominance (σ^2_D / σ^2_A) that uses values greater than unity for all traits except plant exsertion where σ^2_A was larger than σ^2_D , with the degree of dominance being less than unity. Selections based on the predominance of dominance variance suggested the exploitation of heterosis.

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Chapter I

Introduction

Plant breeding is driven by the need to continually increase yield while improving the ability of crop plants to be sustainable across variable environments. Increasing yield of cereal grains is an important aspect of “2050 - feeding the nine billion people” because it will require increasing food production by 70% during the next 33 years (FAO, 2011).

As the amount of water in the Ogallala Aquifer continues to decrease, there is an increased need to improve agricultural production under limited water or dryland conditions with minimal inputs. Sorghum [*Sorghum bicolor* (L.) Moench] because of its drought tolerance is important in semi-arid regions. Sorghum has been cultivated in arid and semi-arid regions of Africa for thousands of years. Sorghum is the fifth-most important cereal crop in the world and the third-most important in the United States, after maize, *Zea mays* L., and wheat, *Triticum aestivum* L. (FAS/USDA, 2012). Increasing global food demand and decreasing water, coupled with potential effects of climate change, push plant breeders to epic heights.

Worldwide, 66.2 million Mg of sorghum were produced during the 2010-2011 trade year (September through August), with the United States being the second largest producer in the world, behind Nigeria. Worldwide sorghum production increased to 67.8 million Mg during the 2014-2015 trade year (September through August), with the United States leading world production, followed by Mexico and then Nigeria

(FAS/USDA, 2015). In 2010, more than 7.5 million Mg of sorghum worth \$1.7 billion were harvested in the United States (USDA, 2011). The United States, with 2.4 million Mg in trade year 2010-2011, is the largest exporter of grain sorghum in the world, while Mexico and Japan were the leading importers of sorghum (FAS/USDA, 2011). Chief importers during the 2014-2015 U.S. crop year (September through August) were China (97%) and Japan (2%) (U.S. Grains, 2016).

Sorghum has a high yield potential, and the greatest recorded yield of 20.1 tons per hectare for the crop was in Texas in the 1980s (Boyer, 1987). However, yields in Africa and India remain low. Access to modern machinery, improved technology, and irrigation, along with sorghum breeding efforts through hybrid production in developed countries might, in part, explain the vast difference in yield in developed countries compared to that in developing countries.

By 1905, the Texas Agricultural Experiment Station and USDA began research at Chillicothe, Texas that focused mainly on creating plants that were short statured for mechanical harvesting, early maturing, and higher yielding than varieties introduced from the tropics (Rooney and Smith, 2000). Dwarf sorghum cultivars were created by making three of the four independent height-gene alleles (Dw1 through Dw4) recessive (three-gene dwarfs). Maturity in grain sorghum was shortened by using six gene loci (Ma1 through Ma6) that can be manipulated to adapt the plants to shorter growing seasons (Quinby and Karper, 1954; Poehlman, 1987; Rooney and Aydin, 1999). Grain yield potential in the United States has increased markedly since the introduction of sorghum

in the 1800s, and the increase is attributed to development of hybrids and short-statured plants that yield well in different environments.

Commercial production of sorghum seed relies on development of hybrids. In sorghum, a mostly self-pollinated crop, development of hybrids depends on cytoplasmic-genetic male sterility, in which self-pollination is averted by using male-sterile lines as females, thus avoiding laborious emasculation by hand, yet enabling crossing between lines as if the crop had been cross-pollinated (Quinby and Martin, 1954; Stephens and Holland, 1954). Production of hybrid sorghum seed has evolved into a system where introduced germplasm can be used either as a male parent or a female parent. This classification depends on the presence or absence of fertility-restoring genes. If the line possesses fertility-restoring genes, it is designated an R line (restorer line) that can be used as a male; otherwise, it is designated a B line and can be sterilized by backcrossing with a male sterile designated as an A line. The system of hybrid development has caused sorghum breeding programs to develop two breeding groups: a male-parent group (R line/fertility-restorer) and a female-parent group (an A/B line, lacking the fertility-restoring gene of the A1 male-sterility system). New germplasm is usually placed in one of the two groups based on whether or not it possesses fertility-restoring genes. Stephens and Holland (1954) discovered the cytoplasmic male sterility (CMS) system that is dependent on the presence of male-sterile cytoplasm and nuclear fertility-restoring genes.

Heterosis or hybrid vigor is defined as the difference between a hybrid and the mean of the two parents (Falconer and Mackay, 1996). Exploitation of heterosis began in the United States in the 1950s, resulting in a dramatic increase in yields for maize. The

more genetically divergent a hybrid is, based on genetic relatedness of the parental lines, the greater the degree of heterosis. Much sorghum research and advancement to date have been based on phenotypic classification. Grain yield is a complex trait controlled by polygenes and has low heritability especially in stressful environments. For example, selection for grain yield under severe drought stress has often been considered inefficient because the estimate of heritability of grain yield has been observed to decrease with reduced yield (Bolaños and Edmeades, 1993).

Such traits as grain yield and its components are governed by polygenes with complex gene action. Hence, a comprehensive understanding of the nature and magnitude of gene action and knowledge of the combining ability of parents to develop new hybrids through suitable breeding methods were essential for implementing a systematic crop improvement program. Combining ability analysis is a powerful tool to estimate combining ability effects and aids in selecting desirable parents and crosses for exploitation of heterosis and involving them in production of desirable hybrids and segregates (Sarker et al., 2002; Rashid et al., 2007). Combining ability is useful for plant breeders to better understand genetic variance and inbred lines to identify desirable parents to use in commercial hybrid production. Plant breeders use results of research on combining ability to help select the best parents for development of hybrids or varieties. The concept of general and specific combining ability was introduced by Sprague and Tatum (1942) who designated general combining ability (GCA) as the average performance of a line in hybrid combination. The term specific combining ability (SCA) was applied to cases where certain hybrid combinations did relatively better or worse

than would be expected on the basis of the average performance of the lines involved. Commercial production of hybrids, however, depends upon two factors: the behavior of a line itself and the behavior of a line in hybrid combination assessed through the estimation of GCA and SCA effects. The importance of both GCA and SCA has been reported by Kambal and Webster (1965). GCA is the result of additive gene effect, while SCA is considered to be composed of non-allelic interaction.

The line x tester (L x T) mating design for combining ability suggested by Kempthorne (1957) is an appropriate method to identify superior parents and hybrids based on GCA and SCA, respectively. It is also helpful for assessing the nature and magnitude of gene action controlling quantitative traits. Information on combining ability and heterosis is a valuable tool in determining superior parents and hybrid combinations in a hybrid breeding program.

Correlation studies provide information on the association of yield with its component characters and help to formulate a selection index. Because yield is a complex quantitative character, it tends to be dependent on various component characters, and path analysis reveals the direct and indirect effects of component traits on grain yield.

The purpose of the present investigation was to obtain information on combining ability of sorghum lines for traits of economic value. The overall goal of this research was to determine if there was additional benefit for sorghum breeders to increase efficiency by decreasing the workload of the breeding process, thus allowing quicker

release of newly cultivated sorghum hybrids. The primary objective of many plant breeding programs is to increase yield and its components to improve the quality (grain yield and its components including plant height) of crop plants. The objective of this study was to use the recovered genetic proportion of exotic sorghum genotypes from different taxonomic groups through the evaluation of F_1 hybrids produced from exotic germplasm in the generation of $F_{2:3}$ or $BC_1F_{2:3}$ to determine if there was additional benefit from backcrossing to an exotic parent in the USDA ARS Sorghum Conversion Program. Considering this, the investigation entitled “Evaluation of Partially Converted Lines from the Sorghum Conversion Program to Determine Combining Ability and Heterosis for Early Testing” was undertaken with the following objectives to determine:

1. if a difference exists between high and low percentage of recovery for the genetic proportion recovered,
2. if a difference exists between the $F_{2:3}$ and $BC_1F_{2:3}$ for high and low percentage of recovery for the genetic proportion recovered,
3. effects of high and low percentage of recovery for genetic similarity to the recurrent parent across different levels of genetics and maturities, across different environments, and across the taxonomic groups used in this study,
4. effects of combining ability across different females, with different genetics and maturity, within the Reinstated Sorghum Conversion (RSC) families and generation selections,
5. effects of midparent heterosis (MPH) across different females, with different genetics and maturities, within a RSC population,

6. correlations among grain yield and yield components and to understand their direct and indirect effects on grain yield, and
7. determine the benefits of early testing in the F₂ population

This should provide insight into combining ability of selected males across different levels of maturity for elite females in a current breeding program. The information will be useful to sorghum breeders who can more efficiently and systematically exploit heterosis through selection of high combining ability based on the parents used.

Understanding combining ability is crucial for sorghum breeders to make much-needed advances in development of new hybrids. At present, yield potential of sorghum has reached a plateau, where increases in yield are minimal.

Chapter II

Review of Literature

Texas Panhandle and the Ogallala Aquifer

As the amount of water in the Ogallala Aquifer on the Great Plains of the United States continues to decrease, improved crop production under limited water or dryland conditions with minimal input is required. Producers will be making difficult decisions to grow crops that more efficiently use water with the decreasing amount of water in the Aquifer. Understanding the impact of the decreased availability of water in the Ogallala Aquifer increases the need to identify higher yielding food and feed crops that can be produced with limited irrigation or dryland conditions on the Texas High Plains.

The Ogallala Aquifer, one of the largest fresh-water aquifers in the world, underlies parts of the states of South Dakota, Nebraska, Wyoming, Colorado, Kansas, Oklahoma, New Mexico, and Texas (Colaizzi et al., 2008). The Ogallala Aquifer is the main supplier of irrigation for agricultural production on the Texas High Plains, with 95% of the water pumped from the Aquifer used for irrigation (Conkwright, 2012; Dudensing et al., 2008). Colaizzi et al. (2008) stated that almost all irrigation water used on the Texas High Plains is from the Ogallala Aquifer. Infiltration and evaporation are key components to recharge the Aquifer, but currently, water is being removed faster than the natural recharge rate, which varies by region. Discharge from the Ogallala Aquifer occurs through natural outlets such as seeps, springs, and leakage to underlying

geological formations (Nativ and Smith, 1987). Thus far, withdrawal through irrigation is currently the most significant discharge component. Pumping from the Ogallala Aquifer started in 1911 and increased drastically after World War II (Nativ and Smith, 1987). Intensive irrigation on the Texas High Plains began in the 1930s and was facilitated mostly by the development of the internal combustion engine, turbine pumps, and rotary well drilling (Musick et al., 1988; Musick and Walker, 1987). The amount of water currently being withdrawn from the Aquifer is much greater than the annual recharge by precipitation. Annual recharge ranges from 0.15 to 2.1 cm yr⁻¹ (0.058 to 0.833 in yr⁻¹) (Knowles et al., 1984), while Nativ and Smith (1987) reviewed articles for the Ogallala Aquifer south of the Canadian River on the Southern High Plains of Texas and New Mexico and found that annual removal by pumping ranged from 16.6 to 113 cm year⁻¹ (0.5 to 6.55 in yr⁻¹). The Texas Panhandle is one of the areas where the amount of annual withdrawal exceeds the recharge rate. The sustainability of current agroecosystems depends on irrigation water on the Texas High Plains. Irrigation increases yield potential two to seven times and reduces yield risk by 75 to 90% over non-irrigated agricultural production (Lust et al., 2009). The increasing interest in agricultural production with limited irrigation (Norwood, 2000), coupled with producers seeking alternative crops that could reduce water consumption and lengthen the utility of the Aquifer (Howell et al., 2004), have become two of the agricultural priorities on the Southern High Plains.

Carpenter et al. (2011) reported that agricultural activities account for 76% of the global fresh water used by humans. Because water is becoming a scarce commodity

through erratic rainfall patterns, the water in aquifers is being depleted, and agricultural commodities are being produced in marginal areas to satiate the demand of the escalating number of humans in the world (Hanjra and Qureshi, 2010). To maintain or increase agricultural output, a more efficient water management system is needed to maximize crop water-use efficiency -- the concept of evaluating agricultural output based on the amount of water consumed rather than land area occupied (Bennett, 2003).

Sorghum History and Characteristics

Sorghum is a self-pollinating monocot in the grass family Poaceae. It is thought to have originated in the Ethiopia-Sudan region of northeastern Africa (Doggett, 1998; FAO, 1995), then spread throughout Africa, Southeast Asia, India, Australia, and the United States (FAO, 2007). Broomcorn might have been introduced into the United States by Benjamin Franklin, as early as 1757 (Kimber, 2000; Smith and Frederiksen, 2000). Grain sorghum was first carried from West Africa to the United States with the slave trade, when guinea corn and probably chicken corn were introduced (Doggett, 1970; Smith and Frederiksen, 2000). Introductions from North Africa included brown and white durras in 1874, milo about 1880, feterita in 1906, and hegari in 1908 (Doggett, 1970; Dillons et al., 2007).

Originally intended to feed slaves, sorghum continues to be the staple food of many of the poorest people in developing countries of the world (Rooney and Smith, 2000). Sorghum also has evolved as a primary feed source for livestock and is used in ethanol production in developed countries. This versatile crop is grown primarily in the semiarid regions of Africa, India, China, South America, and the United States. Grain

yields are significantly less in Africa and India, averaging 300-2,000 kg ha⁻¹ under rain-fed conditions, compared to 4,500-6,500 kg ha⁻¹ of hybrids grown under irrigated conditions in the United States (FAO, 2007).

Worldwide, sorghum ranks fifth behind maize, rice (*Oryza sativa* L.), wheat, and barley (*Hordeum vulgare* L.) with approximately 825, 680, 650, 150, and 60 million Mg of grain produced on 160, 162, 226, 54, and 40 million hectares, respectively (FAO, 2011; Grains, 2011; USDA-ERS, 2011). Sorghum is the third most-important cereal crop in the United States and the fifth most-important cereal crop in the world (Grains, 2007-2008). Assefa et al. (2010) and Assefa and Staggenborg (2010) reported that more than 80% of sorghum in the world is cultivated as a dryland crop because of its drought tolerance.

Sorghum is a frost-sensitive, short-day plant that grows on a wide range of soils at altitudes from sea level to 3,000 m (Kimber, 2000; FAO, 2007). As a C₄ species, sorghum has greater transpiration efficiency and hence survives and grows better than most other cereal crops under water-stress conditions (Doggett, 1988; Rooney, 2004). Sorghum is well known for its ability to tolerate limited water and produce during periods of extended drought that deter production of most other grain crops. Several factors that can limit sorghum yields include: prolonged drought or delayed rainfall; erratic rainfall; harsh rainfall and hail; early frost, snow, and extreme cold conditions; hot, dry summers; high-wind conditions; nutrient deficiencies; weed and insect pests; and attack by birds (Assefa and Staggenborg, 2010).

Currently, sorghum hybrids are classified by maturity into three groups – early, medium, or late-maturing (Smith, 1995). The growing season ranges from as few as 60 days for early-maturing hybrids, to as many as 150 days for late/full-season hybrids. Sorghum is produced commercially as an annual cereal grain adapted to hot, semiarid tropical and dry temperate areas of the world (NRC, 1996). Based on temperature, three kinds of sorghum -- tropical lowland, temperate, and cool-tolerant tropical high altitude -- are grown in the world (Doggett, 1988). Temperate sorghums are grown throughout the midwestern United States.

Since the 1920s, researchers and plant breeders in the United States have worked to improve sorghum by developing hybrids. In the 1950s, sorghum hybrids were produced to be shorter, higher yielding, and temperately adapted. The development and use of sorghum hybrids more than tripled in the United States from 1950-1954 to 1975-1979 (Jackson et al., 1980). Sorghum can tolerate some salinity and poorly drained soils with pH from 5.5 to 8.5.

Various pests attack sorghum. Major insect pests of sorghum in the United States include: Banks grass mite, *Oligonychus pratensis* (Banks); chinch bug, *Blissus leucopterus leucopterus* (Say); corn earworm, *Helicoverpa zea* (Boddie); fall armyworm, *Spodoptera frugiperda* (J.E. Smith); greenbug, *Schizaphis graminum* (Rondani); sugarcane aphid, *Melanaphis sacchari* (Zehntner), sorghum midge, *Stenodiplosis sorghicola* (Coquillett); sorghum webworm, *Nola sorghiella* Riley; stink bugs, family Pentatomidae; white grubs, *Phyllophaga crinita* (Burmeister); and wireworms, families Elateridae and Tenebrionidae (Teetes et al., 1983; Teetes and Pendleton, 2000). Of the

major sorghum insect pests, greenbug and sorghum midge are considered the key pests of sorghum. In 2013, the sugarcane aphid invaded sorghum in 38 counties and four states of the United States (Bowling et al., 2016). The sugarcane aphid reproduces rapidly and damages sorghum by removing sap and covering plants with honeydew, causing general plant decline and yield loss ranging from 10 to 50%.

Sorghum Water Requirements

Challenges with the declining recharge of the Ogallala Aquifer in the Southern High Plains, coupled with rising energy costs to pump irrigation from the aquifer, has strained agricultural producers and resulted in the need for high-yielding crops that use less water. Grain sorghum is adapted to the Southern and Central Great Plains, but water stress at the critical reproductive stage can sharply reduce grain yields of the crop under dryland conditions (Unger, 1988). In contrast, forage sorghum does not require such timely rainfall to attain good yields and has no critical stage. Sorghum requires 1/3 to 1/2 less water than maize and is better adapted to most regions of Texas (Butler and Bean, 2006). Forage sorghum hybrids produce silage yield similar to that of maize, while using approximately 1/3 less water than is required by maize (Pederson and Rooney, 2004). Stone and Schlegel (2006) reported that grain sorghum began to produce grain at a threshold of approximately 175 mm and generally yielded approximately 230.3 kg m³ above threshold (25.4 mm) while the minimum threshold for maize increased to 279.4 mm; however, maize can produce more kilograms per cubic meter (329.2) than sorghum above the threshold. Spiegel (2015) was in agreement with the thresholds but reported that grain sorghum produced more grain per 25.4 mm of moisture to about 609.6 mm.

Staggenborg (2015) agreed with the thresholds but found grain sorghum more profitable when maize and sorghum prices were equal until maize yields exceeded 8,865 kg ha⁻¹. Less than 533.4 mm of water, grain sorghum was expected to out-yield maize by a slim margin, with the margin increasing as available water for production decreased (Stone, 2013; Staggenborg 2013, 2015). Sorghum requires less moisture for growth than other cereal crops; however, it is very dependent on environmental conditions: studies at Pradesh India, a semi-arid tropical environment, revealed that sorghum required 3.01 kg m³ of water; maize required 2.72 kg m³; barley 2.30 kg m³; and wheat 1.96 kg m³ (House, 1985). The water requirement of sorghum increases as the plant grows, reaching a peak during flowering (using 6-7 mm ha⁻¹ of water a day); after this time, the moisture consumption decreases (House, 1985).

Studies from Colby, KS, reported about 600-650 mm of water to complete the maize-growing season (Lamm et al., 2009). Other studies reported about 450-650 mm of water, depending on the maturity and environment, for maximum sorghum production (Assefa et al., 2010; Lemaire and Hebert, 1996). On the High Plains, evapotranspiration rates of sorghum and maize ranged from 535-628 to 667-789 mm, respectively (Tolk and Howell, 2008; Musick and Duesk, 1980). Howell et al. (1994) studied water use by three crops and reported an average of 578 and 771 mm evapotranspiration for sorghum and maize, respectively, which suggested relatively greater water use by maize than sorghum for maximum production (Assefa et al., 2013). A yield and water relationship curve by Stone and Schlegel (2006) in Tribune, KS, showed that the maximum yield of dryland sorghum (~8 Mg ha⁻¹) can be obtained from 300 mm of soil water at the beginning of

growing season, with an additional 300 mm of water (precipitation) from June to September.

In general, forage sorghum is more water-use efficient than maize and requires less water to produce dry matter (Martin et al., 1976). Typically, maize requires 630 to 762 mm of irrigation in addition to annual precipitation to obtain silage yields of approximately 23 Mg ha⁻¹ (Marsalis et al., 2010). In a two-year study in New Mexico comparing maize, conventional forage sorghum, and brown midrib forage sorghum, Marsalis et al. (2010) found that by reducing the amount of water by 30% (average water applied was 445 mm), average yields for maize and conventional forage sorghum were identical at 24.4 Mg ha⁻¹, while the 21.1 Mg ha⁻¹ yield of brown midrib forage sorghum was different.

Traditionally, grain sorghum has been shown to be superior to forage sorghum for silage. In experiments in Kansas, only high grain-producing forage sorghum hybrids approached the feeding value of grain sorghum hybrids when they were fed as silage (White et. al., 1991). Forage sorghum is considered to be a practical alternative to maize silage when water becomes limited in irrigation systems and when input costs associated with seed and fertilizer are obstacles (Marsalis, 2011). In semi-arid environments, advantages of sorghum compared to maize include: less production cost and greater drought tolerance (Lamm et al., 2007; Marsalis, 2011), slower wilting of leaves and stalks, lower transpiration ratios, and greater ability to recover from drought (Martin, 1930; Sanchez-Diaz and Kramer, 1971). Lamm et al. (2007) compared irrigation rates for major irrigated crops, maize, grain sorghum, soybean (*Glycine max* L.), and

sunflower (*Helianthus annuus* L.), in Northwest Kansas and found less irrigation required and a smaller percentage of decrease in yield of grain sorghum and sunflower over maize and soybean when watered with 25 mm every six days.

Evapotranspiration (ET) is a combination of evaporation from surfaces and transpiration from plants. To date, few data exist comparing the ET of maize and forage sorghum on the Southern High Plains. Howell et al. (2008) showed that forage sorghum had 27% less ET than maize in a study at Bushland, TX, comparing maize and forage sorghum for silage. In circumstances where maize and forage sorghum water-use efficiencies (kg m^{-3}) were similar, maize tended to use more water because of earlier planting and longer growing season (Howell et al., 1997). A study at the USDA-ARS Laboratory at Bushland, TX, compared the ET rates, leaf area index, and dry matter of wheat, maize, and sorghum. The sorghum hybrid was of medium maturity and grown under both irrigation and dryland conditions in 1988 and produced dry matter exceeding 1.4 kg m^{-3} with seasonal ET rates averaging 549 mm (Howell et al., 1996). In 1993, the sorghum hybrid was of longer maturity and planted earlier, which increased the maximum dry matter to more than 2.0 kg m^{-3} with seasonal average ET90 (90% of the ET for sorghum), respectively (Howell et al., 1997). Variable ET rates for maize were reported from 70 to 790 mm by Musick and Dusek (1980), for surface irrigation from 783 to 1,003 mm by Eck (1984), for sprinkler irrigation 883 mm by Howell et al. (1989), and for LEPA-irrigated maize from 786 to 973 mm by Howell et al. (1995).

Sorghum requires less water for cultivation compared to other crops grown in the hot, dry climate of the Texas Panhandle (Smith and Frederiksen, 2000). In general

sorghum requires 25% less water than maize (Martin et al., 1976). Maize has been used predominantly for dairy and beef production in the region; however, it requires large amounts of water (as much as 770 mm yr⁻¹, Al-Kaisi and Yin, 2003; New and Dusek, 2005; Gowda et al., 2007; Howell et al., 2008) to produce high yields and adequate nutrition for the dairy industry (Marsalis et al., 2010). Although sorghum like most crops responds to irrigation, sorghum has a significant advantage over maize as forage because of the lesser water requirement for plant growth. The Texas High Plains is a semiarid region with a high evaporation rate and limited and erratic precipitation (Stewart and Burnett, 1987). Higher yielding hybrids of sorghum because of drought tolerance and better water-use efficiency are a viable alternative to maize. Increasing the quality and yield of sorghum because of its drought tolerance and water-use efficiency has become important in semiarid regions (Marsalis, 2011). New improved breeding methods and higher yielding hybrids have improved the quality of sorghum. Bean and McCollum (2006) determined that some sorghum hybrids had comparable or better quality than maize with equivalent amounts of water.

Economic Outlook of Sorghum

In 2015, production of grain sorghum in the United States totaled 15.2 million Mg, an increase of 9.7 million kg or 64% and a 61% increase in value since 2011 (NASS, 2015). In 2011, less production was caused primarily by extreme drought in the primary sorghum-growing states of Kansas, Oklahoma, and Texas. In 2011, the value of the grain sorghum crop was \$1.26 billion (NASS, 2012) while in 2015, the value was \$2.1 billion (NASS, 2015). Predominantly produced on the southern Great Plains, sorghum is grown

in more than 30 states. Historically, most sorghum has been produced in Kansas and Texas. In 2011, the two states retained their ranking as leading producers, harvesting 78% of the sorghum crop in the United States. Kansas produced 2.79 million tons valued at \$671 million, while Texas produced 1.42 million tons valued at \$331 million. In 2015, Kansas produced 7.2 million Mg while Texas produced 3.8 million Mg. Other states producing large quantities of grain sorghum, more than 40,000 ha harvested in 2015, were Arkansas, Colorado, Missouri, Nebraska, Oklahoma, and South Dakota.

In 2011, lead producers of sorghum in the world included: United States (10.0%), Nigeria (12.6%), India (11.2%), and Mexico (11.2%) (FAS, 2012). In 2015, lead producers in the world included: United States (22.4%), Mexico (10.5%), Nigeria (9.1%), and Sudan and India (each 8.1%) (FAS, 2015).

The United States is the leading exporter of sorghum. In 2011, the United States exported grain sorghum valued at \$948.6 million, a 32% increase from 2010 (FAS, 2012). In 2010, the United States exported grain sorghum valued at \$720.8 million, a 13% increase from 2009. Countries that purchased most of the grain sorghum from the United States were Mexico, Spain, and Japan. Approximately half of the sorghum produced is fed to livestock, and half is consumed by humans and used for many other purposes. In 2014-2015 (August-September), China (97%) was the leading importer of U.S. sorghum, followed by Japan (2%) (U.S. Grains, 2015). In 2015-2016 (August-September), China (83%) continued to lead, followed by Mexico (7%), South Africa (1%), and China (0.8%) (U.S. Grains, 2016).

Sorghum Classification

The ancestors of modern sorghums originated on the continent of Africa, associated with specific eco-geographical regions of sub-Saharan Africa, and consisted of five cultivated races of *S. bicolor* spp., including bicolor, caudatum, durra, guinea, and kafir. Murty and Govil (1967) proposed a system of working groups to classify cultivated races of sorghum. Harlan and DeWet (1972) proposed a simplified system of classification of cultivated sorghum in which five races: 1 = Bicolor (B), 2 = Guinea (G), 3 = Caudatum (C), 4 = Kafir (K), and 5 = Durra (D), and intermediates between the races, were described using 6-15 to include combinations of the basic races. The two approaches were revised to develop a modified numeric classification system by Dahlberg (2000), in which each working group was designated by a two- or three-digit number, the first digit indicating the race (Bicolor-1, Guinea-2, Caudatum-3, Kafir-4, and Durra-5). Various combinations of the races have numbers from 6 to 18, and 19, 20, and 21 are unclassified types. The last digit of a working group number indicated the subtype of the working group. A last digit '0' indicated the working group closest to the essential characteristics of the race, i.e., the actual sorghum Bicolor classification where the working group is most closely related to the actual is 10. Zera zera is a working group designated by the number 37, being a part of the race Caudatum, while understanding there are at least seven known subtypes. Understanding the genetics behind working groups and races would be useful for sorghum researchers by increasing diversity. Working groups such as Zera zeras contain useful sources of tan plant and white grain for use in food systems, while kafirs yield well.

Introduction of Sorghum to U.S.

Many sorghum varieties were introduced to the United States beginning in the 1850s. The kafirs came from South Africa in 1876, and shallu from India in 1890. In 1857, Peter Wray brought 16 cultivars of sorgho from Natal, South Africa (Snowden, 1936). J. H. Martin (1936) suggested that the first sorgho introduction was Chinese Amber introduced in 1853. Some suggest that Benjamin Franklin grew broomcorn, a sorghum relative, in the late 1700's. Most of the introductions were from the kafir and durra races (milos). Many sorghum varieties, including Redlan and Martin, derived from the kafir and milo races, were developed during the early decades of the 20th Century. Martin variety was grown on 80% of the sorghum acreage in the U.S. from the early 1940s to 1955 (Duncan et al., 1991).

History of Hybrid Production

Until the early 1950s, hybrids were made by hand emasculation, or mechanical-sterilization methods such as hot water emasculation. The experimental hybrids documented the heterotic potential of sorghum but they also confirmed that economically viable production of hybrid seed was needed (Karper and Quinby, 1937). In the 1950s, a commercially feasible system for large-scale hybridization in sorghum was developed (Quinby and Martin, 1954; Stephens and Holland, 1954). In 1954, Stephens and Holland proposed a method for creating hybrids based on a cytoplasmic male-sterility system that used sterile cytoplasm from milo. Backcrossing kafir with milo, with kafir as the recurrent parent, would result in kafir nuclear genes in milo male-sterile cytoplasm, in effect making a male-sterile version of the kafir line. The male-sterile kafir line could be

crossed with a durra male or any of a large number of milo/kafir derivative lines, restoring its fertility. Within a few years, most of the sorghum production area in the United States was planted with kafir female x milo/kafir derivative male hybrids (Duncan et al., 1991). The system was used to such a wide extent that much of the hybrid sorghum in the 1960s in the United States had similar cytoplasm, as still is the case. In making hybrids, A-lines (male sterile) are used as the female parents. These A- or male-sterile lines are derived from B-lines, and each A-line is isogenic to its corresponding B-line, from which it was derived, i.e., it is genetically identical to the B-line at all loci but the locus/loci for male sterility. An A-line is different from a B-line only because it is male sterile. This system from a practical standpoint of hybrid development works effectively and enables a sorghum breeder to classify a newly introduced genotype for use in a breeding program. However, research at Texas A&M University is characterizing elite U.S. sorghum lines, and results to date suggest a genetic grouping of lines into five broad groups: Kafir-Milo derivative males, Kafir type females, Zera zera derivative males, Zera zera derivative females, and Feterita derivative males (Menz et al., 2004).

Sorghum Conversion Program

Soon after hybrid sorghums were developed, sorghum breeders realized that the genetic base in the United States was limited, in large part because of difficulty in using tropical, tall, photoperiod-sensitive sorghums in the temperate United States. This concern led to the development of the TAES-USDA Sorghum Conversion Program, initiated in 1963. The purpose of the Sorghum Conversion Program was to convert

exotic tropical photoperiod-sensitive sorghum lines into temperate-adapted photoperiod-insensitive lines suitable for breeding programs in the United States, which enabled diversification of available germplasm (Stephens et al., 1967). The Sorghum Conversion Program has had a dramatic impact on sorghum improvement; it is difficult to find hybrids grown today that do not have sorghum conversion germplasm in their pedigrees.

Another reason for the narrow genetic base was that most hybrid sorghum production was (and is still) based on the same cytoplasm system (known as A1 sterile cytoplasm). Different male-sterility inducing systems, such as A2 and A3 cytoplasm, have been discovered in the last few decades, and hold promise for widening the genetic variability of elite lines. The A2 cytoplasm was reported from IS12662C (Schertz, 1977; Schertz and Ritchey, 1978), belonging to the caudatum-nigricans group. Quinby (1980) reported the sterility-inducing cytoplasm from the line IS1112C and designated it A3 cytoplasm whose limited sources of fertility-restorer genes have precluded widespread utilization (Rooney, 2000). Other cytoplasmic sterile systems also have been reported (Schertz and Pring, 1982). Apart from different cytoplasmic sterility sources, the conversion program has made available agronomically desirable lines with resistance to such economically significant diseases as anthracnose (*Collectotrichum graminicola* (Ces.) G.W. Wils) and downy mildew (*Sclerospora sorghi* (Kulk) Weston & Uppal). Sources of resistance to insect pests such as greenbug and sorghum midge, and to pre- and post-flowering stress, have been found in converted materials (Rosenow and Dahlberg, 2000).

The USDA-TAES Sorghum Conversion Program made available diverse sorghum germplasm, and different male sterility-inducing systems have been introduced. The Sorghum Conversion Program continues to serve as a major source of new germplasm for many breeding programs throughout the world (Smith and Frederiksen, 2000). It is a crucial component for increasing diversity in sorghum breeding programs.

Combining Ability

Combining ability is useful for plant breeders to better understand genetic variance and inbred lines important in identifying hybrids for commercial production. Research on combining ability helps plant breeders to select the best parents for development of hybrids or varieties. The concept of general- and specific-combining ability was conceived by Sprague and Tatum (1942) who designated general-combining ability (GCA) as the average performance of a line in hybrid combination, and the term specific-combining ability (SCA) was applied to cases where certain hybrid combinations did relatively better or worse than would be expected on the basis of the average performance of the lines. GCA measures the average performance of an inbred when crossed with a series of other inbreds. GCA indicates the worth of an inbred as a parent of multiple hybrids. Estimates of GCA are useful for choosing a few key inbreds to use as testers. SCA is because of genetic effects specific to a hybrid combination and not accounted for by GCA effects. SCA measures genetic effects that are specific to a hybrid combination. As a general rule, GCA is the result of additive gene effects, while SCA is the result of non-allelic interactions (Jinks, 1954), is assumed to be a deviation from

additivity (Bernardo, 2014), or is attributed primarily to deviations from the additive gene action caused by dominance and epistasis.

Crossing a plant line with several others provides the mean performance of the line in all its crosses. Mean performance, when expressed as a deviation from the mean of all crosses, is the GCA of the line. Any particular cross, then, has an expected value which is the sum of the GCA of its two parental lines. The cross might, however, deviate by a greater or lesser extent from the expected value. The deviation is the SCA of the two lines in combination. In statistical terms, the GCA is the main effect while the SCA is an interaction (Bernardo, 2014). The SCA will be of interest because it will indicate the degree of heterosis expressed in each cross while representing the dominance deviation value in the simplest case but ignoring epistatic deviation. Therefore, a cross between sorghums with greater combining ability, if from genetically divergent backgrounds, is more likely to result in a hybrid with a greater degree of heterosis, which will also be manifested in a greater SCA for one of the lines in specific combination with the other.

Green (1948b) studied F_2 generations of maize derived from crosses of high x high, high x low, and low x low combining inbred lines and found that combining ability was an inherited character. Comstock and Robinson (1948) and Kempthorne (1957) introduced a method to study combining ability of inbred lines as the line x tester method. In this design, a set of female parents (n) are crossed with a genetically different set of male parents (m) in all possible combinations, resulting in a total of nm progenies. The advantage of this method allows the breeder to test at one time a larger number of inbred

lines. The focus of the research will be narrowed to include only the line-by-tester method.

Griffing (1956) showed that for homozygous parents (inbreeding coefficient $F = 1$), the genetic variance σ^2G (variance among hybrids) could be expressed in terms of combining ability variance as:

$$\sigma^2G = \sigma^2GCA + \sigma^2SCA$$

where σ^2GCA and σ^2SCA are the variances for general- and specific-combining ability effects, respectively. Components of combining ability variance might reflect additive effects and additive interactions, while SCA variance components might reflect dominance and epistasis, and components of additive epistasis (Rojas and Sprague, 1952).

Kambal and Webster (1965) estimated the components of variance caused by GCA and SCA and their interaction with years for five traits in split-plot design and reported that both GCA and SCA were important in determining yield and other characters, but the GCA effects were more important and more stable over years. Beil and Atkins et al. (1967) observed that variances for GCA were three times more than specific effects and found similar ratios with such traits as the number of kernels per panicle, number of panicles per plant, and weight of 100 kernels. They reported that SCA effects were more stable than GCA effects in various environments particularly for the grain yield and number of kernels per panicle.

The concept of a good plant tester has been another question for most of the breeding programs to date. Matzinger (1953) defined a desirable tester as one that combined the greatest simplicity in use with maximum information on the performance to be expected from the tested lines. However, Allison and Curnow (1966) assessed that the best tester was the one that maximized the expected mean yield of the variety produced by random mating the selected genotypes. Green (1948a) compared maize progenies from crosses using two testers and found that the average performance was a better estimate of combining ability than was the top cross performance of either tester alone. Top cross refers to the estimation of combining ability of testers based on matings with specific single lines or a cross between an inbred line and an open-pollinated variety. For heterogeneous populations, Cress (1966) concluded that testers could be selected based on average performance of a test cross, i.e., the tester with the greatest average cross performance was chosen. However, if the selected genotypes are not to be used immediately in hybrid combination with the tester, emphasis on heterotic response is misplaced because it reveals little concerning the genetic potential and nothing concerning the expected rate of progress from selection.

Giriraj and Goud (1982) found that both additive and non-additive gene actions were important for grain yield, panicle length, number of primary branches, length of the primary branch, 100-kernel weight and number of grains per panicle, and number of leaves in grain sorghum. Nayeem and Bapat (1984) reported that the estimates of the mean squares due to GCA were more than SCA for the traits studied, indicating the importance of additive gene action for those traits. Kishan and Borikar et al. (1989)

analyzed a line x tester involving diverse cytoplasmic lines in sorghum and reported that restorers IS 12567C, IS 12662C, and SPV 650 had desirable GCA effects, and the cross (A [SUB2]T x 398) x IS 12662C had the greatest mean yield coupled with good SCA effects. Patel et al. (1991) studied the combining ability of 36 genotypes for seven traits in different environments, indicated the importance of additive and non-additive gene effects in the inheritance of the traits, and concluded that most genetic variation was additive. Reddy and Joshi (1993) studied combining ability for grain yield and its five components from F₁ to F₄ generations. The best parent identified was CSV 10 which was a good combiner for grain yield and 1000-kernel weight. They also reported that the magnitude of SCA variance decreased in F₂ and later generations and identified the best cross combinations as SPV 451 x SPV 474, SPV 474 x IS 508, CSV 10 x SPV 451, and CSV 10 x SPV 474 that can be exploited to improve yield of sorghum grain. Badhe and Patil (1997) found that additive gene action for plant height and non-additive gene action were dominant for grain yield and other attributes. They identified female MS 2077A and male SPV 386 as the best combiners for almost all traits except plant height and 1000-kernel weight. The cross MS 2077A X SPV-245 involving high x low combiners showed significant positive SCA effect for all the panicle traits, suggesting a dominant role of non-additive gene action for panicle traits. Can et al. (1997) studied combining ability in a diallel mating system in early maturing grain sorghum by sowing in spring and summer, observed that GCA and SCA effects were significant for all the traits, and identified some parents having large positive GCA for grain yield and small or negative for culm length and days to panicle emergence.

Biradar et al. (2000) revealed significant variances due to GCA and SCA, suggesting the importance of additive and non-additive gene effects in inheritance of grain yield and other component traits. They identified that females 104A and P2A were good general combiners for grain yield per plant that had positive and significant GCA effects, and they also showed good average performance indicating strong relationship between GCA effects and *per se* performance (the individual performance of the line or tester). Hovny (2000) reported that ICSR-112 was a good combiner for grain yield per plant, and the cross ICSA-1 x ICSR-112 gave large grain yield per plant because it had high SCA. Hovny et al. (2001) reported that the female line ICSA-40 and the restorer ICSR-138 had significant positive GCA for grain yield, and the crosses ICSA-1 x ICSR-93002 and ICSA-40 x ICSR-89037 yielded more grain than the check. Iyanar et al. (2001) studied combining ability for grain yield and its components in sorghum and observed that non-additive gene action was dominant for all characters. The lines 2077A and 88005A expressed superior *per se* performance and GCA effects on grain yield, while testers such as CO 26, SPV 1192, and SPV 881 showed high *per se* performance for days to 50% flowering, panicle length, panicle weight, and grain yield. Kanawade et al. (2001) reported that additive gene effects were important in the inheritance of panicle breadth, 1000-kernel weight, and grain yield per plant, while plant height, days to 50% flowering, days to maturity, number of leaves per plant, and panicle length were under the control of non-additive gene effects. Siddiqui and Baig (2001) reported the ratio of GCA to SCA variances was less than unity for all the characters except days to flowering which indicated dominance of non-additive gene action for the characters.

El-Mottaleb and Asran (2004) analyzed a line x tester and indicated that lines ISCA-88003, ICSR-237, and ICSR-92003 had the most significant GCA effects for grain yield and that SCA variance was more important for all the traits studied except plant height. Kenga et al. (2004) studied combining ability in tropical sorghum and reported significant GCA effects of males for all the traits and detected significant SCA in all traits except inflorescence length. From the ratio of GCA to SCA variances, they concluded that non-additive gene action was dominant for most of the traits. El-Menshawi (2005) studied combining ability in eight environments and observed that non-additive effects were two times greater than additive effects. Kenga et al. (2005) observed that the ratio for GCA to SCA variances ranged from unity to a high of 10, indicating dominance of additive gene effects for most yield-contributing traits in sorghum. Chaudhary et al. (2006) analyzed a line x tester for combining ability and observed non-additive gene action for all the traits and identified the lines 116A and 117A and testers RSLG 112, SPV 1090, SPV 839, and SPV 1167 as good general combiners for yield and its contributing characters. Both GCA and SCA effects were present among the crosses and involved at least one of the parents with large GCA effect. They also exhibited significant SCA effects for most of the traits. Kulakarni et al. (2006) did line x tester analysis using 33 hybrids along with parents and indicated the importance of additive gene effects in the inheritance of days to 50% flowering and fodder yield per plant, while non-additive gene action was important for 1000-kernel weight, plant height, number of leaves per plant, days to maturity, panicle weight, number of primary branches per panicle, and grain yield per plant. Premalatha et al. (2006) studied heterosis and combining ability for grain yield and its components and

revealed non-additive gene action for all the traits and also reported that parents that performed well for *per se* performance and GCA effects could be considered good parents. Salini et al. (2008) did line x tester analysis in dual-purpose sorghum and reported that grain yield and crude protein content of leaves were affected by additive gene action, while green forage yield at 50% flowering, 1000-kernel weight, protein content of grain, and leaf breadth were under the control of additive and non-additive gene action, and the other traits were under the control of non-additive gene action. They also found that the lines SPV 1782, SPV 1714, SPV 1754, and SPV 1616 and tester CSV 15 were good general combiners. SPV 1782 x HC 308, SPV 1730 x HC 308, and SPV 1616 x CSH 16 were good specific combiners. Tadesse et al. (2008) reported that GCA for plant height, panicle exertion, panicle length, grain yield, and kernel weight was significant among male parents, indicating the prevalence of additive gene action in determining the traits while the male x female interaction and SCA effect for all the parameters considered was non-significant, indicating little importance of non-additive genetic effects in expression of the traits.

Degu et al. (2009) did line x tester analysis for yield-related traits in grain sorghum in three low-moisture areas in Ethiopia and observed dominance of additive components in inheritance of most of the developmental, panicle, and grain traits. Aruna et al. (2010) reported that additive and non-additive gene actions were equally important for controlling such traits as the number of kernels per primary branch, number of primary branches, number of secondary branches, and kernels per unit length of primary

branch, while non-additive gene action was particularly important for panicle weight, yield per plant, and number of leaves.

Indhubala et al. (2010) reported the preponderance of non-additive gene action for all the characters studied in sweet sorghum, and based on GCA effects, the lines BJ 3A, CK 60A, and AKMS 22A and the testers RSSV 9, SSV 84, and ASV 9401 were identified as good combiners for most of the characters. The hybrid combinations AKMS 22A x RSSV 9, BJ 3A x VMS 98001, BJ 3A x RSSV 9, AKMS14A x RSSV 9, and CK 60 A x VMS 98001 were the best specific combiners. Makanda et al. (2010) studied combining ability for sorghum grain yield in different tropical low and mid-altitude environments and reported that GCA and SCA effects were significant for all the traits, implying both additive and non-additive gene actions were important in controlling inheritance of the traits. Kanbar et al. (2011) observed that varieties Baladi-1, Baladi-2, and Ezraa-7 had significant positive GCA for grain yield. Mahdy et al. (2011) reported that GCA variance components for days to flowering, plant height, and 1000-kernel weight were larger than those of SCA in different environments, while SCA variance for grain yield was larger than that of GCA. Variance components of GCA and SCA varied greatly from location to location and early to late planting for days to flowering, plant height, and 1000-kernel weight.

Tariq et al. (2014) studied heterosis and combining ability for quantitative traits in sorghum by evaluating nine crosses and six parents in Pakistan during 2009-2010. They found the mean squares for genotype, GCA, and SCA were significant for all the traits studied; however, the phenotypic component of variance was greater than the genotypic

component of variance for all the traits. Similarly, the estimation of GCA variance was less than that of SCA variance for all traits. The studies revealed significant mean squares for GCA and SCA which led to the existence of both additive and non-additive types of gene action. Numerous studies also reported significant variability for genotype, GCA, and SCA for different components in sorghum (Mohammed et al., 2008; Kamdi et al., 2009; Prakash et al., 2010). Tariq et al. (2014) discovered that the dominance variance was greater than the additive variance for all the parameters evaluated, with a degree of dominance greater than unity. The ratio of GCA to SCA is an indicator of the dominance of additive gene effects. Mohammed et al. (2015) researched combining insect resistance with desirable agronomic and morphological traits to increase sorghum productivity and found a larger ratio of $\sigma^2(\text{GCA})/\sigma^2(\text{SCA})$ for 100-kernel weight in the post-rainy season, indicating the dominance of additive gene action, whereas both additive and non-additive gene actions were observed during the rainy season. Grain yield had greater SCA variance, suggesting dominance of dominance (non-additive) type of gene action (Wilson et al, 1978; Singhanian, 1980; Hovny et al., 2000; Girma et al., 2010).

Hallauer and Miranda (1981) reported that some form of early generation testing is included in most breeding programs. However, this does not imply that perfect relationship exists between initial and later generations for inbreeding, because early testing was designed to separate the population of lines into groups of good and poor combining ability. According to Sprague (1946), early testing was based on two assumptions: (i) there are marked differences in combining ability among open-

pollinated plants, and (ii) selected samples based on tests of combining ability of S0 (variety) or S1 (advanced generations of a hybrid) plants offer a larger proportion of superior lines upon inbreeding, and then selection does a more nearly random sampling of combining abilities from the same population than on the basis of visual selection alone.

Kambal and Webster (1965) studied data collected during two years from a set of 190 sorghum hybrids obtained by crossing 10 male-sterile lines and 19 restorers. They concluded that both $\sigma^2\text{GCA}$ and $\sigma^2\text{SCA}$ were important and stable over years for grain yield. Beil and Atkins (1967) studied the performance of 40 F_1 hybrids obtained by crossing five male-sterile lines with eight restorers at three locations during two years and found that GCA variances for grain yield, number of kernels per panicle, number of panicles per plant, and 100-kernel weight were much larger than SCA variances. Malm (1968) studied eight fertility restorer lines developed from African introductions crossed with four male-sterile lines to produce 32 hybrids. The data reported indicated that parents with large kernels produced hybrids that yielded most; however, all sets of exotic hybrids produced larger kernels than the checks while some exotic hybrids produced 50% more protein than the checks. Malm (1968) concluded that for grain yield, kernel size, and protein content, additive gene action was more important than non-additive gene action. Mattei (1974) working in Venezuela with a line x tester cross of eight male-sterile lines and four restorers evaluated at three locations observed that variance caused by additive effects was several times greater than variance caused by non-additive effects for grain yield and concluded that evaluating parents based on GCA should be effective.

Shankaregouda et al. (1972) used a line x tester study to show that plant height and days to flowering were mostly controlled by additive gene effects, whereas, yield and number of kernels per panicle were mostly controlled by non-additive gene effects, which was supported by results of Goud et al. (1973), Shahane and Bapat (1981), Shinde and Sudewad (1981), and Rao et al. (1982).

Heterosis

Plant breeders rely on genetic variability to select plants with traits of interest in a population. Plant breeders for almost two centuries observed expression of hybrid vigor, but it was not until the early part of the 20th Century that the modern concept of hybrid vigor was described. Heterosis is defined as the difference between a hybrid and the mean of the two parents for numerous agronomic traits that can be evaluated (Falconer and Mackay, 1996; Bernardo, 2014). Heterosis is expressed as the percentage increase or decrease of an F₁ hybrid over the mid-parental value. Exploitation of heterosis began in the United States in the 1950s, resulting in large increases in yields of sorghum and maize (USDA National Agricultural Statistics Service, 2007; Troyer and Wellin, 2009). The more genetically divergent a hybrid is, based on genetic relatedness of the parental lines, the greater the degree of heterosis that might exist.

The characteristically superior performance of hybrid sorghums was because of a phenomenon known as “heterosis” or “hybrid vigor”, in which hybrids demonstrated markedly vigorous growth and yield when compared with their parents (Bernardo, 2014). In 1914, Shull proposed the term "heterosis" to describe developmental stimulation resulting from union of different gametes that caused superiority of hybrids over their

parents (Shull, 1952). The terms heterosis and hybrid vigor are synonymous and often used interchangeably. A large degree of heterosis occurs when the parents are genetically divergent or unrelated, resulting in a heterozygous hybrid. Therefore, development of superior high-yielding sorghum hybrids requires a system by which genotypes can be crossed on the basis of the degree of ‘unrelatedness’ between them. Beil and Atkins et al. (1967) observed greatest heterosis for grain yield, suggesting that non-additive effects might be proportionately greater for grain yield than for any of its individual components. Quinby (1974) proposed a complementary interaction between recessive and dominant alleles as a possible cause of heterosis.

Blum et al. (1977) defined heterosis as “the advantage of the hybrid over the best parent.” “Midparent heterosis”, which is used in quantitative genetics, is defined as the superiority of a hybrid over the mean of its parents (Bernardo, 2014). The reason for the phenomenon is not understood, but two principal explanations are the concepts of dominance and overdominance (Crow, 1948, 1952). Davenport (1908) proposed the dominance theory, supported by Bruce (1910), Jones (1917), and Collins (1921) that cites the effect of dominant favorable alleles masking unfavorable recessive alleles as the reason for the superiority of a hybrid (Bernardo, 2014). East (1908) and Shull (1908) independently proposed the overdominance theory, which suggests that the heterozygous condition is responsible for heterosis; it is the inherent superiority of a heterozygote over either homozygote (Bernardo, 2014). Franca et al. (1986) reported midparent heterosis for grain yield per plant, with negative heterosis observed for days to 50% flowering and 1000-kernel weight. Berenji (1988) found that heterosis over the mid-parental value was

greater for number of kernels per panicle in grain sorghum, and the hybrids were taller and yielded more than the parents. Nimbalkar et al. (1988) reported positive and significant relative heterosis for panicle length, 1000-kernel weight, and grain yield per plant. Patel et al. (1990) observed negative heterosis for plant height and significant positive heterosis for panicle length, 1000-kernel weight, and grain yield per plant. Nandanwankar (1990) reported that the number of grains per panicle was the major contributor to heterosis for grain yield in sorghum and among the remaining components, the number of primary branches per panicle exhibited maximum heterosis followed by the number of whorls per panicle, panicle length, and 1000-kernel weight. Rao et al. (1993) reported that hybrids had a large degree of heterosis over their parents in days to 50% flowering, panicle length, and grain yield per plant, and limited heterosis for 1000-kernel weight. Ganesh et al. (1996) noticed midparent heterosis for days to 50% flowering and grain yield per plant, better parent heterosis for plant height and panicle length, and all three types of heterosis for 1000-kernel weight. Madhusudhana and Patil (1996) crossed a random sample of 97 F_3 sorghum segregates from the cross 3660B x MR-75 with a sterile tester and observed that as many as 21 derived F_1 plants showed heterosis for yield, and some derived F_1 plants were superior over a commercial check. Can et al. (1997) reported great positive heterosis in grain yield for more than half of the hybrids, and several cross combinations had high heterosis for grain yield but negative for days to panicle emergence. Lokapur (1997) noticed heterosis over the better parent in the positive direction for plant height, significant positive heterosis for 1000-kernel weight, and negative heterosis for days to 50% flowering. Salunke and Deore (1998) reported that low heterosis was observed for days to 50% flowering and moderate

heterosis for 1000-kernel weight, while for plant height, and significant positive heterosis over a standard check was observed. Iyanar et al. (2001) reported that the preponderance of non-additive gene action for the traits indicated the scope for exploitation of heterosis in improving yield in sorghum. El-Mottaleb and Asran (2004) reported that better parent heterosis was generally manifested for plant height, panicle length, panicle width, and grain yield per plant; heterosis for 1000-kernel weight was observed for few of the crosses; and greatest positive significant heterosis for grain yield (87.88%) was manifested by the cross ICSA-37 x ICSR-93023. Kenga et al. (2005) observed high positive heterosis for sorghum grain yield and its components including days to anthesis and plant height for most of the hybrids in different environments. Premalatha et al. (2006) found that the hybrid CSV 15 x SPV 1521 yielded most grain with 90.00, 86.89, and 33.45% heterosis over the midparent, better parent, and standard check, respectively. The hybrid also was superior in terms of days to 50% flowering, plant height, number of kernels per panicle, and grain yield per plant, but hybrids that exhibited heterosis for grain yield were not heterotic for all the traits. Sharma and Sharma (2006) reported that crosses SPV 1518 x IS 18580, IS 18580 x Raj 13, and SPV 1514 x Raj 36 had high heterosis over the midparent and better parent for grain yield per plant, panicle weight, and panicle length. Salini et al. (2008) reported that high significant mean performance of hybrids with that of parents suggested the existence of heterosis for all the traits and the importance of non-additive gene effects in determining the traits. Makanda et al. (2010) found that hybrids were dominant for grain yield and displayed as much as 285% standard heterosis, and overall hybrid mean yield was significantly greater than that of parents and standard check varieties, which was attributed to high levels of average

heterosis and standard heterosis, respectively. El-Dardeer et al. (2011) studied heterosis under normal and water-stress conditions and reported the better parent heterosis was generally manifested for plant height, panicle length, panicle width, and grain yield per plant, and crosses *viz.*, ICSA-364 x ICSR- 66, ICSA-364 x ICSR-102, and ICSA-490 x ICSR-66 had significant standard heterosis for grain yield over the check. Kanbar et al. (2011) observed that hybrids Baladi-4 x SPL-10A and Baladi-3 x ATX-629 had large significant positive values of heterosis, and high levels of mid- and better-parent heterosis were recorded for grain yield in all hybrids except Ezraa-3 x SPL10-A and Ezraa-5 x ATX-629. Mahdy et al. (2011) reported positive heterosis for grain yield and 1000-kernel weight and negative heterosis for days to 50% flowering in different environments.

The phenomenon of heterosis between genetically distant or unrelated genotypes has been widely reported, and the idea of defining genetic relatedness of genotypes has spurred research to determine genetic distance between lines, based on the degree of similarity in molecular markers shared. An estimate of genetic distance can be used as an index of relatedness, and therefore as a tool for defining the potential for diversity. This approach has been used in development of numerous crop plants.

In rice, good correlations between molecular marker-based distance and hybrid performance, using diallel analysis, were reported by Saghai-Maroo et al. (1997), and between specific marker heterozygosity (solely considering markers exhibiting significant effects on the traits being studied) and heterosis by Zhang et al. (1994, 1995) who also, however, reported low correlations with general heterozygosity based on all the

markers. Xiao et al. (1995) and Hua et al. (2003), using molecular marker techniques for rice, concluded that dominance was the major basis of heterosis. Charlesworth and Charlesworth (1999) concluded that overdominance effects were unimportant in most cases.

In alfalfa (*Medicago sativa* L.), Riday et al. (2003) observed no correlation of SCA or midparent heterosis with genetic distance. They theorized that estimates of genetic distance based on neutral molecular markers (not linked with genes controlling traits of interest) did not reflect heterotic potential between genotypes.

Bernardo (1992) suggested a set of conditions, including large heritability and strong dominance effects, for effective prediction of hybrid performance based on marker heterozygosity. Several studies based on molecular markers in crops such as maize (Stuber et al., 1992; Cockerham and Zeng, 1996), and rice (Yu et al., 1997; Li et al., 2001; Luo et al., 2001) found overdominance to be important. However, the possibility of pseudodominance effects could not be eliminated, while epistasis was shown to play a considerable role in the phenomenon of heterosis (Carr and Dudash, 2003).

Grain yield is a complex trait controlled by polygenes and has low heritability especially in stressful environments. For example, selection for grain yield under severe drought stress has often been considered inefficient because the estimate of heritability of yield has been observed to decrease with reduced yield (Bolaños and Edmeades, 1993). Quinby (1963) reported heterosis of sorghum in the forms of increased grain and forage yields, hastened flowering and maturity, increased height, and larger stalks and panicles.

Enhanced grain yield was reported by Kambal and Webster (1966) and Blum (1969) to be the product of more kernels per panicle and increased kernel weight.

Heritability is a key component of plant breeding programs, because a trait needs to be heritable if any breeding progress is to be made. Defining and calculating heritability depends on how the plant breeder considered the trait, because it can be measured based on single plants or a group of offspring. Heritability is a measure of the relative influence of genetic versus non-genetic effects on the expression of a trait. It can explain the variation that is transferred from parents to offspring.

If a phenotype is determined, in part, by the genotype, the heritability is known as broad-sense heritability (Bernardo, 2014). Broad-sense heritability is the ratio of genotypic to phenotypic variance and is usually denoted by H . A trait considered heritable, in the sense of it being transmitted from a parent to its offspring, would be defined as having narrow-sense heritability. Narrow-sense heritability is denoted by h^2 and is the ratio of additive to phenotypic variance (Bernardo, 2012, 2014). The variance of the additive effect of alleles rather than genotypic variance is considered. Additive variance is because of the average effects of alleles and replaces genotypic variance in the narrow-sense heritability formula.

Both definitions have meaning and application when considering the germplasm and environment in which the plants are grown. Broad-sense heritability is expressed when a plant breeder is trying to exploit all types of genetic variance. Epistatic, dominant, and additive variance can all be exploited among clones in asexually propagated species (Bernardo, 2014). Narrow-sense heredity is expressed when a

breeder uses selection to determine the change or increase in the trait of interest.

Selection involves identifying the best individuals in a population and using the selected plants as parents to produce the next generation. The goal of the plant breeder is to produce offspring with a larger quantitative or qualitative mean than that of the previous generation.

Govil and Murty (1973) reported high heritability estimates for days to flowering, length of the primary branch, plant height, and number of leaves and also reported that heritability estimates for protein and lysine were 9 and 14%, respectively, which were similar to the heritability estimates of grain yield. They also indicated that selection for grain quality did not seem to be more difficult than for grain yield. Nayeem (1992) said additive gene action was important in controlling such traits as protein, lysine, and sugar contents. Desai and Shukla (1995) reported that additive and non-additive gene effects controlled the inheritance of most of the traits, with the latter being more important in sorghum and grain yield. Panicle components were under the control of dominance gene action, and the exploitation of hybrid vigor seemed to be beneficial. Sankarapandian et al. (1996) found that heritability was greatest (90%) for all the traits except kernel density and panicle breadth. High heritability of 99.87% was observed for the number of grains per panicle, followed by grain yield and 1000-kernel weight. They also observed that the genetic advance as a percentage of the mean was greatest for grain yield, 1000-kernel weight, number of rachis branches, peduncle length, and panicle length.

Hoshmand and Rezai (1997) used generation mean analysis to study the heritability estimates and type of gene action involved for grain yield per plant, mean

panicle weight, panicle length, plant height, and number of days to flowering in 17 families and reported great genetic potentials among the lines studied. Different heritability estimates were observed in different families, and their averages ranged from 50.99% for panicle length to 77.54% for plant height. Can et al. (1998) reported that high heritability estimates coupled with high genetic advance were observed for dry weight of leaves, plant height, and 100-kernel weight, indicating the traits were controlled by additive gene action. Audilakshmi and Aruna (2005) reported that grain size was controlled by dominant genes that were polygenic. Predominance of dominance and epistatic interactions in crosses indicated that selection for larger grain size would be more effective if the dominance and epistatic effects were first reduced by a few generations of selfing. Aruna et al. (2010) studied the use of different germplasm lines in developing sorghum varieties and hybrids and reported that genetic improvement of grain yield per plant would be easier through indirect selection because many yield components showed significant positive correlation with yield along with high heritability value.

Correlation

Correlation studies provide information on the association of yield with its component characters and help formulate phenotypic selection to discard the bulk of the segregating base population and select desirable types phenotypically. Improvement of yield is possible through the associated characters. Beil and Atkins et al. (1967) reported that the correlation of grain yield with its components revealed that the number of kernels per panicle was related to yield. Expressions for yield were not affected appreciably by 100-kernel weight, and significant negative association for the number of panicles per

plant with grain yield was observed for hybrids and parental lines. Badwal (1970) observed no association of grain yield with days to 50% flowering, plant height, span of maturity, or 100-kernel weight in sorghum. Goud and Asawa (1978) studied correlation and found that yield was positively and significantly correlated with plant height and significantly correlated with the number of days to 70% maturity, and yield was more correlated with plant height in the parental population than in F₁ plants. However, panicle length and grain yield were negatively correlated. Bohra et al. (1985) reported that grain yield per plant had significant and positive correlation with harvest index and panicle length in sorghum grown in limited moisture or irrigated conditions and also with flag-leaf area in the irrigated condition. The relationship between grain yield and protein content was negative. Sugar content was negatively correlated with panicle length; however, it did not show any association with protein content. Nimbalkar (1988) observed significant positive correlation coefficients between grain yield per plant and panicle weight, panicle breadth, and 1000-kernel weight. Raut et al. (1992) observed that among 20 sorghum genotypes, the number of leaves per plant and panicle weight were positively and significantly associated with yield. Veerbadhiran et al. (1994) noticed that grain yield was positively correlated with the number of days to 50% flowering and panicle weight. Sankarapandian et al. (1996) found no correlation of grain yield with most of the component characters except 1000-kernel weight and the number of kernels per rachis that might be because of elimination effects of one or other characters contributing for grain yield in *rabi* sorghum. Jeyaprakash et al. (1997) reported grain yield was significantly and positively correlated with panicle weight, panicle length, and dry fodder yield, and plant height had a positive significant association with grain yield.

Taurchi and Rezai (1997) observed that plant height, panicle length, and 100-kernel weight were significantly positively correlated with grain yield. Can et al. (1998) found that harvest index and its components were positively correlated with grain yield and suggested that harvest index could be used as a selection criterion for high yield and short-plant genotypes. Iyanar et al. (2001) analyzed correlations of 54 genotypes of sorghum and reported that grain yield was significantly and positively correlated with panicle weight and length. Sunku et al. (2002) reported that correlations were significant and high among yield of dry matter, content of dry matter, yield of green fodder, plant height, number of leaves, leaf length, and leaf width in grain sorghum. Tiwari et al. (2003) noticed that panicle length was an important trait contributing to yield because it was significantly positively correlated with grain yield. Ezeaku and Mohammed (2006) revealed significant and positive correlation between grain yield and panicle weight, grain yield and 1000-kernel weight, and 1000-kernel weight and panicle weight, and similarly, significant but negative correlation between the number of panicles and panicle length. Plant height had high positive phenotypic and genotypic correlation coefficients with panicle weight and grain yield in sorghum. Makanda et al. (2010) observed that grain yield was positively and significantly correlated with panicle length and the number of leaves per plant, suggesting improvement in grain yield potential as the number of leaves and panicle size increased. Prakash et al. (2010) reported that yield of green fodder per plant was significantly and positively correlated with plant height, number of tillers, leaf length, leaf breadth, stalk diameter, hydrocyanic acid content, and crude fiber. The number of days to 50% flowering, crude protein, and in vitro dry matter digestibility were negatively associated with the yield of green fodder per sorghum plant. Warkad et

al. (2010) found that only one character, 1000-kernel weight, was significantly correlated with grain yield per plant at and among the yield components themselves. The number of days to 50% flowering was significantly positively associated with the number of days to maturity, plant height, dry fodder weight per plant, and number of leaves per plant.

Chavan et al. (2011) reported improvement in component traits including panicle length, number of grains per panicle, panicle width, test weight (a measure of bulk density, or the weight of a specified volume of sorghum), number of primary branches per panicle, and harvest index through simple selection methods in sorghum, and the traits had positive and significant association with grain yield per panicle. Mahajan et al. (2011) stated that grain yield per panicle showed positive significant correlation with panicle length, panicle width, plant height, branches per panicle, grains per panicle, test weight, and harvest index in sorghum.

Harvest Index

Harvest index is defined as the ratio of grain yield to dry matter yield (Donald, 1962) or total biomass. Gardner and Gardner (1983) argued that as plants increased in size, a larger portion of the dry matter was grain. Prihar and Stewart (1990) presented evidence from the literature showing that harvest index was independent of the size of mature plants and in some cases, harvest index was observed to increase with decrease in plant size.

Crop simulation models are difficult to develop and often deficient in their predictions of grain growth. Hammer and Muchow (1994) developed a crop model for sorghum that accounted for 94% of the variation in total biomass, but only 64% of the

variation in grain yield when tested using data sets from a broad range of environments. The harvest index approach used a linear increase in harvest index with time from shortly after anthesis until two-thirds of the time between anthesis and physiological maturity had elapsed or a maximum harvest index of 0.55 had been reached (Hammer and Muchow, 1994). Hammer and Broad (2003) studied different maturity sorghum hybrids with contrasting phenology (early, medium, and late) and found that grain yield varied from 4,700 to 9,400 kg ha⁻¹. There was large and significant variation in yield among experiments, despite all being grown under non-limiting water and nutrient conditions. Grain yield is the product of total biomass and harvest index. Hammer and Broad (2003) found large yields were associated with great biomass production (14.3-17.6 t ha⁻¹) and harvest index (0.47-0.57). The harvest index values were close to the maximum harvest index of 0.55 reported as reflecting the genetic potential of most current sorghum hybrids (Hammer and Muchow, 1994). Hammer and Broad (2003) reported low yield associated with low biomass production (10.3 and 13.3 t ha⁻¹) and lower values of harvest index (0.42-0.46) that also contributed to substantially less yield. Hammer and Broad (2003) found differences in grain yield manifested through the difference in total biomass and harvest index; differences in total biomass and yield at maturity can be explained by differences in assimilation during grain-filling which was associated with greater incident radiation, warmer temperature, and slightly greater leaf area index. This reflected the enhanced amounts of light interception and growth between initiation and anthesis, which is known to influence kernel number in sorghum (Rosenthal et al., 1989) and other cereals (Fisher, 1985; Hammer and Broad, 1994).

Harvest index had a negative correlation with plant height and a positive correlation with grain yield, both phenotypically and genotypically (Can and Yoshida, 1999). Mohammad et. al. (1993) studied 54 genotypes of sorghum and found greater forage yield was associated with tallness, late maturity, greater tiller retention, and greater stover yield, but low crude protein and harvest index. High grain yield was associated with high harvest index, greater kernel number and size, but short height while crude protein was associated with high harvest index and short height; however, late maturity and taller plant height were associated with high harvest index (Mohammad et al., 1993). Performance of sorghum under rainfed conditions was significantly associated with retention of green leaf area, plant height, and maturity (Habyarimana et al., 2004). Like harvest index, green fodder and dry matter yield varied by cultivar (Gampawar et al., 2002). Also, grain yield and physiological traits related to development and vegetative growth were significantly negatively correlated in genotypes of *S. bicolor* (Soltani et al., 2001). However, there were significant positive correlations for growth rate, grain filling rate, and harvest index. According to Briggs and Knowles (1967), the heritability of quantitative characters was usually high, because breeding behavior could be predicted. High heritability coupled with genetic advance indicated that additive gene effects were operating, and selection for superior genotype was possible (Arunkumar et al., 2004). In addition to correlation and heritability, knowledge of genetic variability among different parameters contributing to yield was an important criterion for yield enhancement. Khan et al. (2005) observed that estimates of genetic variance were smaller than the respective phenotypic variance. Harvest index might be affected by environment and cultivar (Phihar and Stewart, 1991).

Chapter III

Materials and Methods

Germplasm

The study used sorghum conversion materials selected from extremes, high or low percentage of genome recovery, from $F_{2:3}$ and $BC_1 F_{2:3}$ progeny to determine combining ability for early testing. The experiments consisted of 44 parents (four female testers and four male lines in each of the 10 Reinstated Sorghum Conversion (RSC) families which produced 160 F_1 hybrids. It will determine yield potential based on converted lines and whether a relationship between combining ability and yield exists between various sorghum conversion populations and progeny.

In 2009-2011, populations from the RSC collections were developed at Puerto Vallarta, Mexico (a short-day sorghum nursery), under the direction of Dr. Fred Miller, using standard techniques to emasculate by hand, selected sorghum accessions and B.Tx406 as the female parent (Klein et al., 2016). The F_1 plants were self-pollinated at Puerto Vallarta, resulting in F_2 populations. Phenotypic self-pollinated selections were made based on plant height (<75 cm) and early flowering (<65 days) from the F_2 populations grown at Richardson Seeds LTD at Vega, TX. Twenty selections of early-maturing, dwarf F_2 progeny from each cross were genotyped with restriction-site-associated DNA sequencing technology developed by Morishige et al. (2013) and subsequently reported by Klein et al. (2013, 2016). Sorghum selections were grown at

USDA-ARS at College Station, TX, and seedling tissue was collected and evaluated via Illumina by Dr. Bob Klein.

Single nucleotide polymorphism (SNPs) across all 10 chromosomes was examined to identify F_2 descendants that recovered the greatest percentage of the tropical (exotic) parental genome and showed the least linkage disequilibrium (linkage drag) around loci under intense selection pressure (height and flowering time) (Klein et al., 2016). The F_2 progeny that showed the high percentage of whole-genome SNPs from the exotic parent were classified as having the high percentage of the tropical genome, and one genomic and phenotypically chosen F_2 descendant from each population was advanced by backcrossing to the tropical introduction parent.

The resulting BC_1F_1 was grown, self-pollinated, and subjected to the same rigorous phenotypic selection process. The resulting BC_1F_2 was grown, subjected to the same phenotypic and genomic selection processes to identify part of the whole based on the high or low percentage of exotic genome recovery. One BC_1F_2 plant from each population that showed the high proportion of the tropical genome was identified as described previously and self-pollinated. Grain from each of the $F_{2:3}$ and $BC_1F_{2:3}$ progeny rows were bulked for distribution and made available for the study. Table 1 shows the selected taxonomic groups based on the high or low percentage of genomic recovery in this study, along with the country of origin, sorghum race, and working group. The exotic male parent lines included several race and working group combinations; however, not all taxonomic classes were represented.

Table 1. Sorghum from the Reinstated Sorghum Conversion Program (RSC) with country of origin, race, working group, and percentage of exotic genome recovery value.

Classification				% Recovery		Value
RSC	Country of origin	Race	Working group	RSC	Recovery	
RSC73-0	Sudan	C	C	73	F ₂ High	0.489
				73	BC ₁ F ₂ High	0.660
				73	F ₂ Low	0.171
				73	BC ₁ F ₂ Low	0.530
				83	F ₂ High	0.468
				83	BC ₁ F ₂ High	0.650
RSC83-0	Sudan	C	C	83	F ₂ Low	0.116
				83	BC ₁ F ₂ Low	0.550
				112	F ₂ High	0.464
				112	BC ₁ F ₂ High	0.720
RSC112-0	Ethiopia	D	Nandyal	112	F ₂ Low	0.111
				112	BC ₁ F ₂ Low	0.470
				76	F ₂ High	0.463
				76	BC ₁ F ₂ High	0.840
RSC76-0	Sudan	CD	CD	76	F ₂ Low	0.113
				76	BC ₁ F ₂ Low	0.690
				38	F ₂ High	0.445
RSC38-0	Mali	DB	D-B	38	BC ₁ F ₂ High	0.750
				38	F ₂ Low	0.178
				38	BC ₁ F ₂ Low	0.590
				37	F ₂ High	0.280
RSC37-0	Sudan	C	C-Nigr	37	BC ₁ F ₂ High	0.720
				37	F ₂ Low	0.160
				37	BC ₁ F ₂ Low	0.510
				15	F ₂ High	0.274
RSC15-0	Sudan	CG	CG	15	BC ₁ F ₂ High	0.710
				15	F ₂ Low	0.081
				15	BC ₁ F ₂ Low	0.530
				124	F ₂ High	0.261
RSC124-0	Ethiopia	DK	D-K	124	BC ₁ F ₂ High	0.710
				124	F ₂ Low	0.116
				124	BC ₁ F ₂ Low	0.480
				117	F ₂ High	0.257
RSC117-0	Ethiopia	D	Nandyal	117	BC ₁ F ₂ High	0.680
				117	F ₂ Low	0.117
				117	BC ₁ F ₂ Low	0.440
				19	F ₂ High	0.201
RSC19-0	USA	B	C-Nigr	19	BC ₁ F ₂ High	0.560
				19	F ₂ Low	0.083
				19	BC ₁ F ₂ Low	0.300

Two $F_{2:3}$ progeny and two $BC_1F_{2:3}$ progeny from the F_2 populations of 10 RSC families were chosen based on the extremes of high to low percentage of genome recovery of the exotic parent. The A- (female) and B- (maintainer) lines were provided by Richardson Seeds LTD., and all were kafir types, 301 a four-dwarf early maturing Wheatland Redlan derivative kafir-Martin, 319 a three-dwarf medium early maturity red kafir-kafir, Tx3197 a public three-dwarf medium maturity Std kafir (standard check), and 338 a three-dwarf medium late maturity kafir-modified (unique/elite females with great combining ability). Parental crosses between the selected lines from the Sorghum Conversion Program and the four elite females, previously referenced, were made in the summer of 2014 and 2015 at Vega, TX, and during the winter of 2014 at Bucerias, Mexico. To achieve good synchrony (best “nick”) in flowering time of the male and female parents, all 44 parents were planted twice (15 days apart) in two-row plots (0.762 m spacing) 5.18 m long with 1.5 m borders at Vega, TX. Best agronomic practices were used across years for each location of the parental crossing. The panicles of female lines were bagged before stigmas were visible while unbagged RSC male parents were allowed to reach anthesis. For each combination of crosses, multiple panicles were pollinated to achieve an adequate quantity of kernels. Care was taken to use the most uniform $F_{2:3}$ and $BC_1F_{2:3}$ plants because of the heterozygous nature of the F_2 selections.

Experimental Procedures - Evaluation in Field

Research in the summers of 2015 and 2016 was at various locations (Table 2). In 2015, the 160 F_1 hybrids and four B-line parents corresponding to the A-line used in hybridization were evaluated at two locations, with two replications per location at Vega,

TX (latitude 35.12922 N, longitude -102.51716 E; 1241.9 m above sea level) on Pullman clay loam, and at Hutchinson, KS (latitude 38.13003 N, longitude -97.71266; 437.9 m above sea level) on Crete silt loam.

During the same summer, the 160 F₁ hybrids were planted at Taylor, TX (latitude 30.51867 N, longitude -97.49257 E; 189.1 m above sea level) on Branyon clay, and at Perryton, TX (latitude 36.32269 N, longitude -100.86301 E; 905.1 m above sea level) on Sherman clay loam (NRCS web soil survey) to evaluate and increase the environmental effect against commercial checks. Two groups of commercial checks with three maturity classes in each group consisting of early maturity and low yield, medium maturity and medium yield, and late maturity and high yield were provided by Richardson Seeds, Inc. and used for the research. In 2016, the same 160 F₁ hybrids, checks, and parents were planted at Vega and Dumas, TX (latitude 35.96557 N, longitude -101.93120 E; 1092.8 m above sea level) on Sherm silty clay loam (NRCS web soil survey) while the F₁ hybrids and checks were planted only at Hutchinson, KS. The sorghums, other than the checks, were grown in a completely randomized augmented split-block design with two replications at each location. The hybrids were blocked by female and the RSC family with checks arranged into two groups represented in each female block.

Table 2. Agronomic, environmental, and soil characteristics during two years in seven environments used to grow F₁ sorghum hybrids and parents to measure combining ability and heterosis.

Location	Soil type	Altitude -m-	Latitude -N-	Longitude -E-	Plot length -m-	Row spacing -m-	Date planted	Date harvested	Rainfall* -mm-
Vega Texas (2015)	Pullman clay loam	1241.9	35.12923	-102.51716	5.18	0.762	7 June	4-5 November	538.9
Perryton Texas (2015)	Sherman clay loam	905.1	36.32269	-100.86301	7.92	0.762	3 June	NA	940.8
Hutchinson Kansas (2015)	Crete silt loam	437.9	38.13003	-97.71266	5.18	0.762	26-27 June	7 November	404.6
Taylor Texas (2015)	Branyon clay	189.1	30.51867	-97.49257	5.18	0.762	1 April	4 September	538.7
Vega Texas (2016)	Pullman clay loam	1242.7	35.13222	-102.52174	5.18	0.762	12 May	17-18 October	215.1
Dumas Texas (2016)	Sherm silty clay loam	1092.8	35.96557	-101.93120	5.18	0.762	5-6 May	13-14 October	245.6
Hutchinson Kansas (2016)	Crete silt loam	437.9	38.13002	-97.71265	5.18	0.762	17 June	7 November	746.5

*Rainfall is the amount from 1 January until harvest.

At each location, an experimental unit was designated as two rows, with row length varying with location. Dependent upon location, the genotypes were evaluated for the following agronomic traits:

1. Days to mid-anthesis: number of days from the date of planting to the date when half the plants in a plot reached mid-anthesis.
2. Plant height: average distance in centimeters from the ground to the tip of the panicle at maturity.
3. Flag leaf height: average distance in centimeters from the ground to the flag leaf.
4. Panicle exertion: distance in centimeters from the ligule of the flag leaf to the base of the lowest panicle branch at maturity.
5. Panicle length: distance in centimeters from the lowest panicle branch to the tip of the panicle at maturity.
6. 1000-kernel weight: weight of 1000 kernels, measured in grams, from grain samples of three panicles hand harvested per plot, before harvesting by a combine.
7. Grain yield: weight of grain harvested per plot, expressed in kilograms per hectare (kg ha^{-1})
8. NIR was estimated by Dr. William Rooney at College Station/Bryan, TX, measuring grain protein, starch, fiber, and fat concentration (%), and moisture for calibration. Grain ash concentration was calculated but not analyzed.

Harvest and Grain Yield

In 2015 and 2016, the plots were harvested with an Allis-Chalmers Gleaner K2 combine harvester at Vega, Taylor, and Dumas, TX. The plots at Hutchinson, KS, were harvested with a Kincaid 8XP using a Harvest master Classic Grain Gage for plot weight. The plot yield was converted to Mg ha^{-1} after adjusting for 14% moisture, and a conversion factor based on row width and plot length was used, which differed across environments. In some environments, not all traits could be evaluated. Data on the number of days to anthesis were not collected at Perryton (2015), and no yield data were obtained because of the wet year. Data on panicle height, exertion, and length, and 1000-kernel weight were not collected at Hutchinson in 2015 and 2016 or Taylor in 2015. Data on total plant and flag leaf height, including panicle sampling for 1000-kernel weight were collected on both replications at Perryton and Vega in 2015 and at Dumas and Vega in 2016. Numbers of days to anthesis were determined at Vega during both years. Best agronomic practices were used at each location during both years.

Three-panicle Weight and 1000-kernel Weight

The three panicles harvested from each two-row plot were fully dried, threshed, weighed, and expressed to the nearest tenth of a gram. A key-mat seed counter (Model 946, St. Charles, IL) was used to determine 1000-kernel weight to thousandths of a gram of the kernels from the same three panicles.

Plant Height and Panicle Length

Plant heights were measured for each plot by randomly selecting three uniform plants from the two rows in each plot at Vega and Perryton in 2015 and at Vega and

Dumas in 2016. The length from the base of the plant to the tallest point of the tip of the panicle was measured, and the average of three plants was recorded per two-row plot. The heights to the flag leaf on the same three plants were measured from the base of the plant to the tallest point of the collar of the flag leaf. Three uniform panicles were randomly selected and harvested by hand, and panicle length was measured from the tip to the base of each panicle and averaged. Exsertion was extrapolated from the difference between the base of the panicle and the tip of the collar of the flag leaf and averaged for all three panicles.

Near-infrared Spectroscopy

Cereal grains are predominantly composed of carbohydrates, mostly in the form of starch, with considerable but variable amounts of protein as well as some lipids, vitamins, and minerals. Genetic and environmental effects create significant variation in the amount and quality of each constituent. Near-infrared spectroscopy (NIR) is widely applied in agriculture for determining the quality of forage, grain, grain products, and other products. It is much used to quantify the composition of agricultural products because it meets the criteria of being accurate, reliable, rapid, non-destructive, and inexpensive (Burns, 2007). Organic molecules have specific absorption patterns in the near-infrared region that can be used to estimate the chemical composition of the material being analyzed (Williams and Norris, 2001). Bulk whole-grain samples can be evaluated rapidly, require no sample preparation, and preserve the kernels after measurement for further analysis or for propagation (Velasco et al., 1999; Baye and Becker, 2004).

Statistical Analysis

The class of augmented experimental design was first introduced by Walter T. Federer in 1955 (Federer, 1961). An augmented experimental design is any standard design augmented with additional treatments in a complete or incomplete block. Mejza (1998) presented a class of split-block experimental designs wherein a check treatment represented one of the treatments for either or both of the two factors involved. The designs of Mejza had similarities to augmented experimental designs (Federer, 1993, 2002, 2004) and gave rise to a new class of augmented split-block experimental designs (Federer, 2004).

The sorghums, other than the checks, were grown in a completely randomized spilt-block design with two replications at each location. The hybrids were blocked by female and the RSC family, with checks arranged into two groups randomly represented in each female block. At each location, an experimental unit was designated as two rows, with row length varying with location (Table 2). In each environment, data were analyzed as a randomized complete block design with two replications per environment. Genotypes were considered fixed effects, while replications and environments were considered random effects.

The model used was $Y_{ijl} = \mu + e_l + r_{il} + g_j + (gl)_{jl} + e_{ijl}$

where Y_{ijl} = value of the ijl^{th} plot,

μ = grand mean,

e_l = effect of l -th environment, $l = 1, 2, 3, 4, 5$

r_{il} = effect of i -th replication at l^{th} environment, $i = 1, 2$

g_j = effect of j -th genotype, $j = 1, 2, \dots, 211$

$(ge)_{jl}$ = effect of interaction of j^{th} genotype with l^{th} environment

e_{ijl} = error associated with the ijl^{th} observation

Variation due to genotype was partitioned into variation within hybrids, parental lines, and check hybrids. Contrasts were analyzed between hybrids, parents, RSC families, and selections by percentage of exotic genome recovery using highest and lowest BC₁F_{2:3} and F_{2:3} generations. The percentage of exotic genome recovery was previously calculated for each exotic parent in the Sorghum Conversion Program used in the study (Table 1). Analysis of variance (ANOVA) across environments and for individual environments was done using PROC GLM 2002-2012 by SAS Institute Inc., Cary, NC, using line x tester design by Singh and Chaudhary (1985). Contrasts were analyzed to evaluate differences in combining abilities and heterosis between lines, testers, RSC families, and the generation to aid sorghum breeders in early detection of potential new hybrids. Bartlett's test for heterogeneity of error variances assessed the validity of combining the data from individual environments for a combined analysis (Little and Hills, 1978; Steel and Torrie, 1980). Heterogeneous error variances were calculated for all traits evaluated in the seven environments. Because there were no egregious problems with the data, the data from individual environments were combined for analysis in addition to the individual analysis.

Evaluation of the materials for heterosis and combining ability should provide sorghum breeders the information necessary to determine the impact on early testing based on the proportion of genetic recovery. The additional benefit is to determine if

there is a difference between sorghum conversion populations and their combining ability and genetic recovery. This will provide insight into early testing for sorghum breeders worldwide.

Line x Tester Analysis

The data recorded on the material generated as per the line x tester model of Kempthorne (1957) were subjected to analysis of variance as per the line x tester model by Singh and Chaudhary (1985).

Combining Ability Analysis and Heterosis

GCA effects of parents, SCA effects of BC₁F₁ and F₁ hybrids, the corresponding standard errors, and their mean squares were estimated using R Studio (R version 3.2.2 (2015-08-14) © 2015 The R Foundation for Statistical Computing Platform: x86_64_w64-mingw32/x64 (64 bit). Percentage of heterosis of all hybrids over the mid-parental value (midparent heterosis) was calculated using the following formulas:

$$\text{Midparent heterosis (MPH)} = (F_1 - MP) / MP \times 100$$

$$\text{Bestparent heterosis (BPH)} = (F_1 - BP) / BP \times 100$$

Rex Bernardo formula for combining ability.

General Combining Ability was calculated as $GCA_i = (Y_i - Y_{..} / Y_{..}) \times 100$

$$GCA_j = (Y_j - Y_{..} / Y_{..}) \times 100$$

Specific Combining Ability as $SCA_{ij} = ((Y_{ij} - Y_i - Y_j + Y_{..}) / Y_{..}) \times 100$

where Y_i = mean of i th male

Y_j = mean of j th female

Y_{ij} = mean of $j \times i$ hybrid

$Y_{..}$ = mean of all hybrids

R-studio model for combining ability.

The additive model used to estimate GCA and SCA effects of the ijk observation follows:

$$X_{ijk} = \mu + g_i + g_j + s_{ij} + e_{ijk}$$

where

X_{ijk} = any character measured of the cross ($i \times j$) in the k th replication

μ = population mean

g_i = *gca* effect of i th line

g_j = *gca* effect of j th tester

s_{ij} = *sca* effect of ($i \times j$)th cross

e_{ijk} = error associated with observation Y_{ijk}

i = number of lines

j = number of testers

k = number of replications

The variances for GCA and SCA were tested against their respective error variances, derived from the analysis of variance of the different traits based on Fellahi et al. (2013):

Covariance of half-sib of line

$$= \text{Cov.H.S.}(\text{line})$$

$$= \frac{Ml - Ml \times t}{rt}$$

Covariance of half-sib of tester

$$= \text{Cov.H.S.}(\text{tester})$$

$$= \frac{Mt - Ml \times t}{rl}$$

Covariance of full sib

$$= \text{Cov.F.S.}$$

$$= \frac{(Ml - Me) + (Mt - Me) + (Ml \times t - Me)}{3r} + \frac{6r \text{Cov.H.S.} - r(l+t) \text{Cov.H.S.}}{3r}$$

while Cov. H.S. (average) was calculated by the formula

$$\text{Cov. H.S. (average)} = 1/r [2lt - l - t] [(l-1)(Ml) + (t-1)(Mt)] / [l + t - 2 - Ml \times t]$$

Assuming no epistasis, variance caused by GCA (σ^2_{gca}) and variance caused by SCA (σ^2_{sca}) were calculated by:

$$\sigma^2_{\text{gca}} = \text{Cov. H.S.} = (1 + F/4) \sigma^2_A$$

$$\sigma^2_{\text{sca}} = (1 + F/2) 2D$$

Additive and dominance genetic variance (σ^2_A and σ^2_D) were calculated by taking inbreeding coefficient F equal to 1; that is, $F = 1$ because both lines and testers were considered inbred (Fellahi et al., 2013). Significance for GCA and SCA effects were determined by using a t -test. Midparent heterosis was estimated from mean values, and its significance was determined using a t -test.

Correlation Estimates

Pearson's correlation coefficients between all the traits were estimated using SAS statistical software. Correlations were estimated separately for parents, for hybrids, and for all the genotypes combined. Correlations were also estimated between indices of heterosis – SCA and midparent heterosis.

Chapter IV

Results and Discussion

Means of Genotypes

Means for the traits evaluated – number of days to anthesis, height to the flag leaf, total plant height, panicle exertion, panicle length, three-panicle weight, 1000-kernel weight, and grain yield – varied across environments (Table 3). The means are averages over the seven environments analyzed, barring a few traits for which data were not available in particular environments, as mentioned in the previous chapter. The two high grain yield averages of 8.52 and 8.24 Mg ha⁻¹ were for the commercial check hybrids 304/5 and 319/54, respectively. Testers included in the top 20 were A.319 with (seven) crosses, followed by A.301 (four), A.338 (two), and A.Tx3197 (one) crosses. Lines in the top 20 were RSC83-1 BC₁F_{2:3} selected as the low percentage of exotic genome recovered (low), followed by RSC83-14 BC₁F_{2:3} that appeared twice and was selected as the high percentage of exotic genome recovered (high). RSC117-4 BC₁F_{2:3} and RSC112-9 BC₁F_{2:3} also appeared twice, and both were selected for high percentage of exotic genome recovery (high). The hybrids required slightly fewer mean days to anthesis (74.3) compared to lines (76.3) and testers (74.5), but slightly more than checks (72.5).

Table 3. Means for 12 traits of 160 sorghum hybrids, 44 parental lines, and seven hybrid checks ranked by yield in seven environments - Vega, Perryton, and Taylor, TX, and Hutchinson, KS, in 2015, and Vega and Dumas, TX, and Hutchinson, KS, in 2016.

Yield rank	Genotype	Grain yield (Mg ha ⁻¹)	DTF	Flag leaf height (cm)	Total Panicle height (cm)	Panicle length (cm)	3-panicle weight (g)	1000-kernel weight (g)	Fat (%)	Fiber (%)	Protein (%)	Starch (%)
1	A.304/5	8.52	75.0	97.9	140.6	30.4	12.2	214.0	3.20	1.82	8.20	68.62
2	A.319/54	8.24	68.9	93.6	132.8	29.7	9.5	207.6	2.91	1.85	8.22	69.03
3	A.319*((RSC83)RSC83)-1	8.21	72.2	121.8	153.0	23.3	7.9	191.9	2.80	1.78	8.94	67.68
4	A.302/29	8.17	71.5	85.6	127.4	30.7	11.0	176.7	3.03	1.82	8.62	69.62
5	A.338*((RSC117)RSC117)-4	8.06	75.8	104.6	135.8	26.8	4.4	195.1	3.12	1.82	8.80	68.39
6	A.338/4	7.66	74.3	111.4	146.6	26.8	8.5	208.2	3.40	1.90	9.07	68.47
7	A.319/22	7.52	69.7	87.1	130.1	29.9	13.1	159.2	2.06	1.78	7.73	68.83
8	A.301*((RSC83)RSC83)-14	7.47	67.3	98.9	127.5	23.4	5.2	182.3	2.29	1.77	8.76	68.02
9	A.301/41	7.39	67.0	85.6	120.7	27.8	7.3	153.5	1.80	1.70	8.08	68.80
10	A.319*((RSC19)RSC19)-10	7.35	71.3	150.9	185.0	24.4	9.7	188.1	2.82	1.81	9.18	67.81
11	A.301*((RSC112)RSC112)-19	7.22	71.2	82.0	116.1	27.6	6.5	167.9	2.75	1.86	9.24	68.03
12	A.319*((RSC83)RSC83)-14	7.21	72.0	131.1	160.0	25.5	3.5	232.8	2.64	1.76	8.88	67.85
13	A.319*((RSC117)RSC117)-4	7.16	73.0	87.6	120.7	29.6	3.5	168.5	3.09	1.84	8.83	68.64
14	A.301*((RSC19)RSC19)-10	7.14	68.3	102.4	136.5	25.5	8.6	187.4	2.36	1.79	8.56	68.36
15	A.319*((RSC112)RSC112)-19	7.10	72.2	96.8	131.9	25.6	9.5	184.1	2.99	1.80	8.49	68.38
16	A.Tx3197*(RSC112)-5	7.00	71.3	119.2	160.8	27.6	14.0	188.6	2.36	1.61	8.22	69.13
17	A.301*((RSC37)RSC37)-12	6.99	67.7	117.5	149.7	22.7	9.5	195.0	2.55	1.88	9.27	67.22
18	A.338*(RSC117)-2	6.91	77.0	114.4	146.5	26.9	5.1	177.9	2.84	1.76	8.32	68.65
19	A.319*((RSC37)RSC37)-8	6.87	70.5	125.6	162.1	28.1	8.5	204.9	2.60	1.85	8.52	67.73
20	A.319*(RSC37)-12	6.86	74.0	135.5	169.8	24.3	10.1	207.4	3.01	1.77	9.27	67.39
21	A.301*((RSC37)RSC37)-8	6.81	65.2	87.6	123.8	23.5	12.7	150.4	2.43	1.87	8.65	67.86
22	A.301*((RSC19)RSC19)-17	6.81	67.5	120.3	152.9	25.7	6.9	183.4	3.01	1.82	9.81	67.21
23	A.Tx3197*((RSC117)RSC117)-4	6.80	69.0	90.1	128.4	29.1	9.1	199.7	2.98	1.72	8.75	68.62
24	A.319*((RSC37)RSC37)-12	6.79	71.0	126.7	158.9	26.0	6.2	231.3	2.74	1.89	9.10	67.53
25	A.338*((RSC117)RSC117)-3	6.72	79.0	106.8	137.4	25.3	5.4	172.3	2.93	1.74	8.59	68.42
26	A.301*(RSC112)-5	6.70	71.0	95.7	132.4	28.1	8.6	228.7	2.55	1.77	8.11	68.71
27	A.Tx3197*(RSC124)-9	6.69	70.2	91.3	128.3	29.1	7.9	165.4	2.40	1.65	8.33	68.66
28	A.338*((RSC83)RSC83)-14	6.66	75.7	153.1	185.3	22.6	9.6	217.8	2.69	1.69	8.79	67.99
29	A.Tx3197*((RSC112)RSC112)-19	6.66	70.8	110.5	146.5	25.2	10.8	191.0	2.64	1.61	7.89	69.59

30	A.301*(RSC124)RSC124)-16	6.62	72.2	88.7	128.0	28.3	11.1	199.6	25.9	2.57	1.77	8.09	68.49
31	A.Tx3197*((RSC83)RSC83)-1	6.61	69.5	142.8	175.4	23.1	9.5	216.1	31.6	2.59	1.73	8.73	67.94
32	A.301*(RSC76)RSC76)-2	6.59	71.2	145.9	175.3	22.5	6.9	221.9	37.1	1.76	1.72	7.33	69.56
33	A.301*(RSC83)RSC83)-1	6.57	66.7	97.4	127.7	22.3	7.9	163.4	30.7	2.17	1.76	8.63	68.38
34	A.Tx3197*((RSC112)RSC112)-15	6.53	70.5	105.7	144.6	27.1	11.9	190.7	29.1	2.64	1.63	8.29	69.01
35	A.319*(RSC73)-9	6.52	70.5	122.6	159.4	25.0	11.8	171.2	24.0	2.98	1.78	8.53	67.96
36	A.338*((RSC112)RSC112)-15	6.51	77.2	123.3	160.4	25.2	12.0	234.5	27.7	3.12	1.75	8.54	68.04
37	A.319*(RSC124)-9	6.50	69.2	87.5	126.8	30.8	8.5	149.8	23.5	2.51	1.76	8.27	68.42
38	B.Tx3197	6.49	70.7	86.4	112.3	23.8	2.2	128.6	27.9	2.39	1.72	7.84	69.51
39	(RSC19)-12	6.48	71.8	59.6	94.7	22.3	12.8	129.2	22.0	2.89	1.86	9.71	67.13
40	A.319*(RSC112)-5	6.45	72.3	106.8	144.6	28.2	9.6	191.3	28.0	2.63	1.75	7.96	68.75
41	A.319*(RSC73)RSC73)-6	6.41	71.2	117.6	149.6	23.4	8.6	156.5	23.6	2.80	1.76	8.56	67.87
42	A.301*(RSC124)RSC124)-4	6.41	69.5	82.8	113.5	25.7	4.9	180.6	27.9	2.37	1.74	8.90	68.04
43	A.319*(RSC117)RSC117)-3	6.39	72.2	73.1	113.5	28.2	12.2	212.6	25.6	2.99	1.76	8.75	68.58
44	A.319*(RSC38)-5	6.35	72.0	82.8	121.4	31.2	7.4	160.8	20.8	2.90	1.74	9.04	67.49
45	A.301*(RSC124)-9	6.34	68.0	71.8	109.0	28.6	8.6	159.0	27.6	2.25	1.74	8.84	68.13
46	A.Tx3197*((RSC124)RSC124)-16	6.30	70.5	131.9	168.2	27.1	9.2	193.8	25.6	2.36	1.56	8.24	68.91
47	A.338*(RSC124)-9	6.30	78.5	108.2	139.1	25.9	4.9	182.7	26.6	2.67	1.68	8.62	67.99
48	A.301*(RSC73)RSC73)-5	6.28	66.2	113.2	146.8	22.1	11.5	185.6	30.9	2.01	1.76	8.05	68.59
49	B.MMR338	6.23	78.0	99.5	128.6	22.0	7.1	153.7	25.4	2.88	1.83	8.40	68.74
50	A.301*(RSC38)RSC38)-15	6.23	65.0	116.1	143.4	23.3	4.0	179.5	28.1	2.76	1.76	9.27	67.20
51	A.319*(RSC117)-2	6.22	71.2	81.0	115.0	28.5	5.5	168.8	23.9	2.89	1.83	8.41	68.73
52	A.319*(RSC124)RSC124)-16	6.21	72.0	122.8	163.5	28.8	11.8	224.1	25.4	2.55	1.74	7.93	68.61
53	A.338*(RSC112)RSC112)-19	6.20	75.5	128.0	157.8	24.5	5.3	188.1	25.7	3.13	1.70	8.44	68.19
54	(RSC124)-9(RSC124)-4	6.20	83.2	86.9	127.4	26.6	13.9	119.6	22.8	2.84	1.66	9.08	67.63
55	A.Tx3197*((RSC15)RSC15)-15	6.19	67.8	99.2	141.5	27.8	14.6	159.6	28.8	3.02	1.73	9.55	68.47
56	A.301*(RSC73)-9	6.19	65.8	94.2	127.1	23.5	9.4	113.0	24.8	2.59	1.78	8.62	67.82
57	A.Tx3197*((RSC83)RSC83)-14	6.18	69.0	139.1	166.5	23.1	4.3	228.9	30.3	2.64	1.68	8.41	68.21
58	A.319*(RSC37)-7	6.18	71.7	138.8	178.7	25.4	14.5	208.3	25.0	2.79	1.80	8.55	67.73
59	A.319*(RSC83)-1	6.17	72.7	123.6	158.3	26.5	8.2	160.0	27.0	2.56	1.79	8.60	68.10
60	A.301*(RSC73)RSC73)-6	6.16	65.8	95.4	128.0	22.8	9.9	164.0	26.4	2.46	1.77	8.50	68.04
61	A.301*(RSC117)RSC117)-4	6.16	67.5	69.5	102.6	29.1	4.0	179.7	29.9	2.73	1.89	9.46	68.40

62	A.301*(RSC19)-3	6.15	65.5	93.4	136.9	26.9	16.5	155.2	25.3	2.67	1.86	9.36	67.64
63	A.301*(RSC112)RSC112)-15	6.14	66.7	70.2	103.8	25.8	7.7	184.4	31.1	2.79	1.84	9.36	67.81
64	A.Tx3197*(RSC73)-9	6.13	67.7	134.2	170.5	22.4	13.8	153.1	28.6	2.75	1.74	7.87	68.02
65	(RSC112)-5(RSC112)-15	6.12	74.0	91.4	130.5	26.5	12.6	124.5	29.4	2.50	1.76	9.17	68.34
66	(RSC19)-8(RSC19)-7	6.12	68.8	133.1	169.0	24.8	11.2	126.7	21.6	2.91	1.68	9.56	68.34
67	A.301*(RSC15)-13	6.12	65.5	113.2	144.5	22.4	9.0	122.6	29.1	2.90	1.77	9.90	68.30
68	A.301*(RSC76)-4	6.11	68.3	140.6	167.3	20.5	6.2	198.3	37.0	1.78	1.67	7.73	69.05
69	A.319**((RSC112)RSC112)-15	6.10	71.3	96.1	134.5	27.5	10.9	164.7	27.8	2.81	1.82	8.61	68.45
70	A.338**((RSC83)RSC83)-1	6.09	76.2	154.7	185.4	21.1	9.6	218.1	29.2	2.50	1.75	8.28	68.37
71	A.301*(RSC117)RSC117)-3	6.09	68.3	91.0	127.2	26.3	10.0	160.6	29.3	2.87	1.86	9.81	67.56
72	A.301*(RSC117)-2	6.06	66.3	71.7	104.1	29.0	3.4	164.7	27.8	2.37	1.82	8.96	68.55
73	A.338*(RSC83)-10-10	6.06	76.5	110.1	144.9	24.5	10.3	163.6	28.9	3.07	1.82	9.11	67.65
74	A.Tx3197*(RSC124)RSC124)-4	6.06	70.5	104.9	138.4	24.8	8.8	150.0	26.5	2.46	1.67	8.05	68.67
75	A.319*(RSC117)-10-10	6.04	71.3	74.7	115.4	27.7	13.1	164.9	23.0	2.89	1.76	8.82	67.92
76	A.301*(RSC76)RSC76)-16	6.03	71.3	141.3	173.6	21.7	10.7	213.4	37.2	1.80	1.73	7.76	69.37
77	(RSC37)-2(RSC37)-12	6.01	77.0	94.4	117.1	24.3	-1.6	124.2	25.0	2.30	1.78	8.32	67.81
78	A.338*(RSC112)-5	6.00	73.3	103.3	140.1	23.1	13.7	159.4	32.2	2.68	1.78	8.37	68.34
79	A.338*(RSC83)-1	5.99	76.0	163.8	194.5	22.7	8.0	224.4	30.2	2.55	1.73	8.31	68.13
80	B.MMR319	5.99	68.7	70.7	104.3	26.0	7.6	132.3	20.8	2.41	1.96	8.20	68.80
81	A.319*(RSC73)RSC73)-5	5.99	74.7	154.5	189.8	23.7	11.6	163.6	27.7	2.31	1.79	8.13	68.40
82	A.338*(RSC37)RSC37)-8	5.98	75.7	165.8	194.6	23.2	5.6	248.6	25.2	2.68	1.82	8.25	68.02
83	A.338*(RSC19)-3	5.97	74.8	141.2	184.7	26.4	17.2	211.4	28.4	2.73	1.81	8.89	67.71
84	A.319*(RSC124)RSC124)-4	5.95	76.3	92.5	131.1	28.8	9.8	155.1	21.6	2.55	1.81	8.27	68.45
85	A.301*(RSC38)-8	5.91	65.3	93.6	130.1	22.3	14.3	152.6	28.2	2.23	1.69	8.40	68.59
86	A.319*(RSC76)RSC76)-16	5.90	74.0	174.7	212.5	23.9	13.9	262.4	33.1	2.26	1.79	8.29	68.49
87	A.338*(RSC124)RSC124)-4	5.90	76.0	155.9	190.9	23.8	11.2	162.7	28.4	2.72	1.73	8.71	68.13
88	A.319*(RSC15)-11	5.89	72.0	134.3	174.8	26.4	14.1	138.7	23.3	3.21	1.86	9.25	68.49
89	A.301*(RSC83)-1	5.89	67.5	96.2	123.3	22.6	4.5	149.4	28.0	2.21	1.79	8.70	68.26
90	A.301*(RSC37)-12	5.88	64.0	82.0	117.9	25.4	10.5	152.4	27.2	2.62	1.82	9.00	67.51
91	(RSC76)-4(RSC76)-2	5.87	81.2	149.0	175.8	20.0	6.8	184.7	34.8	1.41	1.57	7.38	70.42
92	A.319*(RSC38)-8	5.87	70.2	138.2	172.9	24.5	10.1	207.7	25.7	2.71	1.77	8.19	68.29

93	A.Tx3197*(RSC37)RSC37)-12	5.86	68.8	151.8	189.7	23.6	14.2	208.9	28.7	2.68	1.76	7.65	68.12
94	A.338*(RSC124)-RSC124)-16	5.82	76.5	163.8	200.1	27.1	9.1	233.5	28.6	2.83	1.70	8.21	68.28
95	A.319*(RSC124)-3	5.81	69.2	102.2	144.4	30.0	12.2	139.6	24.9	2.52	1.78	8.96	68.16
96	A.338*(RSC117)-10	5.78	76.0	97.2	131.5	25.7	8.6	185.0	27.3	2.75	1.72	8.42	68.31
97	(RSC19)-8(RSC19)-10	5.78	76.3	122.1	154.7	22.6	10.0	109.6	18.8	2.16	1.73	8.98	68.83
98	A.319*(RSC19)-12-1	5.77	74.3	75.6	112.6	27.8	9.2	140.9	20.9	2.96	1.86	8.91	67.96
99	A.Tx3197*(RSC112)-8	5.75	70.8	92.9	130.2	24.4	13.0	146.9	29.1	2.02	1.62	7.89	69.50
100	(RSC117)-2(RSC117)-4	5.72	75.5	80.3	104.6	28.0	-3.6	143.0	35.2	3.12	1.86	10.07	67.95
101*	A.338*(RSC37)-12	5.72	71.5	129.8	166.0	23.8	12.3	175.7	26.2	2.73	1.85	8.56	67.83
102	A.301*(RSC15)RSC15)-15	5.72	66.7	78.2	113.6	26.8	8.6	150.8	27.7	2.83	1.84	10.00	68.25
103	A.Tx3197*(RSC117)RSC117)-3	5.72	70.7	100.6	130.2	26.3	3.3	209.3	29.9	3.08	1.70	8.58	68.14
104	A.319*(RSC38)RSC38)-15	5.71	70.5	155.1	188.1	24.7	8.4	189.7	25.5	3.05	1.68	8.89	67.50
105	A.338*(RSC15)-13	5.71	74.2	163.9	194.4	22.6	7.8	164.6	32.5	3.27	1.76	10.15	67.72
106	(RSC73)-9(RSC73)-6	5.70	68.3	66.0	81.9	19.9	-4.1	58.2	21.4	2.03	1.61	8.38	68.97
107	A.319*(RSC112)-8	5.69	72.2	87.1	121.0	26.8	7.1	150.8	25.9	2.56	1.80	8.40	68.75
108	A.301*(RSC15)-11	5.68	65.3	142.4	180.6	24.5	13.8	135.5	27.6	3.04	1.83	10.63	67.70
109	A.301*(RSC76)-13	5.67	68.0	135.7	169.7	22.2	11.8	148.4	32.8	2.09	1.77	8.67	68.65
110	A.Tx3197*(RSC19)-3-3	5.65	69.8	125.2	169.8	27.3	17.3	173.2	27.2	2.53	1.77	8.16	68.26
111	A.319*(RSC15)-13	5.64	68.0	93.0	134.1	27.6	13.5	142.9	27.0	2.94	1.78	9.53	68.68
112	A.Tx3197*(RSC37)-12	5.64	67.8	131.9	167.4	22.4	13.0	130.9	25.8	2.60	1.79	7.93	68.38
113	A.338*(RSC15)-11	5.64	73.8	171.9	202.9	23.8	7.2	181.3	29.0	3.50	1.86	10.24	67.31
114	A.Tx3197*(RSC117)-2-2	5.62	69.2	98.6	132.0	23.1	10.3	140.6	31.5	2.64	1.69	8.54	68.77
115	A.Tx3197*(RSC38)RSC38)-9	5.59	66.3	134.5	166.9	26.1	6.4	159.1	28.6	2.67	1.74	8.36	67.58
116	A.338*(RSC19)RSC19)-10	5.59	76.8	173.7	202.5	20.9	7.9	152.6	28.2	2.61	1.80	9.08	67.74
117	A.338*(RSC15)RSC15)-14	5.58	73.7	163.2	194.9	22.6	9.1	190.3	32.7	3.13	1.80	10.22	67.82
118	A.338*(RSC112)-8	5.57	77.5	106.5	139.3	24.8	7.9	165.7	26.2	2.88	1.76	8.39	68.32
119	A.319*(RSC76)-13	5.57	71.0	162.7	198.4	23.8	11.9	187.0	32.1	2.25	1.74	8.65	68.34
120	A.319*(RSC38)RSC38)-9	5.57	66.7	118.1	158.5	30.7	9.7	179.1	24.0	2.94	1.81	9.02	67.36
121	A.338*(RSC73)RSC73)-6	5.54	74.0	152.7	182.9	20.8	9.4	213.4	30.9	2.74	1.73	8.14	68.32
122	A.301*(RSC124)-3	5.52	67.7	86.1	123.5	28.0	9.5	135.8	30.8	2.12	1.70	8.88	68.55
123	A.319*(RSC83)-10	5.52	73.2	87.2	120.4	27.9	5.4	170.2	22.6	3.11	1.85	9.54	67.44
124	A.301*(RSC19)-12	5.49	66.5	64.5	104.7	24.0	16.2	143.8	25.2	2.48	1.84	9.08	68.04

125	A.Tx3197*((RSC37)RSC37)-8	5.48	67.0	137.4	174.0	24.4	12.2	223.1	25.8	2.60	1.72	7.32	68.90
126	(RSC37)-7	5.47	73.2	94.7	129.5	23.6	11.2	123.4	24.9	2.73	1.64	8.45	67.48
127	A.319*(RSC19)-3	5.46	71.8	117.0	161.6	29.2	15.4	131.7	23.4	2.95	1.88	9.33	67.49
128	A.Tx3197*((RSC117)-10-10	5.46	68.2	83.5	120.4	25.9	11.0	162.7	28.9	2.49	1.64	8.32	68.74
129	A.Tx3197*((RSC73)RSC73)-6	5.46	67.3	127.9	156.9	22.6	6.5	160.4	27.7	2.70	1.71	7.60	68.43
130	A.301*(RSC117)-10	5.46	67.8	59.7	92.6	25.5	7.4	155.3	28.1	2.28	1.75	8.78	68.52
131	A.301*(RSC83)-10	5.44	67.7	70.1	105.8	26.1	9.6	150.4	29.4	2.47	1.85	9.52	67.83
132	(RSC83)-1	5.44	75.5	85.0	112.8	22.4	5.5	132.9	29.8	2.42	1.75	8.80	68.12
133	A.Tx3197*((RSC38)-5	5.42	68.7	88.5	125.9	26.8	10.7	156.3	25.8	2.75	1.63	8.53	67.63
134	A.319*((RSC19)RSC19)-7	5.40	71.7	167.8	205.9	25.7	12.4	159.8	25.0	3.23	1.78	10.09	67.56
135	A.338*((RSC15)RSC15)-15	5.40	74.7	119.4	156.2	27.8	9.1	197.7	28.7	3.18	1.84	9.31	68.09
136	344/79	5.40	58.2	75.5	116.2	26.9	13.9	132.2	27.5	2.45	1.69	9.25	69.36
137	A.301*((RSC15)RSC15)-14	5.39	66.0	106.5	140.3	24.1	9.6	155.0	29.7	2.79	1.80	9.71	68.90
138	A.Tx3197*((RSC73)RSC73)-5	5.38	70.8	170.8	209.1	21.9	16.5	166.1	30.1	1.85	1.72	8.29	68.92
139	A.319*((RSC15)RSC15)-14	5.37	68.8	141.8	177.1	24.0	11.3	139.8	28.3	3.13	1.82	9.51	68.79
140	(RSC124)-9(RSC124)-16	5.37	79.2	132.9	168.8	27.6	8.2	181.6	27.2	2.35	1.62	9.11	67.97
141	(RSC117)-2(RSC117)-3	5.32	76.8	72.5	100.8	26.2	2.0	107.3	30.4	2.98	1.80	10.28	66.92
142	A.301*((RSC38)RSC38)-9	5.32	65.2	86.2	122.4	27.8	8.4	158.5	27.5	2.62	1.76	8.99	67.67
143	(RSC38)-5(RSC38)-15	5.32	71.8	124.0	151.8	23.0	4.7	119.0	22.6	2.72	1.61	9.31	67.52
144	A.319*(RSC73)-1	5.29	72.3	128.8	161.2	23.4	8.9	179.2	26.8	2.49	1.81	8.74	67.99
145	A.338*(RSC73)-9	5.28	74.8	157.2	193.0	21.7	14.2	212.1	29.7	2.73	1.70	8.47	67.93
146	A.338*((RSC37)RSC37)-12	5.28	74.5	171.5	203.9	23.2	9.2	231.9	27.8	2.70	1.78	8.30	68.05
147	(RSC19)-3	5.28	69.7	103.2	138.4	23.0	12.2	123.5	24.9	2.28	1.85	9.34	67.79
148	A.Tx3197*((RSC38)-8-8	5.27	68.2	146.9	182.9	23.3	12.7	154.7	29.6	2.47	1.68	8.51	68.44
149	(RSC38)-8	5.27	72.5	116.0	146.2	22.1	8.1	141.9	23.2	2.53	1.65	8.79	68.05
150	(RSC112)-8	5.26	77.5	78.0	113.7	24.0	11.7	111.5	26.1	2.77	1.70	8.74	68.09
151	(RSC112)-5	5.19	77.5	88.8	134.3	28.7	16.9	124.2	26.1	2.24	1.61	8.84	68.19
152	B.MMR301	5.19	64.0	59.6	92.0	23.6	8.8	132.6	27.1	2.33	1.85	8.59	68.91
153	A.301*(RSC112)-8	5.18	66.7	69.8	104.6	26.4	8.4	186.1	28.0	2.44	1.81	8.94	68.95
154	A.319*((RSC15)RSC15)-15	5.18	70.5	94.7	129.2	29.1	5.4	162.2	24.2	3.04	1.89	8.85	68.66
155	(RSC73)-1	5.18	67.3	91.2	119.7	23.8	4.8	111.4	25.6	1.88	1.68	8.76	68.56

156	A.338*(RSC19)-12-1	5.18	75.2	89.0	127.3	24.2	14.1	148.7	24.8	2.98	1.81	8.90	67.89
157	(RSC37)-2(RSC37)-8	5.17	72.8	81.3	102.2	25.6	-4.7	127.6	24.1	2.18	1.72	7.97	68.36
158	A.Tx3197*(RSC15)RSC15)-14	5.17	67.8	141.6	177.0	25.8	9.6	117.9	29.5	2.96	1.70	9.83	68.33
159	A.301*(RSC73)-1	5.16	65.2	85.0	119.4	23.7	10.7	140.1	27.2	2.35	1.75	8.57	68.52
160	(RSC112)-5(RSC112)-19	5.14	81.8	91.6	125.4	22.8	11.1	148.0	29.9	2.86	1.65	9.38	67.71
161	A.Tx3197*(RSC73)-1	5.10	72.0	143.1	173.9	22.8	8.0	164.5	29.3	2.35	1.73	8.35	68.33
162	(RSC38)-5	5.10	71.0	65.8	99.0	27.4	5.8	87.4	19.7	2.67	1.59	8.74	67.56
163	A.Tx3197*(RSC83)-10	5.09	72.3	92.9	128.0	26.3	8.8	125.7	30.4	2.64	1.76	9.00	68.22
164	(RSC117)-2	5.08	70.7	71.0	104.1	25.4	7.8	103.5	27.0	2.52	1.77	9.03	68.64
165	A.319*(RSC76)-4	5.08	73.7	178.4	211.8	22.6	10.8	225.4	32.4	1.97	1.72	7.98	68.42
166	A.301*(RSC37)-7	5.07	64.5	69.1	101.0	23.9	8.0	142.2	24.9	2.36	1.75	8.65	68.13
167	(RSC15)-13(RSC15)-14	5.07	74.2	103.3	135.1	20.2	11.6	86.2	26.9	2.80	1.83	10.12	68.06
168	(RSC76)-13	5.06	70.0	139.7	162.6	18.8	4.1	107.5	30.3	1.93	1.67	8.11	69.01
169	A.338*(RSC73)-1-1	5.01	75.5	165.5	198.1	20.5	12.1	162.2	30.5	2.79	1.77	8.59	68.07
170	A.338*(RSC37)-7-7	5.00	76.8	177.7	210.4	22.0	10.7	220.1	28.6	2.74	1.73	8.28	68.02
171	(RSC124)-3	5.00	72.3	140.8	184.2	26.7	16.7	94.0	25.7	2.38	1.66	9.58	67.72
172	(RSC76)-4(RSC76)-16	4.98	77.3	167.4	204.9	18.3	19.2	163.2	33.6	1.65	1.68	7.59	70.28
173	(RSC83)-1(RSC83)-14	4.98	74.3	65.8	85.9	18.2	2.0	121.8	25.1	1.89	1.59	8.78	68.54
174	(RSC37)-12	4.95	69.0	59.2	85.2	23.5	2.5	88.9	23.7	2.36	1.69	9.08	67.59
175	(RSC15)-13	4.92	68.5	103.7	135.2	22.9	8.6	93.9	27.5	2.61	1.65	10.38	68.39
176	(RSC83)-10	4.88	77.8	80.3	117.3	26.1	10.9	126.0	20.8	2.69	1.83	8.76	68.12
177	A.338*(RSC19)RSC19)-7	4.87	74.2	188.5	222.0	23.5	10.0	209.5	26.1	3.60	1.74	9.56	67.67
178	A.338*(RSC73)RSC73)-5	4.87	77.2	200.0	235.9	21.9	14.1	215.2	29.0	2.29	1.74	7.92	68.44
179	(RSC117)-10	4.85	72.8	63.8	97.4	24.4	9.1	110.8	23.0	2.84	1.64	9.01	67.23
180	A.Tx3197*(RSC76)RSC76)-2	4.85	71.2	204.8	242.0	23.2	14.1	263.7	35.1	1.86	1.65	7.82	69.63
181	(RSC15)-13(RSC15)-15	4.81	75.7	84.0	120.5	28.3	8.3	94.7	27.8	3.18	1.99	10.99	66.47
182	A.301*(RSC38)-5	4.78	64.3	61.4	95.5	28.2	6.0	136.2	24.3	2.45	1.75	9.30	67.71
183	A.338*(RSC124)-3	4.77	76.8	140.4	182.9	25.2	17.3	166.5	28.5	2.59	1.74	8.40	68.34
184	A.Tx3197*(RSC37)-7	4.75	68.8	142.8	181.9	23.4	15.7	170.1	29.7	2.54	1.74	8.16	68.22
185	(RSC38)-5(RSC38)-9	4.70	65.0	101.6	128.5	26.6	0.3	126.8	25.5	2.40	1.64	8.74	67.78
186	A.319*(RSC76)RSC76)-2	4.68	75.2	178.1	219.9	25.0	16.7	267.1	30.7	1.91	1.72	7.49	68.91

187	A.338*((RSC38)RSC38)-15	4.66	76.3	215.9	241.3	22.5	2.9	193.7	28.9	3.06	1.64	8.77	67.26
188	A.Tx3197*(RSC19)-12-1	4.65	69.8	80.2	116.3	24.5	11.6	135.1	26.0	2.47	1.76	8.45	68.50
189	A.Tx3197*(RSC76)-4	4.58	72.7	194.2	227.9	21.7	12.0	199.0	34.4	1.71	1.66	8.34	68.96
190	(RSC73)-9(RSC73)-5	4.58	76.5	100.6	122.8	18.3	3.9	126.1	24.1	1.63	1.68	8.18	68.80
191	A.Tx3197*((RSC19)RSC19)-10	4.52	70.7	159.7	189.3	22.5	7.1	184.8	27.5	2.51	1.72	8.57	68.88
192	A.Tx3197*((RSC38)RSC38)-15	4.50	67.2	167.6	200.5	21.6	11.3	169.6	29.1	2.79	1.65	9.17	67.05
193	(RSC76)-4	4.48	76.8	146.3	167.4	19.7	1.4	140.8	32.4	1.52	1.54	7.59	69.66
194	A.Tx3197*((RSC19)RSC19)-7	4.46	72.8	164.7	204.3	24.2	15.4	184.5	28.4	3.29	1.77	9.06	67.82
195	A.Tx3197*(RSC124)-3	4.40	69.3	125.6	164.5	25.7	13.3	126.4	29.3	2.16	1.66	8.72	68.48
196	A.338*(RSC38)-8	4.39	76.8	179.6	212.2	22.6	10.0	270.8	29.9	2.69	1.70	8.43	68.17
197	A.Tx3197*((RSC76)RSC76)-16	4.35	71.3	198.3	240.7	22.5	20.0	236.9	34.9	1.76	1.68	7.87	69.75
198	A.Tx3197*(RSC83)-1	4.25	74.7	130.5	164.4	25.0	8.8	182.6	30.9	2.58	1.71	8.50	68.11
199	A.338*((RSC38)RSC38)-9	4.14	75.8	167.9	199.3	24.4	6.9	169.2	28.5	2.93	1.75	8.72	67.47
200	(RSC73)-9	4.14	66.7	74.2	95.7	23.0	-1.6	95.6	17.8	2.55	1.66	8.23	68.05
201	A.338*(RSC76)-13	4.07	75.5	189.4	220.3	22.0	8.9	188.3	34.2	2.46	1.76	8.68	68.07
202	(RSC83)-1(RSC83)-1	4.06	71.8	65.1	80.4	19.3	-4.0	96.9	26.6	2.10	1.64	9.28	68.06
203	A.338*((RSC76)RSC76)-2	4.01	77.8	217.0	252.9	23.4	12.4	305.0	35.4	1.94	1.67	7.54	69.14
204	A.Tx3197*(RSC15)-13	4.01	66.7	146.8	185.6	22.8	16.0	106.4	30.1	2.94	1.65	10.12	68.07
205	A.Tx3197*(RSC15)-11	3.92	68.3	154.8	194.5	24.6	15.1	147.1	27.4	3.13	1.73	9.51	68.34
206	A.Tx3197*(RSC76)-13	3.77	71.2	180.9	213.1	23.1	9.1	158.7	31.7	2.12	1.71	8.38	68.90
207	A.338*(RSC38)-5	3.54	75.0	179.6	212.8	25.0	8.2	192.7	26.7	3.01	1.70	9.08	67.26
208	(RSC15)-11	3.50	67.7	160.4	200.4	21.2	18.8	83.0	21.8	3.23	1.80	11.22	67.09
209	A.338*(RSC76)-4	3.34	75.3	221.1	251.6	21.3	9.2	219.5	36.2	1.71	1.68	7.60	69.00
210	(RSC124)-9	3.30	77.2	84.8	121.1	28.1	8.3	90.7	23.6	2.53	1.64	8.69	67.71
211	A.338*((RSC76)RSC76)-16	3.07	77.2	221.0	255.6	22.0	12.6	234.3	37.2	2.13	1.70	7.86	68.77

DTF = days to flowering

†Exs = plant exsertion

Analysis of Variance

In the combined analysis, differences among levels of effects were detected for most sources of variation (Table 4). Grain yield was obtained for six environments; 2015 was a very wet year at Perryton, and the grain could not be harvested; however, this was the only common dependent variable among the six environments, and removing parents and checks allowed for an overall grain yield ANOVA. Of that, only four environments had parents, and traits could not be evaluated in every environment (number of days to anthesis, height to the flag leaf, total plant height, panicle length, three-panicle weight, 1000-kernel weight, and quality data), for which reason, degrees of freedom were modified for those traits in the analysis of variance across environments with parents. This also was reflected in the ANOVA tables for individual environments. By removing parents and checks, a combined mean squared analysis showed significant differences for environment, replication (environment), hybrids, lines, testers, RSC, selections, the interaction between hybrids, lines, tester x environment, and line x tester interaction ($P < 0.01$) for almost every trait considered. Line x tester x environment interaction was significant ($P < 0.01$) for grain yield, whereas total plant height, height to the flag leaf, panicle length, and percentage of fat concentration in grain had significant interaction at $P < 0.05$. The interaction between RSC x environment showed significant differences ($P < 0.01$) for grain yield, total plant height, height to the flag leaf, number of days to anthesis, and percentage of starch, fiber, and fat concentration in grain, but only significant differences at the $P < 0.05$ for plant exsertion, panicle length, three-panicle weight, 1000-kernel weight, and the percentage of concentration of protein in grain. RSC

Table 4. Mean squares of grain yield and agronomic traits of sorghum hybrids in seven environments – Vega, Perryton, and Taylor, TX, and Hutchinson, KS, in 2015, and Vega and Dumas, TX, and Hutchinson, KS, in 2016. Ten hybrids and yield results from Perryton (2015) and NIR data of one hybrid sorghum at Vega (2016) are missing.

Source of variation	Df†	Grain yield Mg ha ⁻¹	Total height cm	Flag leaf height cm	Plant exsertion cm	Panicle length cm	3-panicle weight g	kernel weight g	Df	Days to anthesis	Df	Protein% [‡]	Starch% [‡]	Fiber% [¶]	Fat% [¶]
Environment (E)	5	1095.5**	3 146003.3**	83577.8**	5682.3**	1598.6**	1009747.3**	13.9	2	11343.3**	3	4.45**	4.87**	0.48**	7.39**
Rep (E)	6	13.6**	4 523.8*	834.6**	853.6**	39.1**	17229.4**	95.9**	3	42.6**	3	7.47**	6.65**	0.03**	0.50**
Hybrids (H)	159	9.2**	159 11715.8**	12200.1**	84.5**	48.9**	10070.1**	78.5**	159	79.5**	159	2.38**	1.92**	0.02**	0.81**
H X E	795	3.3**	467 441.7**	433.4**	39.4**	4.9**	2630.6**	7.9*	318	17.8**	466	0.32*	0.35*	0.00**	0.07**
Lines (L)	39	18.2**	39 28079.0**	29718.4**	186.1**	138.3**	22644.4**	221.8**	39	41.8**	39	7.46**	5.66**	0.04**	2.65**
Testers (T)	3	73.7**	3 205521.7**	211615.2**	457.2**	455.7**	95806.2**	782.3	3	3140.1**	3	6.77**	5.82**	0.35**	5.92**
L X T	117	4.5**	117 1151.3**	1150.6**	40.0	8.3**	3340.7**	12.3**	117	12.8**	117	0.63**	0.58**	0.01**	0.07**
L X E	195	5.2**	117 900.6**	919.8**	44.4*	6.9**	3394.2**	9.8**	78	10.2*	117	0.44**	0.60**	0.00**	0.10**
T X E	15	11.6**	9 2668.3**	3021.9**	174.3**	16.0**	7862.9**	25.4**	6	548.8**	9	2.68**	2.02**	0.05**	0.23**
L X T X E	585	2.5**	341 227.0*	201.2*	34.2	4.1*	2247.0	6.9	234	6.6	340	0.21	0.21	0.00	0.06*
RSC†	9	45.0**	9 82873.1**	86108.5**	391.6**	376.6**	41364.6**	751.6**	9	122.1**	9	23.85**	18.04**	0.12**	8.03**
RSC X E	45	7.6**	27 2663.3**	2744.4**	61.0*	11.8*	5337.2*	16.4*	18	23.0**	27	0.66*	1.33**	0.01**	0.26**
RSC X T	27	10.1**	27 2808.8**	2817.8**	42.8	12.7*	3837.3	20.7**	27	17.0*	27	1.39**	1.41**	0.01**	0.14*
RSC X T X E	135	3.8**	81 428.5	398.9	47.9*	5.9	2695.9	9.1	54	9.8	81	0.19	0.30	0.00	0.08
Selections (S)δ	3	47.8**	3 18400.2**	24354.3**	486.6**	18.0	69201.5**	55.6*	3	13.5	3	0.87	1.09	0.00	1.62**
S X E	15	6.3*	9 348.4	436.6	65.0	6.9	2012.1	6.4	6	9.3	9	0.70	0.50	0.00	0.25
S X T	9	2.2	9 376.7	360.0	49.8	4.4	812.0	9.1	9	17.9	9	0.09	0.40	0.00	0.05
S X T X E	45	3.3	27 126.2	109.3	30.1	3.7	2342.3	9.4	18	7.9	27	0.23	0.24	0.00	0.06
Error	947	1.99	615 168.04	158.41	34.2	3.45	1915.8	6.01	473	7.57	456	0.27	0.26	0.00	0.05

DfE = days to flowering.

*Significant at the 0.05 probability level.

**Significant at the 0.01 probability level.

†Separate columns for degrees of freedom for particular traits are because of differences in the number of replications or environments from which data were collected for the traits.

‡RSC represents different lines from the Reinstated Sorghum Conversion (RSC) program and their respective families used in the study.

δSelections (S) are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2,3}, BC₁F_{2,3}, and the Low F_{2,3}, BC₁F_{2,3}.

¶Content in grain

x tester interaction showed significance ($P < 0.01$) for grain yield, total plant height, height to the flag leaf, 1000-kernel weight, and percentage of concentration of protein, starch, and fiber in grain but also showed significance at $P < 0.05$ for panicle length, number of days to anthesis, and percentage of concentration of fat in grain. RSC x tester x site interaction was significant at $P < 0.01$ for grain yield but only $P < 0.05$ for plant exsertion. The sorghum selections, which represent generations with high or low percentage of exotic genome recovery, were significant at $P < 0.01$ for grain yield, total plant height, height to the flag leaf, plant exsertion, three-panicle weight, and percentage of concentration of fat in grain, but significantly different only at $P < 0.05$ for 1000-kernel weight and interaction between selection x environment for grain yield. No significant differences were found for selection x tester or selection x tester x site. The significant differences indicated sufficient genetic variability among the hybrids and will be further partitioned into a line x tester analysis of variance with parents and checks.

In the combined line x tester analysis of variance with parents and checks, differences among levels of effects were detected for most sources of variation (Table 5). Environment, genotypes, hybrids, lines, testers, RSC, parents, and checks were significant ($P < 0.01$) for almost all the traits evaluated. Genotype x environment interaction was significant for all traits with at least a $P < 0.05$ difference. Replication (environment) showed significant differences at $P < 0.01$ for the height to the flag leaf, plant exsertion, panicle length, 1000-kernel weight, number of days to anthesis, and percentage of concentration of protein, starch, fiber, and fat in grain, and a significant difference at $P < 0.05$ was found for three-panicle weight. Hybrid x environment

Table 5. Mean squares of grain yield and agronomic traits of sorghum hybrids, including parents and checks, in four environments -Vega, TX, and Hutchinson, KS, in 2015, and Vega and Dumas, TX, in 2016.

Source of variation Environment (E)	Df†	Grain yield (Mg ha ⁻¹)	Total height (cm)	Flag leaf height (cm)	Plant		Panicle length (cm)	3-panicle weight (g)	1000- kernel		Days to anthesis	Df	Protein† (%)	Starch† (%)	Fiber† (%)	Fat† (%)
					exsertion (cm)	length (cm)			kernel weight (g)	Df						
Rep (E)	3	510.5**	2 187012.2**	106502.8**	7421.4**	2813.4**	1652954.2**	9.1	2	14496.1**	2	9.82**	0.51	0.33**	17.22**	
Genotypes (G)	4	9.1*	3 352.9	834.5**	1433.3**	70.4**	5584.7*	154.3**	3	70.4**	2	2.51**	2.78	0.01**	0.37**	
G X E	210	11.9**	210 8886.9**	8803.0**	111.6**	51.7**	10298.8**	77.7**	210	115.9**	210	2.03**	1.66**	0.03**	0.71**	
Hybrids (H)	630	4.3**	420 465.5**	446.2**	42.2*	5.4**	2048.8*	9.9**	420	22.2**	419	0.33*	0.40**	0.00**	0.08*	
H X E	159	8.7**	159 8192.8**	8387.6**	68.4**	34.5**	6953.9**	61.3**	159	79.5**	159	1.68**	1.26**	0.02**	0.60**	
H X E	477	4.0**	318 465.5**	446.5**	42.1	4.8	2278.4	8.5*	318	17.8**	317	0.31*	0.32*	0.00*	0.07*	
Lines (L)	39	16.2**	39 19630.0**	20344.5**	144.9**	94.8**	16719.0**	163.3**	39	41.8**	39	5.00**	3.39**	0.03**	1.89**	
Testers (T)	3	69.5**	3 142479.9**	144569.9**	266.5**	350.0**	48360.3**	705.8**	3	3140.1**	3	6.90**	6.56**	0.37**	4.63**	
L X T	117	4.5**	117 929.3**	914.6**	37.6	6.5**	2589.8*	11.3**	117	12.8**	117	0.46**	0.40**	0.01**	0.07	
L X E	117	6.2**	78 1045.5**	1063.0**	40.8	6.3**	3143.8**	10.6*	78	10.2*	78	0.39*	0.52**	0.00*	0.09*	
T X E	9	15.5**	6 2617.3**	2908.5**	195.8**	21.8**	2865.2	28.0**	6	548.8**	6	3.64**	2.13**	0.05**	0.32**	
L X T X E	351	3.0*	234 217.3*	178.9	38.6	3.9	1958.5	7.3	234	6.6	233	0.20	0.20	0.00	0.06	
RSC‡	9	39.0**	9 57951.3**	59150.2**	275.8**	257.1**	32796.4**	556.7**	9	122.1**	9	16.47**	10.30**	0.09**	5.95**	
RSC X E	27	8.5**	18 3132.4**	3105.2**	56.3	10.5*	4845.3*	17.0*	18	23.0**	18	0.59*	1.16**	0.00*	0.18*	
RSC X T	27	10.7**	27 1978.6**	1976.4**	38.4	7.4	3438.8	17.2*	27	17.0*	27	0.98**	0.93**	0.01**	0.11	
RSC X T X E	81	4.2*	54 370.0	273.1	58.4*	5.3	2148.3	9.0	54	9.8	54	0.20	0.32	0.00	0.09	
Selections (S)§	3	38.6**	3 12037.6**	15224.4**	190.3*	29.2*	57132.7**	50.2*	3	13.5	3	0.92	0.62	0.00	1.17**	
S X E	9	7.7*	6 278.2	297.95	38.7	4.1	2657.9	5.4	6	9.3	6	0.70	0.59	0.00	0.32	
S X T	9	2.2	9 227.7	279.7	50.6	4.1	1186.1	9.3	9	17.9	9	0.07	0.28	0.01	0.12	
S X T X E	27	4.6	18 128.2	104.0	34.9	4.4	1654.6	9.9	18	8.0	18	0.25	0.24	0.00	0.05	
Parents (P)	43	4.0*	43 5749.9**	5034.3**	223.0**	52.7**	3822.1**	96.9**	43	122.4**	43	3.17**	2.96**	0.05**	0.86**	
P X E	129	3.0*	86 339.9**	296.7**	41.8	5.6	969.9	12.2*	86	31.6**	86	0.43	0.64*	0.01**	0.11	
Checks (C)	6	33.0**	6 2697.4**	3122.0**	139.0**	66.8**	22289.5**	154.4**	6	765.1**	6	1.44**	0.78*	0.03**	1.73**	
C vs P vs H	2	370.2**	2 147959.7**	140200.6**	1078.2**	1364.6**	377713.7**	746.2**	2	925.2**	2	6.84**	8.38**	0.18**	2.83**	
C vs P vs H X E	6	45.3**	4 4266.9**	4616.6*	88.5	39.2*	10303.3*	32.4	4	36.6	4	0.48	2.59*	0.01	0.59*	
P vs H	1	468.7**	1 227198.7**	172145.4**	1822.5**	363.3**	743383.6**	1397.9**	1	1096.0**	1	10.66**	0.21	0.29**	5.44**	
P vs H X E	3	49.9**	2 4433.4	3143.1	80.2	59.6*	13614.4	15.3	2	55.0	2	0.66	3.47*	0.02	0.78*	
C vs H	1	154.5**	1 116151.8**	156801.4**	111.5	1989.3**	1755.6	256.3**	1	453.8**	1	1.97	15.97**	0.05*	0.06	
Error	987	2.21	737 149.1	129.89	32.4	3.55	1556.5	6.43	744	6.9	418	0.28	0.26	0.00	0.06	

*and** Significant at the 0.05 and 0.01 probability level

†Separate columns for degrees of freedom for particular traits are because of differences in the number of replications or environments from which data were collected for the traits.

‡RSC represents the different lines from the Reinstated Sorghum Conversion (RSC) program and their respective families used in the study.

δ Selections (S) are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2,3}, BC₁F_{2,3}, and the Low F_{2,3}, BC₁F₂.

¶ Content in grain

interaction was significant at $P < 0.01$ for grain yield, total plant height, height to the flag leaf, and number of days to anthesis; 1000-kernel weight and percentage of grain of protein, starch, fiber, and fat in grain were significant at $P < 0.05$. Line x tester interaction was significant at $P < 0.01$ for all traits evaluated except percentage of concentration of fat in grain. Line x environment interaction was significant at $P < 0.01$ for grain yield, total plant height, height to the flag leaf, panicle length, three-panicle weight, and percentage of concentration of starch in grain, while 1000-kernel weight, number of days to anthesis, and percentage of concentration of protein, fiber, and fat in grain showed significance at $P < 0.05$. Tester x environment interaction was significant at $P < 0.01$ for all traits except three-panicle weight that was not significant. Line x tester x environment was significant ($P < 0.05$) only for grain yield and total plant height. RSC x environment was significant at $P < 0.01$ for grain yield, total plant height, height to the flag leaf, number of days to anthesis, and percentage of concentration of starch in grain, while panicle length, three-panicle weight, 1000-kernel weight, and percentage of concentration of protein, fiber, and fat in grain was significant at $P < 0.05$. RSC x tester interaction was significant at $P < 0.05$ for grain yield, total plant height, height to the flag leaf, and percentage of concentration of protein, starch, and fiber in grain. The 1000-kernel weight and number of days to anthesis were significant for RSC x tester at $P < 0.05$. RSC x tester x site interaction was significant at $P < 0.05$ for grain yield and plant exsertion. Grain yield, total plant height, height to the flag leaf, three-panicle weight, and percentage of concentration of fat in grain were significant at $P < 0.01$ for selections, while plant exsertion, panicle length, and 1000-kernel weight were significant at $P < 0.05$. Selection x environment was significant ($P < 0.05$) only for grain yield. There was

no interaction for selection x tester x environment. The sorghum parents were significant ($P < 0.01$) for all traits evaluated except grain yield that was significant at $P < 0.05$.

Parent x environment was significant at $P < 0.01$ for total plant height, height to the flag leaf, and number of days to anthesis, with significance at $P < 0.05$ for grain yield, 1000-kernel weight, and percentage of concentration of starch in grain. Checks and checks versus parents versus hybrids were significant at $P < 0.01$ for all traits except percentage of concentration of fat in grain. Checks versus parents versus hybrid x environment interaction were significant at $P < 0.01$ only for grain; however, total plant height, height to the flag leaf, panicle length, three-panicle weight, and percentage of concentration of starch and fat in grain were significant at $P < 0.05$. Parent versus hybrid was significant ($P < 0.01$) for all traits except percentage of concentration of starch in grain. Parent versus hybrid x environment was significant ($P < 0.01$) for grain yield only with panicle length and with percentage of concentration of starch and fat in grain ($P < 0.05$). The check versus hybrid was significant at $P < 0.01$ for grain yield, total plant height, height to the flag leaf, panicle length, 1000-kernel weight, number of days to anthesis, and percentage of concentration of starch in grain, with fiber concentration significant only at $P < 0.05$. The significant differences in mean square between parents and hybrids for almost all traits indicated they were suitable for studies of combining ability. Significant mean squares of parent versus hybrid indicated good scope for manifestation of heterosis in all the studied traits except percentage of concentration of starch in grain. This provided evidence of the presence of sufficient genetic variability among lines, testers, and hybrids and allows further analysis of GCA and heterosis. The results coincided with

the findings of Jayasudha and Sharma (2009) and Rahimi et al. (2010) who also found significant difference among parents and hybrids.

The significant differences between the interaction of line x tester for the traits indicated SCA attributed in the expression of the traits and provide the importance of dominance or non-additive variance for all traits. Variation among parents, hybrids, parents versus hybrids, lines, testers, and line x tester also was observed in several other sorghum studies (Hovny and El-Dsouky, 2007; Abedel-Mottaleb, 2009; Essa, 2009; Mahdy et al., 2011).

Significant differences between genotypes including parents and checks were found for all traits in three of the four environments (Tables 6-10), while significant differences were found in all traits in the across-environment analysis for genotype (Table 5). No differences were found in NIR for sorghum at Vega in 2016 because of low kernel quantity and analysis of differences only between replications. Combined analysis of the various components of the genotype, hybrid, and line, and the interaction with genotype x environment showed significant differences for all traits, as was the case at Dumas, TX, in 2016. Significant differences between RSC families were recorded for all traits in all four environments (Tables 6-10), as was the case in the across-environment analysis (Table 5).

RSC x tester interaction showed various significant differences at individual locations, whereas in the combined location analysis (Table 5), RSC x tester and RSC * environment showed significant differences for grain yield, total plant height, height to

Table 6. Mean squares of grain yield and agronomic traits of sorghum hybrids at Vega, TX, in 2015, with means of various categories of experimental entries.

Source of variation	Df†	Grain yield Mg ha ⁻¹	Days to anthesis	Total height cm	Flag leaf height cm	Plant exsertion cm	Panicle length cm	3-panicle weight g	1000-kernel weight g	Protein‡ %	Starch‡ %	Fiber‡ %	Fat‡ %
Replications	1	11.0*	132.8**	0.6	2243.0*	3924.4**	210.7**	1.6	458.2**	0.40	0.34	0.00	0.03
Genotypes (G)	210	6.8**	62.0**	3994.6**	3849.1**	100.2**	21.3**	5567.4**	35.5**	1.10**	1.66**	0.02**	0.39**
Hybrids (H)	159	5.6**	49.6**	3746.5**	3753.1**	80.9	13.6**	4893.6**	28.3**	0.94**	1.25**	0.01**	0.34**
Lines (L)	39	12.5**	14.5*	8489.4**	8557.7**	98.3	31.7**	9447.1**	67.7**	2.44**	3.34**	0.02**	0.97**
Testers (T)	3	34.24**	2038.6**	64640.3**	67055.5**	382.3*	116.2**	22744.1**	257.6**	1.54*	4.55**	0.10**	1.29**
L X T	117	2.5**	8.6	591.0**	524.1**	67.4	5.0	2817.9	9.6	0.44*	0.47*	0.00*	0.10**
RSC‡	9	32.2**	36.9**	26595.1**	25913.7**	184.1*	77.6**	16148.3**	206.4**	6.98**	10.01**	0.06**	2.88**
RSC X T	27	3.8*	11.5	1069.6*	921.1	111.1*	5.9	3607.2	16.3*	0.88**	1.09**	0.01*	0.17*
Selections (S)δ	3	11.5*	6.3	6214.5	6787.8*	34.4	5.5	21215.8*	16.2	0.56	0.87	0.00	0.46
S X T	9	0.8	11.6	185.6	214.4	74.2	7.7	1953.2	10.2	0.26	0.42	0.00	0.10
Parents (P)	43	2.62**	57.5**	2555.5**	2138.3**	141.5**	18.0**	1392.8	47.3**	1.65**	2.94**	0.03**	0.53*
Checks (C)	6	14.4**	347.7**	859.1**	1383.4**	130.1	43.0**	9302.8**	46.2**	0.92*	0.59	0.00	0.29**
C vs P vs H	2	171.3**	291.5**	64059.8**	55660.8**	654.1**	637.4**	137683.7**	318.5**	2.57*	10.08**	0.15**	1.98**
P vs H	1	110.4**	162.2*	104009.3**	72285.2**	1149.9**	389.5**	260135.3**	631.1**	2.05	2.42	0.27**	3.73**
C vs H	1	172.8**	321.9*	43878.9**	58420.6**	40.9	667.3**	1044.9	4.4	2.60*	18.85**	0.01	0.42
Error	242/206	1.19	10.19	198.79	178.85	54.19	4.62	2036.95	7.27	0.29	0.31	0.00	0.06
Mean of genotypes		5.50	72.5	162.1	121.1	14.3	26.8	170.9	27.6	8.5	68.22	1.71	2.34
Mean of hybrids		5.52	72.5	173.2	131.4	15.0	26.8	182.2	28.2	8.5	68.14	1.72	2.40
Mean of parents		4.25	74.0	134.3	99.0	10.8	24.4	120.7	25.2	8.7	68.33	1.66	2.16
Mean of checks		7.48	69.8	141.8	95.3	15.8	30.7	170.9	27.9	8.1	69.33	1.76	2.22
Mean of lines		4.08	74.2	135.5	100.4	10.8	24.4	120.0	25.3	8.8	68.25	1.65	2.40
Mean of testers		5.93	72.5	122.6	85.9	10.8	25.9	127.7	23.9	7.8	69.08	1.79	2.40

*and** Significant at the 0.05 and 0.01 probability levels.

†Df separated for NIR quality analysis (protein, starch, fiber, and ash) because only one replication was analyzed: difference = checks plus one entry missing.

‡RSC represents the different lines from the Reinstated Sorghum Conversion (RSC) program and their respective families used in the study.

δ Selections (S) are equal to the High and Low percentage of exotic genome recovery and the generation: High F2:3, BC1F2:3, and the Low F2:3, BC1F2:3.

¶ Content in grain

Table 7. Mean squares of grain yield and agronomic traits of sorghum hybrids at Vega, TX, in 2016, with means of various categories of experimental entries.

Source of variation	Df†	Grain yield Mg ha ⁻¹	Days to anthesis	Total height cm	Flag leaf height cm	Plant exsertion cm	Panicle length cm	3-panicle weight g	1000-kernel weight g	Protein‡ %	Starch‡ %	Fiber¶ %	Fat¶ %
Replications	1/0	3.6	78.5**	1039.3*	186.9	352.1**	0.04	16439.0**	2.9	N/A	N/A	N/A	N/A
Genotypes (G)	210/209	3.7**	70.0**	1582.9**	1483.0**	43.4**	15.5**	3080.8**	26.5**	0.68	0.45	0.01	0.23
Hybrids (H)	159/158	2.7**	55.3**	1442.5**	1406.4**	35.7*	11.2**	2571.6**	20.4**	0.61	0.36	0.01	0.19
Lines (L)	39	4.1**	34.9**	2697.4**	2770.1**	58.9**	24.7**	4805.1**	47.8**	1.43	0.66	0.01	0.46
Testers (T)	3	10.5**	2123.2**	31469.9**	29833.7**	131.8*	120.1**	14148.8**	161.4**	6.23**	3.20**	0.16**	2.24**
L X T	117/116	2.0	8.9*	254.3**	222.9**	25.5	3.9*	1530.3**	7.6*	0.19	0.18	0.00	0.04
RSC‡	9	7.2**	102.2**	6483.5**	6923.4**	73.2*	66.3**	10068.2**	152.9**	4.88**	2.07**	0.02**	1.50**
RSC X T	27	2.9*	15.4*	507.3*	8498.9*	23.9	5.7	1517.8	9.0	0.19	0.31*	0.00	0.06
Selections (S)δ	3	9.1*	13.8	2297.8*	2806.3**	41.8	13.4	8825.3*	36.9*	0.69	0.17	0.00	0.30
S X T	9	4.5*	6.3	66.1	34.7	33.4	1.6	1042.3	6.8	0.07	0.20	0.00	0.08
Parents (P)	43	2.2*	57.5**	1168.2**	936.5**	71.5*	17.7**	1410.4**	35.9**	0.99	0.81	0.02	0.28
Checks (C)	6	11.0**	490.0**	947.7**	825.4**	19.4	15.4**	4392.9**	68.9**	0.37	0.22	0.01	0.66
C vs P vs H	2	95.4**	252.9*	23563.5**	21299.2**	129.3*	305.5**	75530.9**	183.2**	1.12	0.87	0.02	1.50*
P vs H	1	147.7**	300.7*	394.9**	30629.8**	258.6*	66.5*	141053.4**	302.9**	2.10	0.05	0.03	2.60**
C vs H	1	18.9*	123.0	14278.8**	19079.4**	8.6	465.0**	1109.5	118.0*	0.05	1.62	0.01	0.23
Error	248/N/A	1.4	4.68	98.83	72.40	24.50	2.53	659.24	1.28	N/A	N/A	N/A	N/A
Mean of genotypes		5.65	76.4	127.1	98.9	6.0	22.1	104.8	27.7	8.9	68.26	1.77	2.75
Mean of hybrids		5.85	76.2	133.7	105.3	6.4	22.0	112.9	28.3	8.8	68.25	1.77	2.79
Mean of parents		4.39	78.3	109.7	84.2	4.5	21.0	67.7	26.2	9.1	68.21	1.75	2.52
Mean of checks		6.50	74.5	115.8	84.6	6.0	25.2	117.9	26.7	8.7	67.74	1.82	2.98
Mean of lines		4.38	78.5	111.2	85.7	4.6	21.0	67.1	26.2	9.1	68.13	1.73	2.80
Mean of testers		4.54	76.5	94.2	69.4	4.0	20.8	73.4	25.8	8.5	69.02	1.88	2.74

*and** Significant at the 0.05 and 0.01 probability levels.

†Df separated for NIR quality analysis (protein, starch, fiber, and ash) because only one replication was analyzed: difference = checks plus one entry missing.

‡RSC represents the different lines from the Reinstated Sorghum Conversion (RSC) program and their respective families used in the study.

δ Selections (S) are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, and the Low F_{2:3}, BC₁F_{2:3}.

¶ Content in grain

Table 8. Mean squares of grain yield and agronomic traits of sorghum hybrids at Dumas, TX, in 2016, with means of various categories of experimental entries.

Source of variation	Df†	Grain yield Mg ha ⁻¹	Total height cm	Flag leaf height cm	Plant exsertion cm	Panicle length cm	3-panicle weight g	1000-kernel weight g	Protein‡ %	Starch§ %	Fiber¶ %	Fat¶ %
Replications	1	0.01	18.8	73.7	23.3	0.3	313.6	1.8	4.63**	5.22**	0.01*	0.71**
Genotypes (G)	210	5.07**	4242.3**	4365.3**	52.3**	25.9**	5747.3**	35.8**	1.10**	0.63**	0.01**	0.31**
Hybrids (H)	159	5.48**	3939.9**	4124.7**	35.9**	19.4**	4067.7**	29.8**	0.00**	0.50**	0.01**	0.27**
Lines (L)	39	4.76*	10511.9**	11122.1**	69.4**	50.9**	8759.5**	68.9**	2.32**	1.06**	0.01**	0.83**
Testers (T)	3	51.97**	51061.6**	52828.6**	142.4**	160.6**	16945.9**	343.6**	5.37**	3.98**	0.21**	1.31**
L X T	117	4.49*	519.4**	526.2**	21.9	5.3*	2161.9	8.7	0.29*	0.22*	0.00*	0.06
RSC‡	9	10.84**	31059.9**	32448.9**	130.7**	133.8**	16303.7**	231.2**	6.88**	2.48**	0.02**	2.47**
RSC X T	27	9.05**	1131.7*	1095.1	19.6	6.3	2542.5	9.9	0.56*	0.41*	0.01*	0.07
Selections (S)δ	3	3.47	454.1	6197.4*	191.7**	18.5	32279.7**	7.8	0.88	0.85*	0.00	0.62*
S X T	9	5.41	231.5	238.2	12.7	3.6	1508.4	12.1	0.24	0.14	0.00	0.03
Parents (P)	43	3.77	2806.0**	2553.0**	93.6**	28.2**	2958.1*	38.0**	1.74**	0.97*	0.03**	0.33**
Checks (C)	6	4.59	1131.9**	1193.3**	52.8**	23.6**	10371.3*	91.2**	0.71	0.24	0.03**	1.03**
C vs P vs H	2	1.53	68484.0**	71943.6	468.4**	497.3**	185293.7**	308.2**	5.42**	4.55**	0.10**	0.24
P vs H	1	3.00	92276.9**	75464.7**	573.1**	26.1	369303.6**	493.9**	9.30**	4.71*	0.10**	0.21
C vs H	1	0.02	67727.3**	92856.2**	215.5**	887.3**	18583.9*	215.5*	0.81	3.40*	0.03*	0.24
Error	245/212	3.03	150.51	139.32	18.68	3.52	1986.25	7.77	0.27	0.21	0.00	0.07
Mean of genotypes		6.53	162.0	127.2	8.3	26.4	226.8	27.6	8.78	68.29	1.76	2.69
Mean of hybrids		6.57	173.3	138.6	8.6	26.1	243.2	28.4	8.71	68.32	1.76	2.70
Mean of parents		6.36	136.7	105.5	5.7	25.5	170.0	25.7	9.08	68.07	1.73	2.65
Mean of checks		6.55	133.7	92.3	10.9	30.6	222.4	26.1	8.47	68.81	1.81	2.81
Mean of lines		6.33	139.3	107.9	5.9	25.1	166.0	25.6	9.14	67.98	1.71	2.70
Mean of testers		6.73	111.3	82.1	4.5	24.8	209.3	26.2	8.46	68.90	1.86	2.70

*and** Significant at the 0.05 and 0.01 probability levels.

†Df separated for NIR quality analysis (protein, starch, fiber, and ash) because only two replications were analyzed: difference = checks.

‡RSC represents the different lines from the Reinstated Sorghum Conversion (RSC) program and their respective families used in the study.

δ Selections (S) are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2,3}, BC₁F_{2,3}, and the Low F_{2,3}, BC₁F_{2,3}

¶ Content in grain

Table 9. Mean squares of grain yield and agronomic traits of sorghum hybrids at Perryton, TX, in 2015, with means of various categories of experimental entries. Data missing 10 hybrids and no parents planted.

Source of variation	Df†	Total height cm	Flag leaf height cm	Plant exsertion cm	Panicle length cm	3-panicle weight g	1000-kernel weight g	Protein‡ %	Starch‡ %	Fiber¶ %	Fat¶ %
Replications	1	1807.5**	1794.4**	2.04	1.7	59356.6**	22.1*	18.16**	39.20	0.07**	0.25*
Genotypes (G)	156	4569.3**	4992.5**	50.8**	29.1**	7385.9**	26.7**	1.13**	9.74	0.01**	0.34**
Hybrids (H)	149	4165.3**	4485.9**	50.8**	20.6**	6572.9**	25.1**	1.14**	1.21**	0.01**	0.31**
Lines (L)	39	9300.2**	10195.9**	94.3**	52.7**	10101.7**	67.5**	3.14**	3.33**	0.02**	0.94**
Testers (T)	3	67720.7**	71177.0**	284.8**	110.4**	62726.6**	105.3**	0.92*	1.01*	0.02**	1.43**
L X T	107	481.7**	497.0**	27.2	6.3**	3578.5*	7.1*	0.43*	0.43	0.00**	0.06**
RSC‡	9	26914.0**	29133.4**	179.3**	139.7**	16342.8**	219.7**	8.28**	10.35**	0.05**	2.49**
RSC X T	27	1238.9**	1315.1**	34.5	10.5*	4225.4	13.6*	0.59	0.70*	0.01**	1.51**
Selections (S)δ	3	67524.5*	9601.0*	401.2**	2.5	13276.4*	14.4	0.70	0.87	0.00	0.49*
S X T	9	262.3	196.5	20.0	2.4	3252.7	8.3	0.24	0.34	0.00	0.05
Checks (C)	6	1213.4**	1170.7**	54.3*	41.7**	27505.0**	25.5**	1.21*	175.9	0.01*	0.90**
C vs H	1	84908.4**	103417.0**	23.2	1225.3**	7749.5	264.0**	0.05	165.4*	0.01	0.21
Error	191/150	105.89	117.55	25.01	2.53	2207.18	3.99	0.30	19.6	0.00	0.03
Mean of genotypes		177.2	135.5	14.8	26.9	217.9	28.5	8.8	68.28	1.68	2.59
Mean of hybrids		183.9	142.9	14.9	26.1	215.8	28.8	8.5	68.44	1.68	2.59
Mean of checks		141.2	95.7	14.2	31.2	229.0	26.4	8.6	65.01	1.70	2.47

DTF = days to 50% flowering; *and** Significant at the 0.05 and 0.01 probability levels.

†Separate columns for degrees of freedom for particular traits are because of differences in the number of replications or environments from which data were collected for the traits.

‡RSC represents different lines from the Reinstated Sorghum Conversion (RSC) program and their respective families used in the study.

δSelections (S) are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2,3}, BC₁F_{2,3}, and the Low F_{2,3}, BC₁F_{2,3}.

¶ Content in grain

Table 10. Mean squares of grain yield of sorghum hybrids at Taylor, TX, in 2015, and Hutchinson, KS, in 2015 and 2016, with means of various categories of experimental entries. Data on days to flowering (DTF) were collected only at Hutchinson, KS, in 2015.

Source of variation	Df	Hutchinson		Df	Taylor 2015	
		2015	2016		2015	2016
		Grain yield Mg ha ⁻¹	DTF		Grain yield Mg ha ⁻¹	DTF
Replications	1	21.79*	0.01	1	42.19**	1
Genotypes (G)	210	9.12**	28.3**	166	4.24**	.
Hybrids (H)	159	7.00**	10.2	159	3.44*	159
Lines (L)	39	13.44**	13.12*	39	6.71**	39
Testers (T)	3	18.52*	44.76*	3	10.32*	3
L X T	117	4.56	8.30	117	2.18	117
RSC†	9	13.81*	29.61**	9	13.72**	9
RSC X T	27	8.64*	13.13*	27	3.93*	27
Selections (S)‡	3	37.81**	11.96	3	16.32*	3
S X T	9	5.24	15.81	9	0.54	9
Parents (P)	43	4.30	70.6**	.	.	.
Checks (C)	6	22.18**	63.6**	6	9.51*	.
C vs P vs H	2	241.91**	454.0**	.	.	.
P vs H	1	357.59**	743.8**	.	.	.
C vs H	1	58.74*	59.6*	1	99.35**	.
Error	252	3.19	5.92	204	2.18	159
Mean of genotypes		7.864	65.03		6.11	.
Mean of hybrids		8.162	64.54		5.90	.
Mean of parents		5.886	67.83		.	2.50
Mean of checks		9.272	63.43		7.39	.
Mean of lines		5.799	68.41		5.897	.
Mean of testers		6.743	62.00		5.897	.

*Significant at the 0.05 probability level.

**Significant at the 0.01 probability level.

†RSC represents the different lines from the Reinstated Sorghum Conversion (RSC) program and their respective families used in the study.

‡Selections (S) are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, and Low F_{2:3}, BC₁F₂

the flag leaf, and percentage of concentration of starch in grain. The sorghum selections were significant at Vega, TX, in 2015 for grain yield, height to the flag leaf, and three-panicle weight that also were recorded for Vega in 2016, but with the inclusion of total plant height and 1000-kernel weight. Selections showing significant differences at Dumas, TX, in 2016 were height to the flag leaf, three-panicle weight, and concentrations of starch and fat in grain, with significance of only $P < 0.05$ for grain yield at Vega in 2015. In the combined analysis, sorghum selections showed significant differences for all traits except concentrations of protein, starch, and fiber in the grain. Parents showed

significant differences in individual locations with the exception of three-panicle weight at Vega in 2015; NIR data at Vega in 2016 as previously explained; and grain yield at Dumas in 2016. In the combined analysis, parents showed significant differences for all traits. The seven commercial check hybrids were significantly different in the across-environment analysis (Table 5); at Vega in 2015, there were significant differences among the checks for all traits except plant exsertion and starch concentration in grain, which also was found at Vega in 2016 with the exclusion of concentrations of protein, starch, fiber, and fat in grain. Commercial check sorghums at Dumas, TX, showed significant differences for all traits except grain yield and concentrations of protein and starch in grain. All seven commercial checks at Hutchinson, KS, in 2015 showed significant differences for grain yield and number of days to anthesis.

The greatest overall grain yield for genotypes and hybrids, 7.86 and 8.16 Mg ha⁻¹, respectively, were at Hutchinson in 2015, followed by Dumas, Vega (2016), and Vega (2015), with 6.53, 6.57; 5.65, 5.85; and 5.50, 5.52 Mg ha⁻¹, respectively. Mean grain yield of parents was greatest at Dumas in 2016, followed by Hutchinson in 2015, Vega in 2016, and Vega in 2015, with yields of 6.36, 5.89, 4.39, and 4.25 Mg ha⁻¹, respectively. The greatest overall mean yield was at Hutchinson in 2015, with 9.27 Mg ha⁻¹, followed by Vega in 2015, Dumas in 2016, and Vega in 2016, with 7.48, 6.55, and 6.50 Mg ha⁻¹, respectively. The greatest overall mean yield of lines was at Dumas in 2016, followed by Hutchinson in 2015, Vega in 2016, and Vega in 2015, with 6.33, 5.80, 4.38, and 4.08 Mg ha⁻¹, respectively. The testers yielded more than the lines at all four locations, with greatest yield at Hutchinson in 2015, followed by Dumas in 2016, Vega in 2015, and

Vega in 2016, with mean yields of 6.73, 6.73, 5.93, and 4.54 Mg ha⁻¹, respectively. Sorghum at Vega was planted later than usual in 2015 because of increased rainfall during the planting season but still had 144 growing days, which might have affected yield. Sorghum at Hutchinson was planted 10 days earlier in 2015 than in 2016, with only 133-134 and 143 growing days, respectively; this might account for negative GCA for A.Tx3197 and A.338 because they were probably harvested too early. Sorghum at Vega in 2016 and Dumas in 2016 was planted during the target date for the season in the Texas Panhandle, with 159 and 162 growing days, respectively.

General Combining Ability (GCA) Effects

General combining ability (GCA) effects represent the fixable component of genetic variance, and are important for developing superior genotypes. Individual GCA estimates for Perryton in 2015 could not be obtained because of the absence of 10 experimental hybrids in the line x tester analysis, while individual NIR data could not be obtained for Vega in 2016 because of one missing hybrid. GCA estimates were variable for all traits measured at each location (Table 11). Standard errors were greater for lines than testers across all traits evaluated while both standard errors for lines and testers were one or less for all traits except total height, height to the flag leaf, and three-panicle weight, indicating more variability in lines than testers. The range of combined GCA effects for grain yield varied from -0.982 (RSC76-13 F_{2:3} low) to 1.295 (RSC117-4 BC₁F_{2:3} high) among the lines and from -0.359 (A.Tx3197) to 0.359 (A.319) among the testers (Table 12). The magnitude of variance due to specific combining ability (SCA) was greater than that of GCA for grain yield, number of days to anthesis, total plant

Table 11. Combined means of General Combining Ability (GCA) estimates for 12 traits for each parental line in a line x tester evaluated across environments.

Parent	Selections¶	Grain yield Mg ha ⁻¹	Days to anthesis	Total height cm	Height to flag leaf cm	Plant exertion cm	Panicle length cm	3-1000-				Protein‡ %	Starch‡ %	Fiber‡ %	Fat‡ %
								panicle weight g	kernel weight g						
RSCδ															
RSC73-9	F _{2,3} High	0.28	-1.38	1.75	2.38	1.20	-1.90	-22.48	-1.35			-0.32	-0.08	0.00	-0.05
RSC73-6	BC ₁ F _{2,3} High	0.14	-1.50	-7.08	-2.49	-1.83	-2.72	-11.30	-1.62			-0.08	-0.11	0.01	0.10
RSC73-1	F _{2,3} Low	-0.61	0.17	5.48	7.77	0.35	-2.42	-18.96	0.13			-0.42	0.30	-0.02	-0.35
RSC73-5	BC ₁ F _{2,3} Low	-0.12	1.12	36.32	36.75	2.11	-2.64	3.02	1.28			-0.12	0.17	0.03	-0.16
RSC83-1	F _{2,3} High	-0.18	1.62	0.09	4.23	-3.22	-0.93	21.41	0.89			-0.28	0.05	-0.01	-0.14
RSC83-14	BC ₁ F _{2,3} High	1.13	-0.08	1.64	7.66	-4.57	-1.49	36.34	0.41			0.24	-0.62	-0.01	0.10
RSC83-10	F _{2,3} Low	-0.22	1.33	-35.86	-36.33	-1.43	1.82	-25.71	-1.06			-0.03	-0.06	0.03	-0.05
RSC83-1	BC ₁ F _{2,3} Low	1.12	0.04	1.01	4.74	-1.93	-1.82	17.98	0.71			0.25	-0.46	0.00	0.11
RSC112-5	F _{2,3} High	0.79	0.92	-22.30	-25.40	1.66	1.34	4.47	1.40			-0.53	0.68	-0.01	-0.14
RSC112-19	BC ₁ F _{2,3} High	1.05	1.29	-22.34	-20.93	-2.08	0.72	15.59	-0.52			-0.11	0.39	0.01	0.13
RSC112-8	F _{2,3} Low	-0.21	0.71	-38.76	-40.11	0.37	0.90	-17.54	-1.17			-0.08	0.01	0.02	-0.09
RSC112-15	BC ₁ F _{2,3} Low	0.56	0.33	-26.00	-28.66	0.99	1.64	5.70	0.17			-0.21	0.48	0.02	0.05
RSC76-4	F _{2,3} High	-0.98	1.42	58.77	61.28	1.19	-3.70	22.50	6.85			-0.50	0.32	-0.05	-0.54
RSC76-16	BC ₁ F _{2,3} High	-0.92	2.37	63.82	62.88	3.34	-2.49	55.21	7.36			-0.32	0.40	0.00	-0.32
RSC76-13	F _{2,3} Low	-0.98	0.33	44.27	45.63	1.38	-2.64	-11.02	4.72			-0.23	0.14	-0.02	-0.28
RSC76-2	BC ₁ F _{2,3} Low	-0.72	2.75	64.64	64.89	1.41	-1.58	90.48	6.54			-0.27	0.20	-0.02	-0.24
RSC38-5	F _{2,3} High	-0.73	-1.08	-26.29	-27.11	-2.15	2.98	-20.89	-3.71			-0.01	-0.21	-0.05	-0.12
RSC38-15	BC ₁ F _{2,3} High	-0.48	-1.33	32.56	37.36	-2.52	-2.07	1.76	-0.45			0.54	-0.21	-0.04	0.32
RSC38-8	F _{2,3} Low	-0.40	-0.96	15.41	15.70	1.81	-2.19	18.21	-0.04			-0.49	0.31	-0.06	-0.35
RSC38-9	BC ₁ F _{2,3} Low	-0.60	-2.58	3.26	2.14	-1.32	2.38	-13.24	-1.19			0.31	-0.05	0.02	0.14
RSC37-12	F _{2,3} High	0.27	-1.75	-4.27	-5.03	1.83	-1.17	-16.01	-2.10			-0.29	-0.11	0.02	-0.08
RSC37-12	BC ₁ F _{2,3} High	0.48	-0.58	15.52	17.26	-0.87	-0.97	38.55	-0.73			0.15	-0.17	0.04	0.09
RSC37-7	F _{2,3} Low	-0.50	-0.63	9.10	8.07	2.46	-1.52	10.96	-0.91			-0.36	-0.01	0.00	-0.13
RSC37-8	BC ₁ F _{2,3} Low	0.53	-1.50	3.44	4.56	-1.02	-0.19	24.27	-3.08			0.13	0.00	0.03	0.15
RSC15-13	F _{2,3} High	-0.38	-2.50	-1.07	-0.97	0.81	-1.00	-39.35	1.28			0.72	-0.38	-0.02	0.24

RSC15-15	BC ₁ F _{2:3} High	-0.13	-1.17	-26.97	-30.02	-0.16	3.46	-6.70	-1.64	0.39	0.13	-0.01	0.14
RSC15-11	F _{2:3} Low	-0.46	-1.25	25.09	22.81	2.83	-0.63	-35.54	-1.22	0.60	-0.46	0.00	0.36
RSC15-14	BC ₁ F _{2:3} Low	-0.37	-2.04	6.54	6.97	0.06	-0.59	-23.83	1.48	0.75	0.07	0.00	0.28
RSC124-9	F _{2:3} High	0.70	0.37	-36.98	-37.18	-3.08	3.32	-18.51	-2.62	-0.21	-0.17	-0.04	0.00
RSC124-16	BC ₁ F _{2:3} High	0.49	1.71	3.20	1.46	-1.28	2.93	36.18	-1.97	-0.04	0.16	0.01	0.11
RSC124-3	F _{2:3} Low	-0.63	-0.33	-4.58	-11.88	4.67	2.55	-39.35	0.48	0.15	-0.24	-0.03	-0.11
RSC124-4	BC ₁ F _{2:3} Low	0.33	1.96	-14.87	-13.99	-1.26	0.37	-21.01	-1.82	0.03	0.16	0.01	0.04
RSC117-2	F _{2:3} High	0.45	-0.17	-33.28	-32.01	-2.97	1.89	-12.33	-0.57	-0.15	0.03	0.01	-0.02
RSC117-4	BC ₁ F _{2:3} High	1.30	0.25	-39.82	-38.85	-4.96	4.25	9.76	3.29	0.24	0.24	0.03	0.24
RSC117-10	F _{2:3} Low	-0.07	-0.25	-47.00	-48.61	-0.13	1.64	-8.05	-1.44	-0.08	-0.05	0.00	-0.08
RSC117-3	BC ₁ F _{2:3} Low	0.48	1.46	-29.20	-28.73	-2.26	1.97	16.37	1.14	0.20	-0.01	0.01	0.21
RSC19-3	F _{2:3} High	0.05	-0.58	1.97	-6.87	6.40	2.34	-16.29	-2.65	-0.06	-0.29	0.03	0.02
RSC19-17	BC ₁ F _{2:3} High	-0.37	0.46	39.41	38.48	0.76	0.08	-11.11	-1.65	0.64	-0.41	0.01	0.42
RSC19-1	F _{2:3} Low	-0.48	0.37	-46.66	-50.32	3.13	0.50	-41.36	-4.14	-0.02	-0.25	0.03	0.10
RSC19-10	BC ₁ F _{2:3} Low	0.40	0.71	21.78	23.98	-0.49	-1.47	-3.46	-1.51	-0.12	0.09	0.00	-0.05
Testers†													
A.301	Tester	0.32	-3.93	-33.95	-32.96	-0.93	-0.09	-15.14	0.40	0.19	-0.05	0.01	-0.13
A.319	Tester	0.36	0.66	-3.96	-5.74	0.16	1.68	3.36	-2.35	0.09	-0.04	0.01	0.00
A.Tx.3197	Tester	-0.36	-1.40	9.73	8.50	1.67	-0.44	-12.80	0.85	-0.13	0.08	-0.03	0.01
A.338	Tester	-0.32	4.67	27.75	29.79	-0.90	-1.16	24.38	1.10	-0.14	0.00	0.00	0.12
SE Lines		0.269	0.933	3.747	3.680	1.091	0.415	8.869	0.524	0.166	0.160	0.017	0.091
SE Testers		0.085	0.295	1.185	1.164	0.345	0.131	2.805	0.166	0.053	0.051	0.005	0.029

†Testers were analyzed separately

‡Content in grain

δRSC represents different lines from the Reinstated Sorghum Conversion (RSC) program and their respective families used in the study.

¶(Selections (S) are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, and the Low F_{2:3}, BC₁F_{2:3}.

Table 12. General Combining Ability (GCA) estimates and the combined analysis for grain yield (Mg ha⁻¹) for each parental line in a line x tester evaluated in six environments. Because of the absence of parental lines or hybrids, individual GCA estimates were not reported for Taylor, TX, in 2015, or Hutchinson, KS, in 2016. All six locations are included in the combined analysis.

Parents		Vega	Hutchinson	Vega	Dumas	Combined	
RSC†Lines	Selectionsδ	2015	2015	2016	2016	locations	Rank
RSC73-9	F _{2:3} High	0.019	1.542	-0.616	0.074	0.276	15
RSC73-6	BC ₁ F _{2:3} High	1.119	1.764	-0.782	-1.019	0.141	17
RSC73-1	F _{2:3} Low	-0.96	-1.797	-0.866	0.591	-0.613	34
RSC73-5	BC ₁ F _{2:3} Low	-1.341	-0.28	-0.116	0.396	-0.124	20
RSC83-1	F _{2:3} High	-0.998	-1.58	0.789	0.082	-0.177	22
RSC83-14	BC ₁ F _{2:3} High	2.074	0.898	0.348	1.581	1.130	2
RSC83-10	F _{2:3} Low	0.273	-1.307	-0.861	0.682	-0.223	24
RSC83-1	BC ₁ F _{2:3} Low	1.024	1.856	0.276	1.782	1.118	3
RSC112-5	F _{2:3} High	2.147	0.034	1.114	-0.620	0.787	5
RSC112-19	BC ₁ F _{2:3} High	2.191	2.034	2.028	0.490	1.048	4
RSC112-8	F _{2:3} Low	0.819	0.156	0.004	-0.370	-0.207	23
RSC112-15	BC ₁ F _{2:3} Low	1.970	0.653	0.756	0.113	0.557	7
RSC76-4	F _{2:3} High	-1.447	-1.650	-0.184	-0.826	-0.978	39
RSC76-16	BC ₁ F _{2:3} High	-2.302	-2.442	0.335	-0.157	-0.915	38
RSC76-13	F _{2:3} Low	-1.818	-1.120	-0.225	-1.260	-0.982	40
RSC76-2	BC ₁ F _{2:3} Low	-1.947	-0.438	-0.031	-0.861	-0.720	36
RSC38-5	F _{2:3} High	0.413	-1.385	-0.908	-0.586	-0.731	37
RSC38-15	BC ₁ F _{2:3} High	-1.870	0.066	0.225	-1.271	-0.478	30
RSC38-8	F _{2:3} Low	-1.217	-0.203	0.011	-0.529	-0.396	28
RSC38-9	BC ₁ F _{2:3} Low	-0.602	-0.354	0.085	-1.006	-0.598	33
RSC37-12	F _{2:3} High	0.271	-0.068	1.014	-1.151	0.272	16
RSC37-12	BC ₁ F _{2:3} High	-0.748	1.821	-0.242	0.347	0.477	11
RSC37-7	F _{2:3} Low	-0.951	-0.872	-0.476	-0.987	-0.500	32
RSC37-8	BC ₁ F _{2:3} Low	0.334	0.582	0.851	-0.219	0.534	8
RSC15-13	F _{2:3} High	-0.396	0.069	-1.067	0.611	-0.384	27
RSC15-15	BC ₁ F _{2:3} High	1.550	-1.531	-0.483	0.240	-0.131	21
RSC15-11	F _{2:3} Low	-1.884	-0.714	-0.272	0.514	-0.463	29
RSC15-14	BC ₁ F _{2:3} Low	-0.736	0.200	-0.877	-0.459	-0.369	26
RSC124-9	F _{2:3} High	0.996	2.595	-0.045	0.490	0.703	6
RSC124-16	BC ₁ F _{2:3} High	0.383	-0.781	0.958	0.533	0.485	9
RSC124-3	F _{2:3} Low	-0.088	-0.924	-0.936	-0.615	-0.628	35
RSC124-4	BC ₁ F _{2:3} Low	0.325	1.486	-1.122	1.090	0.331	14
RSC117-2	F _{2:3} High	0.606	0.756	0.110	0.140	0.451	12
RSC117-4	BC ₁ F _{2:3} High	2.363	1.567	0.952	0.714	1.295	1
RSC117-10	F _{2:3} Low	0.985	-0.831	-0.481	-0.231	-0.070	19
RSC117-3	BC ₁ F _{2:3} Low	0.455	0.727	-0.019	1.085	0.481	10
RSC19-3	F _{2:3} High	0.752	0.080	0.569	-0.481	0.054	18
RSC19-17	BC ₁ F _{2:3} High	-0.846	-0.904	0.048	-0.019	-0.367	25
RSC19-1	F _{2:3} Low	0.178	-1.932	-0.579	1.018	-0.480	31
RSC19-10	BC ₁ F _{2:3} Low	-0.806	2.226	0.718	0.156	0.396	13
Testers							
A.301	Tester	0.732	0.438	-0.239	0.935	0.321	2
A.319	Tester	0.378	0.255	0.446	0.271	0.359	1
A.Tx.3197	Tester	-0.635	-0.66	-0.344	-0.193	-0.359	4
A.338	Tester	-0.477	-0.033	0.137	-1.002	-0.322	3
SE Lines		0.376	0.652	0.417	0.606	0.269	
SE Testers		0.119	0.206	0.132	0.192	0.085	

†Testers were analyzed separately from the lines.

‡RSC represents different lines from the Reinstated Sorghum Conversion (RSC) program and their respective families used in the study.

δSelections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, and the Low F_{2:3}, BC₁F_{2:3}.

height, height to the flag leaf, panicle length, three-panicle weight, and 1000-kernel weight, revealing that non-additive gene action was dominant in the inheritance of grain yield and all its component traits. Rani et al. (2015) studied six lines and eight testers of grain sorghum and found similar results for grain yield and its components. Similar observations also were reported by Chaudhary et al. (2006) and Premalatha et al. (2006). Non-additive gene action for grain yield and its components in the present study was in accordance with findings by Aruna et al. (2010), Mahdy et al (2011), and Rani et al. (2015), whereas additive gene action controlling the inheritance of grain yield per plant was reported by Prabhakar et al. (2013). Line RSC117-4, a BC generation with a large percentage of exotic genome recovery ($BC_1F_{2:3}$ high) had the greatest GCA (1.295) and proved to be a good combiner for grain yield. Tester A.319 had the greatest GCA (0.359), but was followed closely by A.301 with a GCA effect of 0.321 for grain yield. This grain yield was very encouraging because plant breeders continually strive for earlier, better-yielding varieties. A.301 was generally a good combiner for the number of days to anthesis, total plant height, height to the flag leaf, and concentration of protein and fiber in grain, with GCA effects of -5.252, -33.947, -32.964, 0.189, and 0.013, respectively.

RSC76-2 $BC_1F_{2:3}$ low had the greatest GCA effect of 3.548 for the number of days to anthesis; however, if sorghum breeders are seeking earlier hybrids, line RSC37-12 $BC_1F_{2:3}$ low with a GCA effect of -3.639 would be a good combiner to shorten the duration of vegetative growth. Tester A.301 had a combined GCA effect of -5.252 and proved to be a good combiner for early maturity. For total plant height and height to the

flag leaf, RSC76-2 ranked first with GCA effects of 64.638 and 64.890, respectively; however, RSC117-10, a $F_{2:3}$ generation with small percentage of exotic genome recovery ($F_{2:3}$ low) ranked 40th and 39th with GCA effects of -47.003 and -48.605, respectively. RSC117-10 is a good general combiner to reduce plant height with the greatest panicle length GCA effect (Tables 11-16). Negative GCA effect for plant height and height to the flag leaf is useful for development of dwarf plant material (Fellahi et al., 2013).

Tester A.338 had the least GCA effect for panicle length but the greatest GCA for three-panicle and 1000-kernel weight, with -1.158, 24.383, and 1.104, respectively (Tables 16, 18-19). RSC family 76 ranked 1st and 2nd for three-panicle weight, with GCA effects of 90.480 and 55.208, respectively; in addition, all four family members ranked 1-4 for 1000-kernel weight, with GCA effects of 7.360, 6.849, 6.536, and 4.720 for BC_1 and $F_{2:3}$ generation with large percentage of exotic genome recovery and BC_1 and $F_{2:3}$ generation with small percentage of exotic genome recovery, respectively. Others with large GCA effect for 1000-kernel weight were RSC117-4, RSC15-14, RSC112-5, and RSC73-5 that consisted of $BC_1F_{2:3}$ high, $BC_1F_{2:3}$ low, $F_{2:3}$ high, and $BC_1F_{2:3}$ low, respectively (Table 19). The greatest GCA effect for environmentally affected exertion was in tester B.Tx3197 while the lines with the greatest GCA effects were RSC19-3 ($F_{2:3}$ high), RSC124-3 ($F_{2:3}$ low), RSC76-6 ($BC_1F_{2:3}$ high), RSC19-1 ($F_{2:3}$ low), and RSC15-11 ($F_{2:3}$ low) with GCA effects of 6.403, 4.666, 3.34, 3.131, and 2.825, respectively (Table 17).

In sorghum, the plant height, number of days to anthesis, maturity, plant exertion, panicle size, and kernel size in addition to grain yield are major selection criteria. A.301 had the greatest combined GCA effect (0.189) for concentration of protein in the grain that was very influenced by environment (Table 19). The greatest GCA effect (0.335)

Table 13. General Combining Ability (GCA) estimates and combined analysis for the number of days to anthesis for each parental line in a line x tester evaluated in two environments.

Parents		Vega		Combined	Rank
RSC†Lines	Selectionsδ	2015	2016	locations	
RSC73-9	F _{2:3} High	-0.468	-2.572	-1.827	33
RSC73-6	BC ₁ F _{2:3} High	-0.343	-3.697	-2.389	36
RSC73-1	F _{2:3} Low	0.657	-0.072	0.611	16
RSC73-5	BC ₁ F _{2:3} Low	1.782	-0.197	0.423	18
RSC83-1	F _{2:3} High	3.157	0.803	2.486	4
RSC83-14	BC ₁ F _{2:3} High	0.532	0.303	0.923	14
RSC83-10	F _{2:3} Low	0.532	1.428	1.173	10
RSC83-1	BC ₁ F _{2:3} Low	0.282	-0.447	0.548	17
RSC112-5	F _{2:3} High	1.907	1.428	1.611	8
RSC112-19	BC ₁ F _{2:3} High	1.675	3.178	2.861	3
RSC112-8	F _{2:3} Low	0.782	0.928	1.298	9
RSC112-15	BC ₁ F _{2:3} Low	1.782	0.553	0.798	15
RSC76-4	F _{2:3} High	0.782	2.428	2.173	5
RSC76-16	BC ₁ F _{2:3} High	1.782	4.428	3.423	2
RSC76-13	F _{2:3} Low	0.282	0.428	-0.202	23
RSC76-2	BC ₁ F _{2:3} Low	1.782	4.678	3.548	1
RSC38-5	F _{2:3} High	-0.343	-1.322	-1.014	29
RSC38-15	BC ₁ F _{2:3} High	-1.218	-0.947	-1.827	34
RSC38-8	F _{2:3} Low	-2.093	-1.322	-2.014	35
RSC38-9	BC ₁ F _{2:3} Low	-3.093	-2.947	-3.514	39
RSC37-12	F _{2:3} High	-3.093	-2.072	-3.639	40
RSC37-12	BC ₁ F _{2:3} High	-0.218	-0.572	-0.452	24
RSC37-7	F _{2:3} Low	-0.843	-1.072	-0.764	27
RSC37-8	BC ₁ F _{2:3} Low	-0.843	-2.322	-1.139	30
RSC15-13	F _{2:3} High	-1.968	-4.197	-3.327	37
RSC15-15	BC ₁ F _{2:3} High	-0.343	-1.697	-1.139	31
RSC15-11	F _{2:3} Low	-1.611	-0.697	-0.577	26
RSC15-14	BC ₁ F _{2:3} Low	-2.754	-3.697	-3.327	38
RSC124-9	F _{2:3} High	-1.093	0.678	-0.514	25
RSC124-16	BC ₁ F _{2:3} High	1.157	3.178	2.111	6
RSC124-3	F _{2:3} Low	-0.343	-1.072	-0.764	28
RSC124-4	BC ₁ F _{2:3} Low	-0.468	2.553	0.986	13
RSC117-2	F _{2:3} High	-0.843	-0.822	-1.139	32
RSC117-4	BC ₁ F _{2:3} High	0.907	-1.072	0.111	21
RSC117-10	F _{2:3} Low	0.032	0.178	0.298	19
RSC117-3	BC ₁ F _{2:3} Low	1.032	1.303	1.173	11
RSC19-3	F _{2:3} High	-0.593	0.428	-0.139	22
RSC19-17	BC ₁ F _{2:3} High	0.407	1.803	1.736	7
RSC19-1	F _{2:3} Low	0.282	0.928	0.298	20
RSC19-10	BC ₁ F _{2:3} Low	0.657	1.178	1.111	12
Testers					
A.301	Tester	-5.557	-5.297	-5.252	4
A.319	Tester	0.734	0.566	1.023	2
A.Tx.3197	Tester	-1.831	-2.109	-2.358	3
A.338	Tester	6.762	6.841	6.586	1
SE Lines		1.026	0.816	0.612	
SE Testers		0.324	0.258	0.194	

†Testers were analyzed separately from the lines

‡RSC represents different lines from the Reinstated Sorghum Conversion (RSC) program.

δSelections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, and the Low F_{2:3}, BC₁F_{2:3}.

Table 14. General Combining Ability (GCA) estimates and combined analysis for total plant height (cm) for each parental line in a line x tester evaluated in four environments. Because of the absence of 10 hybrids, individual GCA was not estimated for Perryton, TX, in 2015.

Parents		Vega		Dumas	Combined	Rank
RSC [‡] Lines	Selections [§]	2015	2016	2016	locations	
RSC73-9	F _{2:3} High	6.965	-4.23	4.583	1.747	19
RSC73-6	BC ₁ F _{2:3} High	3.472	-10.686	-9.916	-7.076	26
RSC73-1	F _{2:3} Low	-0.179	2.861	6.594	5.479	14
RSC73-5	BC ₁ F _{2:3} Low	35.752	27.52	42.789	36.315	6
RSC83-1	F _{2:3} High	6.542	0.427	-6.741	0.093	22
RSC83-14	BC ₁ F _{2:3} High	-1.449	3.813	-3.037	1.641	20
RSC83-10	F _{2:3} Low	-40.343	-21.375	-44.100	-35.863	35
RSC83-1	BC ₁ F _{2:3} Low	3.578	-0.843	-1.767	1.006	21
RSC112-5	F _{2:3} High	-39.178	2.967	-10.445	-22.304	28
RSC112-19	BC ₁ F _{2:3} High	-21.429	-13.649	-24.672	-22.335	29
RSC112-8	F _{2:3} Low	-42.248	-27.196	-39.443	-38.761	37
RSC112-15	BC ₁ F _{2:3} Low	-29.601	-12.095	-30.977	-26.002	30
RSC76-4	F _{2:3} High	58.982	31.436	73.375	58.766	3
RSC76-16	BC ₁ F _{2:3} High	66.284	39.585	75.809	63.818	2
RSC76-13	F _{2:3} Low	40.567	22.652	57.712	44.266	4
RSC76-2	BC ₁ F _{2:3} Low	68.983	37.151	81.207	64.638	1
RSC38-5	F _{2:3} High	-18.858	-18.200	-26.32	-26.286	31
RSC38-15	BC ₁ F _{2:3} High	44.112	11.963	43.742	32.558	7
RSC38-8	F _{2:3} Low	17.019	6.353	20.035	15.413	11
RSC38-9	BC ₁ F _{2:3} Low	12.045	-6.452	-0.497	3.255	16
RSC37-12	F _{2:3} High	-2.348	-5.923	-6.106	-4.272	24
RSC37-12	BC ₁ F _{2:3} High	19.877	12.068	14.532	15.519	10
RSC37-7	F _{2:3} Low	13.103	2.120	8.605	9.103	12
RSC37-8	BC ₁ F _{2:3} Low	5.007	4.237	1.514	3.440	15
RSC15-13	F _{2:3} High	0.879	-0.526	13.368	-1.068	23
RSC15-15	BC ₁ F _{2:3} High	-26.108	-11.638	-37.009	-26.974	32
RSC15-11	F _{2:3} Low	27.98	18.842	35.91	25.093	8
RSC15-14	BC ₁ F _{2:3} Low	21.449	-0.843	11.357	6.537	13
RSC124-9	F _{2:3} High	-37.009	-24.973	-40.819	-36.983	36
RSC124-16	BC ₁ F _{2:3} High	-1.661	15.349	0.985	3.197	17
RSC124-3	F _{2:3} Low	-1.819	-6.241	-10.657	-4.577	25
RSC124-4	BC ₁ F _{2:3} Low	-16.288	-7.828	-24.732	-14.870	27
RSC117-2	F _{2:3} High	-44.999	-21.057	-40.925	-33.284	34
RSC117-4	BC ₁ F _{2:3} High	-42.512	-23.703	-48.333	-39.819	38
RSC117-10	F _{2:3} Low	-55.477	-25.714	-54.048	-47.003	40
RSC117-3	BC ₁ F _{2:3} Low	-43.835	-17.565	-37.538	-29.195	33
RSC19-3	F _{2:3} High	7.071	7.729	-5.259	1.972	18
RSC19-17	BC ₁ F _{2:3} High	32.841	29.531	46.282	39.411	5
RSC19-1	F _{2:3} Low	-45.74	-30.371	-58.387	-46.658	39
RSC19-10	BC ₁ F _{2:3} Low	20.035	14.503	20.247	21.776	9
Testers						
A.301	Tester	-36.046	-25.061	-33.033	-33.947	4
A.319	Tester	-6.633	0.257	-5.026	-3.960	3
A.Tx.3197	Tester	11.839	1.348	11.283	9.725	2
A.338	Tester	31.084	23.456	26.364	27.752	1
SE Lines		5.396	3.695	4.6997	3.747	
SE Testers		1.706	1.169	1.4862	1.185	

[†]Testers were analyzed separately from the lines.

[‡]RSC represents different lines from the Reinstated Sorghum Conversion (RSC) program.

[§]Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, and the Low F_{2:3}, BC₁F_{2:3}.

Table 15. General Combining Ability (GCA) estimates and combined analysis for height to the flag leaf (cm) for each parental line in a line x tester evaluated in four environments. Because of the absence of 10 hybrids, individual GCA was not estimated for Perryton, TX, in 2015.

Parents		Vega		Dumas	Combined	Rank
RSC‡Lines	Selectionsδ	2015	2016	2016	locations	
RSC73-9	F _{2:3} High	8.761	-5.787	2.863	2.380	19
RSC73-6	BC ₁ F _{2:3} High	5.639	-4.835	-6.027	-2.486	23
RSC73-1	F _{2:3} Low	1.935	6.701	7.838	7.765	13
RSC73-5	BC ₁ F _{2:3} Low	35.908	24.164	43.398	36.751	7
RSC83-1	F _{2:3} High	11.143	2.362	-3.169	4.233	18
RSC83-14	BC ₁ F _{2:3} High	8.708	5.008	2.652	7.659	14
RSC83-10	F _{2:3} Low	-37.117	-22.403	-45.608	-36.328	35
RSC83-1	BC ₁ F _{2:3} Low	9.396	0.88	1.911	4.736	16
RSC112-5	F _{2:3} High	-39.552	-2.295	-14.705	-25.40	29
RSC112-19	BC ₁ F _{2:3} High	-20.139	-11.291	-24.940	-20.925	28
RSC112-8	F _{2:3} Low	-41.827	-26.636	-39.682	-40.112	38
RSC112-15	BC ₁ F _{2:3} Low	-30.027	-14.858	-33.967	-28.66	31
RSC76-4	F _{2:3} High	61.096	36.123	78.111	61.277	3
RSC76-16	BC ₁ F _{2:3} High	58.662	39.298	78.111	62.878	2
RSC76-13	F _{2:3} Low	45.168	22.364	58.638	45.626	4
RSC76-2	BC ₁ F _{2:3} Low	62.578	38.557	82.873	64.89	1
RSC38-5	F _{2:3} High	-19.761	-18.699	-27.617	-27.107	30
RSC38-15	BC ₁ F _{2:3} High	49.243	16.332	50.171	37.359	6
RSC38-8	F _{2:3} Low	15.217	8.818	19.373	15.702	11
RSC38-9	BC ₁ F _{2:3} Low	10.613	-4.623	-1.264	2.142	20
RSC37-12	F _{2:3} High	-0.076	-7.692	-8.143	-5.028	24
RSC37-12	BC ₁ F _{2:3} High	21.250	14.533	14.505	17.264	10
RSC37-7	F _{2:3} Low	12.889	3.314	4.768	8.068	12
RSC37-8	BC ₁ F _{2:3} Low	7.756	3.738	0.535	4.563	17
RSC15-13	F _{2:3} High	-1.928	-0.496	14.717	-0.971	22
RSC15-15	BC ₁ F _{2:3} High	-30.662	-14.042	-37.036	-30.017	33
RSC15-11	F _{2:3} Low	23.525	17.496	33.343	22.807	9
RSC15-14	BC ₁ F _{2:3} Low	21.892	0.034	11.33	6.972	15
RSC124-9	F _{2:3} High	-38.599	-24.52	-43.068	-37.179	36
RSC124-16	BC ₁ F _{2:3} High	-6.637	11.146	0.641	1.458	21
RSC124-3	F _{2:3} Low	-5.367	-10.656	-18.621	-11.880	26
RSC124-4	BC ₁ F _{2:3} Low	-12.035	-8.962	-25.288	-13.994	27
RSC117-2	F _{2:3} High	-39.869	-20.075	-41.057	-32.014	34
RSC117-4	BC ₁ F _{2:3} High	-42.483	-24.414	-44.55	-38.846	37
RSC117-10	F _{2:3} Low	-58.496	-26.425	-54.075	-48.605	39
RSC117-3	BC ₁ F _{2:3} Low	-46.325	-15.418	-34.919	-28.734	32
RSC19-3	F _{2:3} High	-1.399	-2.824	-13.541	-6.866	25
RSC19-17	BC ₁ F _{2:3} High	34.055	27.233	44.35	38.483	5
RSC19-1	F _{2:3} Low	-51.299	-30.129	-61.907	-50.323	40
RSC19-10	BC ₁ F _{2:3} Low	19.821	18.978	25.935	23.984	8
Testers						
A.301	Tester	-34.482	-23.808	-32.674	-32.964	4
A.319	Tester	-9.527	-0.887	-5.794	-5.740	3
A.Tx.3197	Tester	10.341	1.251	9.669	8.496	2
A.338	Tester	33.967	23.444	28.39	29.792	1
SE Lines		5.110	3.176	4.554	3.678	
SE Testers		1.616	1.004	1.440	1.164	

†Testers were analyzed separately from the lines.

‡RSC represents different lines from the Reinstated Sorghum Conversion (RSC) program.

δSelections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, and the Low F_{2:3}, BC₁F_{2:3}.

Table 16. General Combining Ability (GCA) estimates and the combined analysis for panicle length (cm) for each parental line in a line x tester evaluated in four environments. Because of the absence of 10 hybrids, individual GCA was not estimated for Perryton, TX, in 2015.

Parents		Vega		Dumas	Combined	Rank
RSC†Lines	Selectionsδ	2015	2016	2016	locations	
RSC73-9	F _{2:3} High	-2.427	-1.771	-1.193	-1.897	32
RSC73-6	BC ₁ F _{2:3} High	-2.531	-2.188	-2.901	-2.715	39
RSC73-1	F _{2:3} Low	-1.781	-2.583	-2.693	-2.418	35
RSC73-5	BC ₁ F _{2:3} Low	-2.136	-2.271	-3.235	-2.637	38
RSC83-1	F _{2:3} High	-0.011	0.062	-2.318	-0.934	23
RSC83-14	BC ₁ F _{2:3} High	-2.031	-0.021	-1.86	-1.485	28
RSC83-10	F _{2:3} Low	0.031	1.625	2.099	1.817	11
RSC83-1	BC ₁ F _{2:3} Low	-2.573	-1.75	-3.068	-1.819	31
RSC112-5	F _{2:3} High	-0.136	2.375	3.224	1.342	14
RSC112-19	BC ₁ F _{2:3} High	-0.088	0.271	1.504	0.721	16
RSC112-8	F _{2:3} Low	1.344	-1.125	1.765	0.900	15
RSC112-15	BC ₁ F _{2:3} Low	1.135	1.066	2.140	1.641	13
RSC76-4	F _{2:3} High	-3.448	-3.250	-3.526	-3.704	40
RSC76-16	BC ₁ F _{2:3} High	-1.915	-1.833	-3.735	-2.490	36
RSC76-13	F _{2:3} Low	-1.823	-2.00	-2.693	-2.636	37
RSC76-2	BC ₁ F _{2:3} Low	-0.865	-1.208	-2.151	-1.579	30
RSC38-5	F _{2:3} High	1.802	2.479	4.265	2.978	4
RSC38-15	BC ₁ F _{2:3} High	-1.552	-1.646	-2.610	-2.074	33
RSC38-8	F _{2:3} Low	-1.823	-1.083	-2.443	-2.189	34
RSC38-9	BC ₁ F _{2:3} Low	1.260	1.729	3.932	2.375	7
RSC37-12	F _{2:3} High	-0.823	-1.333	-0.776	-1.168	26
RSC37-12	BC ₁ F _{2:3} High	-1.740	-0.833	-0.610	-0.965	24
RSC37-7	F _{2:3} Low	-1.156	-1.021	-1.651	-1.516	29
RSC37-8	BC ₁ F _{2:3} Low	0.989	0.021	-1.526	-0.194	20
RSC15-13	F _{2:3} High	-1.531	-0.938	-0.818	-0.996	25
RSC15-15	BC ₁ F _{2:3} High	3.614	2.271	2.849	3.462	2
RSC15-11	F _{2:3} Low	1.079	-0.833	-1.151	-0.629	22
RSC15-14	BC ₁ F _{2:3} Low	-0.564	-1.271	-0.485	-0.585	21
RSC124-9	F _{2:3} High	4.427	2.50	4.015	3.319	3
RSC124-16	BC ₁ F _{2:3} High	3.802	2.50	2.307	2.926	5
RSC124-3	F _{2:3} Low	2.031	1.25	3.515	2.547	6
RSC124-4	BC ₁ F _{2:3} Low	-0.112	0.583	1.557	0.367	18
RSC117-2	F _{2:3} High	1.427	2.354	1.974	1.894	10
RSC117-4	BC ₁ F _{2:3} High	3.552	3.583	3.974	4.249	1
RSC117-10	F _{2:3} Low	0.698	1.75	1.265	1.644	12
RSC117-3	BC ₁ F _{2:3} Low	0.906	1.521	2.224	1.973	9
RSC19-3	F _{2:3} High	3.635	1.646	2.265	2.343	8
RSC19-17	BC ₁ F _{2:3} High	0.073	0.708	-1.276	0.077	19
RSC19-1	F _{2:3} Low	0.156	-0.208	0.599	0.499	17
RSC19-10	BC ₁ F _{2:3} Low	-1.156	-1.125	-2.568	-1.469	27
Testers						
A.301	Tester	-0.366	0.204	0.118	-0.093	3
A.319	Tester	1.795	1.3	1.824	1.684	1
A.Tx.3197	Tester	-0.848	0.154	-0.343	-0.435	2
A.338	Tester	-0.578	-1.658	-1.597	-1.158	4
SE Lines		0.797	0.546	0.704	0.415	
SE Testers		0.252	0.173	0.223	0.131	

†Testers were analyzed separately from the lines.

‡RSC represents different lines from the Reinstated Sorghum Conversion (RSC) program.

δSelections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, and the Low F_{2:3}, BC₁F_{2:3}.

Table 17. General Combining Ability (GCA) estimates and the combined analysis for plant exsertion (cm) for each parental line in a line x tester evaluated in four environments. Because of the absence of 10 hybrids, individual GCA was not estimated for Perryton, TX, in 2015.

Parents		Vega		Dumas	Combined	Rank
RSC†Lines	Selectionsδ	2015	2016	2016	locations	
RSC73-9	F _{2:3} High	0.627	3.200	2.312	1.198	13
RSC73-6	BC ₁ F _{2:3} High	0.361	-3.749	-0.118	-1.83	30
RSC73-1	F _{2:3} Low	-0.337	-0.271	2.755	0.347	19
RSC73-5	BC ₁ F _{2:3} Low	1.975	5.394	1.452	2.110	7
RSC83-1	F _{2:3} High	-4.594	-2.082	0.317	-3.220	38
RSC83-14	BC ₁ F _{2:3} High	-8.130	-1.189	-4.226	-4.569	39
RSC83-10	F _{2:3} Low	-3.260	-0.832	-1.111	-1.428	29
RSC83-1	BC ₁ F _{2:3} Low	-3.249	0.089	-1.460	-1.927	31
RSC112-5	F _{2:3} High	0.505	2.653	-0.306	1.662	10
RSC112-19	BC ₁ F _{2:3} High	-1.205	-2.330	-3.968	-2.079	32
RSC112-8	F _{2:3} Low	-1.768	0.375	-1.092	0.370	18
RSC112-15	BC ₁ F _{2:3} Low	-0.713	1.496	-1.590	0.987	15
RSC76-4	F _{2:3} High	1.330	-1.330	-0.735	1.185	14
RSC76-16	BC ₁ F _{2:3} High	9.232	1.885	0.108	3.340	3
RSC76-13	F _{2:3} Low	-2.782	2.839	-0.126	1.380	12
RSC76-2	BC ₁ F _{2:3} Low	7.267	0.246	-0.615	1.407	11
RSC38-5	F _{2:3} High	-0.903	-1.840	-0.643	-2.154	33
RSC38-15	BC ₁ F _{2:3} High	-3.428	-2.442	-4.237	-2.519	35
RSC38-8	F _{2:3} Low	3.621	-1.616	3.105	1.807	9
RSC38-9	BC ₁ F _{2:3} Low	0.167	-3.669	-0.556	-1.321	28
RSC37-12	F _{2:3} High	-1.453	2.868	4.44	1.831	8
RSC37-12	BC ₁ F _{2:3} High	0.365	-1.866	1.757	-0.872	24
RSC37-7	F _{2:3} Low	1.367	-0.407	0.279	2.459	6
RSC37-8	BC ₁ F _{2:3} Low	-3.742	0.243	0.960	-1.021	25
RSC15-13	F _{2:3} High	4.336	0.674	0.587	0.807	16
RSC15-15	BC ₁ F _{2:3} High	0.933	0.338	-3.578	-0.155	22
RSC15-11	F _{2:3} Low	3.374	1.944	1.767	2.825	5
RSC15-14	BC ₁ F _{2:3} Low	0.117	0.161	-0.961	0.058	20
RSC124-9	F _{2:3} High	-2.840	-2.924	1.450	-3.078	37
RSC124-16	BC ₁ F _{2:3} High	1.171	1.469	-2.148	-1.278	27
RSC124-3	F _{2:3} Low	1.512	2.930	2.267	4.666	2
RSC124-4	BC ₁ F _{2:3} Low	-4.146	0.568	2.187	-1.263	26
RSC117-2	F _{2:3} High	-6.560	-3.411	1.620	-2.967	36
RSC117-4	BC ₁ F _{2:3} High	-3.584	-3.057	-3.881	-4.964	40
RSC117-10	F _{2:3} Low	2.316	-1.235	-1.506	-0.125	21
RSC117-3	BC ₁ F _{2:3} Low	1.580	-2.852	-5.508	-2.264	34
RSC19-3	F _{2:3} High	4.827	8.674	4.540	6.403	1
RSC19-17	BC ₁ F _{2:3} High	-1.290	1.356	2.779	0.759	17
RSC19-1	F _{2:3} Low	5.401	-0.047	4.618	3.131	4
RSC19-10	BC ₁ F _{2:3} Low	1.368	-2.250	-1.131	-0.492	23
Testers						
A.301	Tester	-1.202	-1.534	-0.531	-0.932	4
A.319	Tester	1.096	-0.088	-0.841	0.156	2
A.Tx.3197	Tester	2.341	0.009	1.862	1.665	1
A.338	Tester	-2.293	1.613	-0.496	-0.902	3
SE Lines		2.7438	1.5721	1.8285	1.0909	
SE Testers		0.8677	0.4972	0.5782	0.3449	

†Testers were analyzed separately from the lines.

‡RSC represents different lines from the Reinstated Sorghum Conversion (RSC) program.

δSelections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, and the Low F_{2:3}, BC₁F_{2:3}.

Table 18. General Combining Ability (GCA) estimates and combined analysis for three-panicle weight (g) for each parental line in a line x tester evaluated in four environments. Because of the absence of 10 hybrids, individual GCA was not estimated for Perryton, TX, in 2015.

Parents		Vega		Dumas	Combined	Rank
RSC†Lines	Selectionsδ	2015	2016	2016	locations	
RSC73-9	F _{2:3} High	-8.509	-22.608	-20.159	-22.482	34
RSC73-6	BC ₁ F _{2:3} High	3.816	-20.146	-1.184	-11.304	24
RSC73-1	F _{2:3} Low	0.554	-11.271	-43.134	-18.96	31
RSC73-5	BC ₁ F _{2:3} Low	-8.459	16.304	1.754	3.021	17
RSC83-1	F _{2:3} High	-5.521	11.229	-6.709	21.405	8
RSC83-14	BC ₁ F _{2:3} High	72.466	24.079	11.454	36.343	4
RSC83-10	F _{2:3} Low	-27.021	-7.883	-45.946	-25.710	36
RSC83-1	BC ₁ F _{2:3} Low	20.091	-6.858	40.591	17.977	10
RSC112-5	F _{2:3} High	-32.821	24.642	45.891	4.468	16
RSC112-19	BC ₁ F _{2:3} High	-26.852	-2.108	33.327	15.592	12
RSC112-8	F _{2:3} Low	-26.659	-34.583	10.029	-17.542	29
RSC112-15	BC ₁ F _{2:3} Low	8.254	16.029	18.154	5.702	15
RSC76-4	F _{2:3} High	40.341	24.917	28.116	22.499	7
RSC76-16	BC ₁ F _{2:3} High	50.429	46.729	74.766	55.208	2
RSC76-13	F _{2:3} Low	-36.071	18.204	-8.721	-11.020	22
RSC76-2	BC ₁ F _{2:3} Low	126.541	63.054	65.329	90.480	1
RSC38-5	F _{2:3} High	-33.059	-21.921	1.216	-20.885	32
RSC38-15	BC ₁ F _{2:3} High	-8.059	-14.683	33.866	1.758	18
RSC38-8	F _{2:3} Low	21.604	6.554	22.854	18.205	9
RSC38-9	BC ₁ F _{2:3} Low	-26.284	-12.633	0.041	-13.242	26
RSC37-12	F _{2:3} High	-7.171	-23.758	-7.521	-16.007	27
RSC37-12	BC ₁ F _{2:3} High	63.416	29.892	18.666	38.546	3
RSC37-7	F _{2:3} Low	11.529	9.129	-3.371	10.962	13
RSC37-8	BC ₁ F _{2:3} Low	40.629	50.567	-9.271	24.271	6
RSC15-13	F _{2:3} High	-47.684	-27.871	-60.359	-39.348	39
RSC15-15	BC ₁ F _{2:3} High	1.616	-11.358	-25.859	-6.704	20
RSC15-11	F _{2:3} Low	-45.452	-20.296	-29.696	-35.541	37
RSC15-14	BC ₁ F _{2:3} Low	-35.909	-31.171	-28.146	-23.826	35
RSC124-9	F _{2:3} High	3.791	-24.008	-25.384	-18.51	30
RSC124-16	BC ₁ F _{2:3} High	10.341	25.817	63.716	36.184	5
RSC124-3	F _{2:3} Low	-25.221	-34.133	-52.721	-39.345	38
RSC124-4	BC ₁ F _{2:3} Low	-9.895	-24.721	-16.521	-21.009	33
RSC117-2	F _{2:3} High	-11.934	-13.433	-23.921	-12.332	25
RSC117-4	BC ₁ F _{2:3} High	-22.884	25.579	16.291	9.758	14
RSC117-10	F _{2:3} Low	-25.768	-1.208	-10.434	-8.047	21
RSC117-3	BC ₁ F _{2:3} Low	-10.371	15.517	22.629	16.371	11
RSC19-3	F _{2:3} High	2.766	-11.033	-26.371	-16.285	28
RSC19-17	BC ₁ F _{2:3} High	-5.621	3.492	16.654	-11.113	23
RSC19-1	F _{2:3} Low	-29.284	-20.733	-61.909	-41.363	40
RSC19-10	BC ₁ F _{2:3} Low	23.566	-13.321	-13.846	-3.457	19
Testers						
A.301	Tester	-15.729	-11.814	-14.29	-15.139	4
A.319	Tester	-6.649	8.251	2.81	3.361	2
A.Tx.3197	Tester	-1.716	-10.799	-8.129	-12.796	3
A.338	Tester	24.426	14.363	19.43	24.383	1
SE Lines		16.379	8.8899	15.6975	8.869	
SE Testers		5.179	2.811	4.964	2.805	

†Testers were analyzed separately from the lines.

‡RSC represents different lines from the Reinstated Sorghum Conversion (RSC) program.

δSelections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, and the Low F_{2:3}, BC₁F_{2:3}.

Table 19. General Combining Ability (GCA) estimates and combined analysis for 1000-kernel weight (g) for each parental line in a line x tester evaluated in four environments. Because of the absence of 10 hybrids, individual GCA was not estimated for Perryton, TX, in 2015.

Parents		Vega		Dumas	Combined	Rank
RSC†Lines	Selectionsδ	2015	2016	2016	locations	
RSC73-9	F _{2:3} High	-0.575	-2.274	-1.665	-1.354	27
RSC73-6	BC ₁ F _{2:3} High	-0.254	-1.308	-1.802	-1.617	30
RSC73-1	F _{2:3} Low	-0.796	0.814	0.424	0.133	16
RSC73-5	BC ₁ F _{2:3} Low	0.96	0.087	2.346	1.283	8
RSC83-1	F _{2:3} High	0.783	0.1000	1.362	0.885	11
RSC83-14	BC ₁ F _{2:3} High	1.035	1.032	-0.565	0.412	14
RSC83-10	F _{2:3} Low	1.280	-1.841	-0.736	-1.055	23
RSC83-1	BC ₁ F _{2:3} Low	2.748	0.724	0.173	0.710	12
RSC112-5	F _{2:3} High	3.994	-0.775	-0.69	1.400	7
RSC112-19	BC ₁ F _{2:3} High	-1.448	1.209	-1.529	-0.517	19
RSC112-8	F _{2:3} Low	-1.102	-0.722	-1.097	-1.170	24
RSC112-15	BC ₁ F _{2:3} Low	0.603	1.675	-0.401	0.173	15
RSC76-4	F _{2:3} High	6.643	6.150	7.345	6.849	2
RSC76-16	BC ₁ F _{2:3} High	7.453	7.062	7.481	7.360	1
RSC76-13	F _{2:3} Low	5.113	3.108	5.073	4.720	4
RSC76-2	BC ₁ F _{2:3} Low	6.514	5.853	6.468	6.536	3
RSC38-5	F _{2:3} High	-4.686	-2.849	-4.089	-3.705	39
RSC38-15	BC ₁ F _{2:3} High	-1.121	-1.365	1.413	-0.446	18
RSC38-8	F _{2:3} Low	0.594	-0.585	0.128	-0.043	17
RSC38-9	BC ₁ F _{2:3} Low	-1.113	-0.893	-1.399	-1.186	25
RSC37-12	F _{2:3} High	-1.223	-1.626	-2.96	-2.097	35
RSC37-12	BC ₁ F _{2:3} High	0.244	0.566	-2.897	-0.733	21
RSC37-7	F _{2:3} Low	-2.096	-0.635	-0.922	-0.909	22
RSC37-8	BC ₁ F _{2:3} Low	-4.577	-2.343	-3.494	-3.078	38
RSC15-13	F _{2:3} High	0.343	0.432	3.383	1.283	9
RSC15-15	BC ₁ F _{2:3} High	-1.773	-0.548	-0.44	-1.640	31
RSC15-11	F _{2:3} Low	-2.097	-2.228	-0.056	-1.219	26
RSC15-14	BC ₁ F _{2:3} Low	1.630	0.938	2.432	1.476	6
RSC124-9	F _{2:3} High	-4.022	-2.03	-1.849	-2.623	36
RSC124-16	BC ₁ F _{2:3} High	-3.289	-1.469	-0.927	-1.968	34
RSC124-3	F _{2:3} Low	0.448	0.155	-0.363	0.484	13
RSC124-4	BC ₁ F _{2:3} Low	-1.109	-1.918	-2.693	-1.818	33
RSC117-2	F _{2:3} High	-0.164	-1.189	-0.498	-0.567	20
RSC117-4	BC ₁ F _{2:3} High	1.642	4.374	4.173	3.287	5
RSC117-10	F _{2:3} Low	-2.939	-0.537	-0.856	-1.441	28
RSC117-3	BC ₁ F _{2:3} Low	-0.797	2.141	1.066	1.144	10
RSC19-3	F _{2:3} High	-2.450	-2.087	-2.106	-2.645	37
RSC19-17	BC ₁ F _{2:3} High	-0.922	-2.157	-1.563	-1.647	32
RSC19-1	F _{2:3} Low	-4.269	-2.915	-4.954	-4.142	40
RSC19-10	BC ₁ F _{2:3} Low	0.417	-2.128	-2.907	-1.509	29
Testers						
A.301	Tester	-0.048	0.668	0.568	0.398	3
A.319	Tester	-2.482	-2.014	-3.075	-2.346	4
A.Tx.3197	Tester	1.547	0.104	1.29	0.849	2
A.338	Tester	0.976	1.241	1.224	1.104	1
SE Lines		0.9494	0.8049	0.9795	0.5241	
SE Testers		0.300	0.255	0.310	0.167	

†Testers were analyzed separately from the lines.

‡RSC represents different lines from the Reinstated Sorghum Conversion (RSC) program.

δSelections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, and the Low F_{2:3}, BC₁F_{2:3}.

for testers was from Dumas in 2016, for A.301; much variation was found across environments (Table 11). The greatest combined GCA effect (0.746) for lines was found in RSC15-14 BC₁F_{2:3} low, followed by RSC-15-13 F_{2:3} high, RSC19-17 BC₁F_{2:3} high, RSC15-11F_{2:3} low, and RSC38-15BC₁F_{2:3} high. For concentration of protein in grain, GCA effects of 0.723, 0.641, 0.599, and 0.543, respectively, were found for the RSC15 family that dominated the GCA effect. A.Tx3197 had the greatest GCA effect (0.081) for testers for concentration of starch in grain, while the RSC112 family ranked 1st, 2nd, and 4th best as a good combiner for starch in grain. A.301 also had the greatest GCA effect (0.013) for concentration of fiber in grain, and RSC37-12, a BC₁ low line, followed by RSC117-4 (BC₁F_{2:3} high), RSC19-3 (F_{2:3} high), RSC19-1 (F_{2:3} low), and RSC73-5 (BC₁F_{2:3} low) had GCA effects of 0.043, 0.033, 0.031, 0.031, and 0.029, respectively. Greatest tester GCA effect for concentration of fat in grain was reported for A.338 (0.118). Greatest line GCA effect was found for RSC19-17 (BC₁F_{2:3} high) with a GCA effect of 0.418, followed by RSC15-11 (F_{2:3} low), RSC38-15 (BC₁F_{2:3} high), and RSC15-14 (BC₁F_{2:3} low) with GCA effects of 0.357, 0.315, and 0.283, respectively. The RSC family seemed to play a greater role in the combining ability for quality grain analysis. The families RSC15, RSC19, RSC112, RSC73, and RSC117 all seemed to be good combiners for high concentrations of protein, starch, fiber, and fat in grain, depending on the quality factors a sorghum breeder is seeking. The lines or testers with positive and significant GCA effects for plant height also showed good mean performance, indicating a strong relationship between GCA status and *per se* performance and can be considered good parents, in agreement with earlier reports by Iyanar et al. (2001), Kanawade et al. (2001), Chaudhary et al. (2006), and Kulakarni et al. (2006). This might help in selection

of parents on the basis of *per se* performance in the absence of information on combining ability of inbreds.

Specific Combining Ability (SCA)

SCA effect represents the non-fixable component of genetic variation that provides information on hybrid performance. The range of combined SCA effects for grain yield varied from -1.443 (A.338*RSC76-16 BC₁F_{2:3} high) to 1.334 (A.338*RSC117-4 BC₁F_{2:3} high), with yields from 4.978 to 8.060 Mg ha⁻¹, respectively. Of the 160 crosses, A.338*RSC117-4, A.301*RSC76-2 (BC₁F_{2:3} low), A.301*RSC19-17 (BC₁F_{2:3} high), A.338*RSC117-2 (F_{2:3} high), and A.301*RSC76-4 (F_{2:3} high) had positive SCA effects greater than one and were identified as good specific combiners for grain yield (Table 20). Respectively, the crosses involved low x high, high x low, high x low, low x high, and high x low combinations in terms of GCA effects, indicating additive x dominance type of gene interaction. Thus, it can be concluded that inter- and intra-allelic interactions were involved in expression of the trait. A.301* RSC117-4 (BC₁F_{2:3} high), A.Tx3197*RSC19-10 (BC₁F_{2:3} high), and A.338*RSC76-16 (BC₁F_{2:3} high) had the most negative SCA effects of -1.206, -1.275, and -1.443, with grain yields of 6.163, 4.515, and 3.072 Mg ha⁻¹, respectively. Although the two selections from RSC76 showed good SCA with A.301, the overall mean performance was less because of poor combining ability with the other three testers.

A.338*RSC117-4 BC₁F_{2:3} high had significant positive MPH and large overall mean; however, the hybrid had a mean of 75.8 days to anthesis and 135.8 cm total plant height, out-yielding three of the best six commercial hybrid checks.

Table 20. Specific Combining Ability (SCA) estimates for grain yield (Mgha⁻¹) and overall rank for each hybrid combination in the combined analysis across six environments.

Lines		A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC†	Selections‡								
RSC73-9	F _{2:3} High	-0.164	101	0.133	63	0.456	31	-0.425	127
RSC73-6	BC ₁ F _{2:3} High	-0.051	88	0.156	57	-0.078	92	-0.027	84
RSC73-1	F _{2:3} Low	-0.300	114	-0.212	104	0.320	41	0.192	53
RSC73-5	BC ₁ F _{2:3} Low	0.331	39	-0.002	80	0.109	66	-0.437	128
RSC83-1	F _{2:3} High	-0.012	82	0.234	45	-0.962	153	0.74	16
RSC83-14	BC ₁ F _{2:3} High	0.269	42	-0.031	86	-0.340	117	0.102	67
RSC83-10	F _{2:3} Low	-0.408	125	-0.369	120	-0.077	91	0.855	10
RSC83-1	BC ₁ F _{2:3} Low	-0.623	143	0.983	6	0.096	69	-0.456	132
RSC112-5	F _{2:3} High	-0.158	99	-0.448	131	0.822	12	-0.215	105
RSC112-19	BC ₁ F _{2:3} High	0.124	64	-0.06	90	0.212	49	-0.276	112
RSC112-8	F _{2:3} Low	-0.685	147	-0.219	106	0.559	25	0.344	37
RSC112-15	BC ₁ F _{2:3} Low	-0.494	137	-0.575	142	0.549	26	0.519	28
RSC76-4	F _{2:3} High	1.009	5	-0.057	89	0.163	55	-1.116	156
RSC76-16	BC ₁ F _{2:3} High	0.872	9	0.701	17	-0.130	96	-1.443	160
RSC76-13	F _{2:3} Low	0.581	23	0.437	33	-0.640	145	-0.378	123
RSC76-2	BC ₁ F _{2:3} Low	1.231	2	-0.709	151	0.177	54	-0.699	149
RSC38-5	F _{2:3} High	-0.565	140	0.968	7	0.752	15	-1.155	157
RSC38-15	BC ₁ F _{2:3} High	0.633	21	0.079	70	-0.418	126	-0.295	113
RSC38-8	F _{2:3} Low	0.231	46	0.151	58	0.269	43	-0.650	146
RSC38-9	BC ₁ F _{2:3} Low	-0.152	98	0.052	75	0.793	14	-0.694	148
RSC37-12	F _{2:3} High	-0.466	134	0.481	30	-0.029	85	0.014	78
RSC37-12	BC ₁ F _{2:3} High	0.438	32	0.201	52	-0.013	83	-0.626	144
RSC37-7	F _{2:3} Low	-0.500	138	0.568	24	-0.140	97	0.072	72
RSC37-8	BC ₁ F _{2:3} Low	0.202	50	0.226	47	-0.446	130	0.018	77
RSC15-13	F _{2:3} High	0.428	34	-0.090	93	-1.003	154	0.665	20
RSC15-15	BC ₁ F _{2:3} High	-0.227	108	-0.800	152	0.926	8	0.101	68
RSC15-11	F _{2:3} Low	0.070	73	0.245	44	-1.008	155	0.694	18
RSC15-14	BC ₁ F _{2:3} Low	-0.313	115	-0.372	122	0.143	59	0.542	27
RSC124-9	F _{2:3} High	-0.442	129	-0.313	116	0.592	22	0.163	56
RSC124-16	BC ₁ F _{2:3} High	0.062	74	-0.388	124	0.419	36	-0.093	94
RSC124-3	F _{2:3} Low	0.076	71	0.329	40	-0.369	121	-0.036	87
RSC124-4	BC ₁ F _{2:3} Low	0.002	79	-0.471	136	0.335	38	0.134	62
RSC117-2	F _{2:3} High	-0.462	133	-0.344	119	-0.226	107	1.032	4
RSC117-4	BC ₁ F _{2:3} High	-1.206	158	-0.243	110	0.115	65	1.334	1
RSC117-10	F _{2:3} Low	-0.549	139	-0.006	81	0.135	61	0.420	35
RSC117-3	BC ₁ F _{2:3} Low	-0.467	135	-0.176	102	-0.160	100	0.804	13
RSC19-3	F _{2:3} High	0.018	76	-0.703	150	0.202	51	0.483	29
RSC19-17	BC ₁ F _{2:3} High	1.103	3	-0.342	118	-0.566	141	-0.195	103
RSC19-1	F _{2:3} Low	-0.102	95	0.14	60	-0.263	111	0.225	48
RSC19-10	BC ₁ F _{2:3} Low	0.668	19	0.846	11	-1.275	159	-0.239	109
SE hybrid	0.537								

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

A.301*RSC76-2 BC₁F_{2:3} low had large positive MPH and large overall mean *per se* parents; however, the hybrid was earlier (71.2 days to anthesis) and taller (175.3 cm total plant height). A.301*RSC19-17 BC₁F_{2:3} high had significant positive MPH and large overall mean grain yield while being earlier (67.5 days to anthesis) and shorter (152.9 cm total plant height). A.338*RSC117-2 F_{2:3} high had high MPH and large overall mean grain yield, with 77.0 days to anthesis and 146.5 cm total plant height. For grain yield, SCA variance (0.0866) was greater than that of the GCA variance (0.0054), indicating dominance of non-additive gene action in inheritance of the trait (Table 22). Similar trends of results were reported by Kenga et al. (2004), El-Menshawi (2005), Mahdy et al. (2011), and Rani et al. (2015). Additive gene action for controlling the trait was reported by Kenga et al. (2005) and Tadesse et al. (2008). Importance of both additive and non-additive gene action in inheritance of the trait was reported by El-Mottaleb (2009) and Makanda et al. (2010).

The range of combined SCA effects for days to anthesis varied from -5.648 (A.338*RSC37-12 F_{2:3} high) to 6.164 (A.319*RSC37-12 F_{2:3} high), with 79.3 to 75.5 days, respectively (Tables 3 and 21). The earliest hybrids were crosses with tester A.301 and lines RSC124-4 (BC₁F_{2:3} low), RSC19-17 (BC₁F_{2:3} high), RSC15-15 (BC₁F_{2:3} high), RSC37-8 (BC₁F_{2:3} low), and RSC112-15 (BC₁F_{2:3} low), ranging in days to anthesis of 66.3 to 67.0, with SCA effects of 0.814, -0.936, 0.939, -1.311, and -0.998, respectively. The results of combining ability for the number of days to anthesis among lines RSC73-9 F_{2:3} high, RSC73 BC₁F_{2:3} high, and RSC117-2 F_{2:3} high, and the family of RSC38 and RSC15 among testers A.301 and A.Tx3197 flowered early, because they showed

Table 21. Specific Combining Ability (SCA) estimates for day to anthesis for each hybrid combination in the combined analysis across two environments with overall rank. Because of a missing entry, data for Hutchinson, KS, in 2015 were not included.

Lines		A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC†	Selections‡								
RSC73-9	F _{2:3} High	0.127	71	-0.398	88	-1.267	128	1.539	26
RSC73-6	BC ₁ F _{2:3} High	0.439	61	-1.086	119	-0.205	81	0.852	43
RSC73-1	F _{2:3} Low	-4.061	157	1.164	34	3.795	8	-0.898	102
RSC73-5	BC ₁ F _{2:3} Low	-1.873	137	1.352	30	0.983	37	-0.461	93
RSC83-1	F _{2:3} High	-2.436	153	-0.961	108	4.420	5	-1.023	115
RSC83-14	BC ₁ F _{2:3} High	-0.873	99	0.352	66	-0.017	75	0.539	55
RSC83-10	F _{2:3} Low	-1.123	123	1.602	25	1.483	27	-1.961	142
RSC83-1	BC ₁ F _{2:3} Low	-1.998	143	1.977	20	0.608	51	-0.586	94
RSC112-5	F _{2:3} High	3.939	7	0.914	41	-0.205	82	-4.648	159
RSC112-19	BC ₁ F _{2:3} High	3.689	9	-1.086	120	-2.205	147	-0.398	89
RSC112-8	F _{2:3} Low	-2.248	150	-1.023	113	1.608	24	1.664	23
RSC112-15	BC ₁ F _{2:3} Low	-0.998	109	0.227	68	0.108	72	0.664	50
RSC76-4	F _{2:3} High	0.377	64	-0.648	95	3.233	10	-2.961	155
RSC76-16	BC ₁ F _{2:3} High	3.127	12	-0.898	100	0.483	59	-2.711	154
RSC76-13	F _{2:3} Low	0.002	74	-1.023	114	1.358	29	-0.336	86
RSC76-2	BC ₁ F _{2:3} Low	2.502	17	-1.273	129	-0.392	87	-0.836	98
RSC38-5	F _{2:3} High	-1.186	126	-0.211	83	0.17	69	1.227	33
RSC38-15	BC ₁ F _{2:3} High	-0.123	79	-2.398	151	-1.517	133	4.039	6
RSC38-8	F _{2:3} Low	-0.436	91	-2.211	148	-2.08	145	4.727	3
RSC38-9	BC ₁ F _{2:3} Low	0.564	52	-4.211	158	-1.08	118	4.727	4
RSC37-12	F _{2:3} High	0.439	62	6.164	1	-0.955	107	-5.648	160
RSC37-12	BC ₁ F _{2:3} High	0.502	57	1.977	21	-1.142	124	-1.336	131
RSC37-7	F _{2:3} Low	-0.936	105	2.539	16	-1.580	135	-0.023	76
RSC37-8	BC ₁ F _{2:3} Low	-1.311	130	1.664	22	-1.705	136	1.352	31
RSC15-13	F _{2:3} High	1.127	35	-2.398	152	-0.767	97	2.039	18
RSC15-15	BC ₁ F _{2:3} High	0.939	39	-1.086	121	-1.205	127	1.352	32
RSC15-11	F _{2:3} Low	-1.873	138	3.102	13	-1.017	111	-0.211	84
RSC15-14	BC ₁ F _{2:3} Low	0.377	65	-1.898	140	0.983	38	0.539	56
RSC124-9	F _{2:3} High	0.564	53	-2.211	149	0.17	70	1.477	28
RSC124-16	BC ₁ F _{2:3} High	5.689	2	-3.336	156	-1.455	132	-0.898	103
RSC124-3	F _{2:3} Low	0.314	67	-1.961	141	0.92	40	0.727	47
RSC124-4	BC ₁ F _{2:3} Low	0.814	44	2.789	15	-2.08	146	-1.523	134
RSC117-2	F _{2:3} High	-1.061	116	0.414	63	-0.455	92	1.102	36
RSC117-4	BC ₁ F _{2:3} High	-0.311	85	0.664	48	0.545	54	-0.898	104
RSC117-10	F _{2:3} Low	0.502	58	0.477	60	-1.892	139	0.914	42
RSC117-3	BC ₁ F _{2:3} Low	-0.123	80	-0.898	101	-1.017	112	2.039	19
RSC19-3	F _{2:3} High	-1.061	117	0.664	49	0.795	45	-0.398	90
RSC19-17	BC ₁ F _{2:3} High	-0.936	106	0.039	73	2.92	14	-2.023	144
RSC19-1	F _{2:3} Low	-0.998	110	3.227	11	-1.142	125	-1.086	122
RSC19-10	BC ₁ F _{2:3} Low	-0.061	77	-0.086	78	0.795	46	-0.648	96
SE hybrid	1.224								

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

negative GCA effects and projected their use in sorghum breeding programs for earliness (Table 13). Of the 160 crosses, A.338*RSC37-12 $F_{2:3}$ high (high x low), A.338*RSC112-5 $F_{2:3}$ high (high x high), A.319*RSC38-9 $BC_1F_{2:3}$ low (high x low), and A.301*RSC73-1 $F_{2:3}$ low (low x high) had the greatest SCA effect in the negative direction and were considered the best specific crosses for early maturity (Tables 13 and 21). A.301*RSC73-1 $F_{2:3}$ low (low x high) flowered in 68.5 days as compared to the line and testers at 71 and 65.5 days, respectively. For days to anthesis, the estimate of SCA variance (3.357) was greater than that of the GCA variance (0.296), with the ratio of variance due to GCA and SCA 0.046 indicating dominance of non-additive gene action in inheritance of the trait (Table 22). Similar results were reported by Chaudhary et al. (2006) and Premalatha et al. (2006), while opposite results were reported by Kenga et al. (2005) and Mahdy et al. (2011). The importance of both additive and non-additive gene action in inheritance of the trait was reported by Hovny et al. (2005), Mahmoud (2007), and El-Mottaleb (2009).

According to Kenga et al. (2004), a cross combination with large means, favorable SCA estimate, and involving at least one of the parents with great GCA effects probably would enhance the concentration of favorable alleles to improve targeted traits. In crosses with high x low and low x high GCA effects, selection should be delayed until further segregating generations, by which time heterozygosity would be less, while homozygosity and additive genes could be stabilized (Rani et al., 2015).

The range of combined SCA effects for total plant height varied from -35.357 (A.301*RSC37-7 $F_{2:3}$ low) to 37.428 (A.338*RSC38-5 $F_{2:3}$ high), with total heights

Table 22. Estimates of genetic components and proportional contribution to the total variation of the line x tester analysis for the measured traits in 160 sorghum hybrids across environments.

Genetic Component	Grain yield (Mgha ⁻¹)	DTF	Total height (cm)	Height to flag leaf (cm)	Panicle length (cm)	Plant exsertion (cm)	3-panicle weight(g)	1000-kernel weight(g)	Protein (%)	Starch (%)	Fiber (%)	Fat (%)
σ^2 GCA line	0.2895	0.2962	847.8684	895.0326	4.0885	4.6573	633.7928	6.6013	0.00	0.00	0.00	0.00
σ^2 GCA tester	0.1456	13.1754	673.7420	689.4357	1.4352	1.3348	307.6490	2.4800	0.00	0.00	0.00	0.00
σ^2 GCA	0.0054	0.1529	18.3848	19.1820	0.0699	0.0765	11.7141	0.1145	0.00	0.00	0.00	0.00
σ^2 SCA	0.0866	3.3569	92.0435	94.1777	0.3589	0.0506	115.3883	0.4447	0.00	0.00	0.00	0.00
σ^2 GCA/ σ^2 SCA	0.0624	0.0455		0.2037	0.1947	1.5122	0.1015	0.2575	0.00	0.00	0.00	0.00
SCA			0.1997									
σ^2 A	0.0108	0.3057	36.7696	38.3639	0.1398	0.1530	23.4282	0.2290	0.00	0.00	0.00	0.00
σ^2 D	0.0866	3.3569	92.0435	94.1777	0.3589	0.0506	115.3883	0.4447	0.00	0.00	0.00	0.00
$[\sigma^2 D/\sigma^2 A]^{1/2}$	2.8318	3.3135	1.5822	1.5668	1.6024	0.5750	2.2193	1.3935	N/A	N/A	N/A	N/A
Proportional contribution to total variation (%)												
Lines (L)	48.90	12.72	58.33	59.12	69.50	55.35	56.64	69.30	53.02	49.27	44.90	65.79
Testers (T)	15.21	73.77	34.35	33.82	17.96	10.57	18.71	19.22	9.33	1.56	19.83	12.01
L x T	35.89	13.51	7.33	7.06	12.54	34.08	24.64	11.48	37.65	49.17	35.28	22.20

DTF = days to flowering.

σ^2 A: additive genetic variance, σ^2 D: dominance genetic variance, σ^2 GCA: estimate of GCA variance, σ^2 SCA: estimate of SCA variance, σ^2 GCA/ σ^2 SCA: average degree of dominance, $[\sigma^2 D/\sigma^2 A]^{1/2}$: degree of dominance.

*Negative component interpreted as zero.

ranging from 101.04 to 212.80 cm, respectively. A.338*RSC38-5 F_{2:3} high cross combination had positive SCA for total plant height and 1000-kernel weight. The cross is the best specific combiner to increase plant height and 1000-kernel weight with a high x low combination of GCA effects and with a large SCA effect further substantiating the operation of non-additive gene action. However, crosses such as A.301*RSC37-7 F_{2:3} low and A.338*RSC112-5 F_{2:3} high had negative SCA effects with GCA effects of low x high and high x low, respectively, which would enable good combiners to reduce plant height. For total plant height, the estimate of SCA variance (92.04) was greater than that of the GCA variance (18.38), with a ratio of 0.199 indicating dominance of non-additive gene action in inheritance of the trait (Table 23). Rani et al. (2015) reported similar results for plant height. Dominance variance was more than additive variance for the trait. The results are in agreement with earlier reports by Kulakarni et al. (2006), Yadav and Pahuja (2007), Fellahi et al. (2013), and Rani et al. (2015), while opposite results were reported by Tadesse et al. (2008) and Degu et al. (2009).

The range of combined SCA effects for the height to the flag leaf varied from -34.343 (A.301*RSC37-7 F_{2:3} low) to 39.778 (A.338*RSC38-5 F_{2:3} high), with the height to the flag leaf ranging from 69.1 to 179.6 cm, respectively (Table 24). This followed the same trend as total plant height, with crosses such as A.301*RSC37-7 F_{2:3} low and A.338*RSC112-5 F_{2:3} high being the best combiners to reduce plant height. The crosses had negative SCA effects with GCA effects of low x high and high x low, respectively, which would make good combiners to reduce plant height. For height to the flag leaf, the estimate of SCA variance (94.18) was greater than the estimate of GCA variance (19.18),

Table 23. Specific Combining Ability (SCA) estimates for total plant height (cm) for each hybrid combination in the combined analysis across four environments with overall rank.

RSC†	Lines Selections‡	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC73-9	F _{2:3} High	-5.142	117	-1.387	98	0.203	79	6.326	35
RSC73-6	BC ₁ F _{2:3} High	1.248	69	-1.136	97	-4.253	114	4.142	49
RSC73-1	F _{2:3} Low	-12.736	145	0.491	72	4.782	42	7.463	31
RSC73-5	BC ₁ F _{2:3} Low	-20.077	157	1.721	64	2.414	58	15.943	10
RSC83-1	F _{2:3} High	-5.287	118	1.219	70	-6.661	125	10.730	24
RSC83-14	BC ₁ F _{2:3} High	-0.911	96	2.053	62	-0.536	91	-0.607	92
RSC83-10	F _{2:3} Low	14.317	15	-0.765	93	-6.212	123	-7.34	128
RSC83-1	BC ₁ F _{2:3} Low	-0.857	95	-2.867	106	4.492	46	-0.767	94
RSC112-5	F _{2:3} High	16.686	8	5.095	38	4.412	48	-26.193	159
RSC112-19	BC ₁ F _{2:3} High	8.549	28	-2.123	102	0.474	73	-6.900	127
RSC112-8	F _{2:3} Low	14.623	14	0.333	75	-2.416	104	-12.54	144
RSC112-15	BC ₁ F _{2:3} Low	2.361	59	2.815	56	-0.462	90	-4.714	115
RSC76-4	F _{2:3} High	-16.438	149	1.602	65	3.670	52	11.166	23
RSC76-16	BC ₁ F _{2:3} High	-19.007	154	-0.433	89	9.941	25	9.499	26
RSC76-13	F _{2:3} Low	-5.963	121	0.226	77	6.158	36	-0.421	88
RSC76-2	BC ₁ F _{2:3} Low	-18.292	153	1.762	63	8.908	27	7.621	30
RSC38-5	F _{2:3} High	-6.109	122	-12.248	141	-19.071	155	37.428	1
RSC38-15	BC ₁ F _{2:3} High	-19.282	156	-1.878	100	1.143	71	20.018	5
RSC38-8	F _{2:3} Low	-12.407	143	3.470	54	3.843	50	5.095	39
RSC38-9	BC ₁ F _{2:3} Low	-9.454	133	-3.690	112	0.073	83	13.071	17
RSC37-12	F _{2:3} High	-5.366	119	15.692	12	4.846	41	-15.172	147
RSC37-12	BC ₁ F _{2:3} High	1.407	67	-10.079	135	7.387	32	1.285	68
RSC37-7	F _{2:3} Low	-35.357	160	13.59	16	4.700	43	17.068	7
RSC37-8	BC ₁ F _{2:3} Low	-7.735	130	0.465	74	2.744	57	4.525	45
RSC15-13	F _{2:3} High	16.073	9	-22.453	158	6.548	34	-0.168	84
RSC15-15	BC ₁ F _{2:3} High	11.883	20	0.187	80	-0.338	87	-11.733	138
RSC15-11	F _{2:3} Low	28.214	2	-11.741	139	0.224	78	-16.696	150
RSC15-14	BC ₁ F _{2:3} Low	4.462	47	15.31	13	-3.527	110	-16.245	148
RSC124-9	F _{2:3} High	15.734	11	3.561	53	-8.714	132	-10.582	136
RSC124-16	BC ₁ F _{2:3} High	-4.238	113	4.667	44	-7.729	129	7.299	33
RSC124-3	F _{2:3} Low	-0.247	86	-6.812	126	2.135	60	4.924	40
RSC124-4	BC ₁ F _{2:3} Low	0.265	76	-12.05	140	-6.244	124	18.029	6
RSC117-2	F _{2:3} High	11.211	22	-2.975	107	0.151	81	-8.387	131
RSC117-4	BC ₁ F _{2:3} High	12.560	18	2.131	61	-1.887	101	-12.805	146
RSC117-10	F _{2:3} Low	11.383	21	3.812	51	-2.853	105	-12.342	142
RSC117-3	BC ₁ F _{2:3} Low	26.595	3	-4.734	116	-3.618	111	-18.243	152
RSC19-3	F _{2:3} High	6.011	37	-3.04	108	0.138	82	-3.109	109
RSC19-17	BC ₁ F _{2:3} High	-11.214	137	11.962	19	-2.268	103	1.520	66
RSC19-1	F _{2:3} Low	22.414	4	-0.183	85	-5.471	120	-16.76	151
RSC19-10	BC ₁ F _{2:3} Low	-9.879	134	8.43	29	2.876	55	-1.426	99
SE hybrid	7.493								

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

Table 24. Specific Combining Ability (SCA) estimates for height to flag leaf (cm) for each hybrid combination in the combined analysis across four environments with overall rank.

RSC†	Lines Selections‡	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC73-9	F _{2:3} High	-3.570	109	-1.115	91	0.850	79	3.834	50
RSC73-6	BC ₁ F _{2:3} High	-0.343	86	-1.378	96	-1.266	94	2.988	57
RSC73-1	F _{2:3} Low	-15.677	150	1.704	70	7.162	30	6.811	32
RSC73-5	BC ₁ F _{2:3} Low	-18.731	155	2.353	60	1.512	74	14.866	13
RSC83-1	F _{2:3} High	-2.089	99	-0.056	83	-7.670	129	9.814	25
RSC83-14	BC ₁ F _{2:3} High	-1.283	95	3.820	51	1.658	72	-4.195	114
RSC83-10	F _{2:3} Low	12.013	19	2.192	65	-5.420	119	-8.786	131
RSC83-1	BC ₁ F _{2:3} Low	-3.703	110	-0.135	84	5.004	41	-1.166	93
RSC112-5	F _{2:3} High	17.545	9	6.294	35	2.597	59	-26.436	158
RSC112-19	BC ₁ F _{2:3} High	7.844	28	-2.326	102	-0.467	87	-5.051	117
RSC112-8	F _{2:3} Low	13.520	16	1.215	78	-3.487	107	-11.247	136
RSC112-15	BC ₁ F _{2:3} Low	3.741	52	2.250	62	-0.704	89	-5.287	118
RSC76-4	F _{2:3} High	-12.780	142	2.221	63	-0.949	90	11.508	21
RSC76-16	BC ₁ F _{2:3} High	-15.014	148	-1.129	92	5.920	37	10.223	24
RSC76-13	F _{2:3} Low	-6.812	124	-0.652	88	6.976	31	0.488	81
RSC76-2	BC ₁ F _{2:3} Low	-12.580	141	-2.291	101	8.934	26	5.938	36
RSC38-5	F _{2:3} High	-7.474	126	-13.059	146	-19.245	157	39.778	1
RSC38-15	BC ₁ F _{2:3} High	-16.905	152	-1.749	98	-2.272	100	20.926	4
RSC38-8	F _{2:3} Low	-15.305	149	3.926	49	4.939	42	6.440	33
RSC38-9	BC ₁ F _{2:3} Low	-11.324	137	-6.168	121	2.201	64	15.291	12
RSC37-12	F _{2:3} High	-7.644	128	18.307	8	5.452	39	-16.115	151
RSC37-12	BC ₁ F _{2:3} High	1.444	76	-7.054	125	5.918	38	-0.308	85
RSC37-7	F _{2:3} Low	-34.343	160	11.825	20	3.681	53	18.837	6
RSC37-8	BC ₁ F _{2:3} Low	-10.622	135	0.779	80	1.842	67	8.002	27
RSC15-13	F _{2:3} High	18.769	7	-26.505	159	4.511	46	3.225	56
RSC15-15	BC ₁ F _{2:3} High	13.005	17	3.502	54	-3.529	108	-12.978	145
RSC15-11	F _{2:3} Low	26.609	2	-12.950	144	-1.406	97	-12.252	140
RSC15-14	BC ₁ F _{2:3} Low	5.014	40	16.520	10	-2.576	105	-18.958	156
RSC124-9	F _{2:3} High	14.410	14	2.900	58	-7.623	127	-9.686	134
RSC124-16	BC ₁ F _{2:3} High	-6.407	122	4.677	44	-4.665	115	6.396	34
RSC124-3	F _{2:3} Low	2.118	66	-6.644	123	3.259	55	1.268	77
RSC124-4	BC ₁ F _{2:3} Low	4.139	48	-14.151	147	-5.534	120	15.546	11
RSC117-2	F _{2:3} High	10.344	23	-2.435	103	1.750	69	-9.658	133
RSC117-4	BC ₁ F _{2:3} High	12.257	18	4.181	47	-4.774	116	-11.664	138
RSC117-10	F _{2:3} Low	13.706	15	1.77	68	-3.727	111	-11.749	139
RSC117-3	BC ₁ F _{2:3} Low	24.846	3	-8.522	130	1.486	75	-17.809	154
RSC19-3	F _{2:3} High	4.618	45	-2.503	104	1.684	71	-3.799	112
RSC19-17	BC ₁ F _{2:3} High	-9.407	132	10.990	22	-3.924	113	2.341	61
RSC19-1	F _{2:3} Low	18.866	5	1.636	73	-2.801	106	-17.702	153
RSC19-10	BC ₁ F _{2:3} Low	-12.793	143	7.763	29	4.702	43	0.328	82
SE hybrid	7.359								

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

with a ratio of 0.2037 indicating dominance of non-additive gene action in inheritance of the trait (Table 22). Dominance variance was more than the additive variance for the trait. The results are in accordance with those obtained by Nayakar et al. (1989), Pillai et al. (1995), El-Mottaleb and Asran (2004), El-Menshawi (2005), Chaudhary et al. (2006), and Rani et al. (2015), while opposite results were reported by Tadesse et al. (2008).

The range of combined SCA effects for panicle length varied from -2.904 (A.Tx3197*RSC117-2 F_{2:3} high) to 2.724 (A.319*RSC15-13 F_{2:3} high). Among the hybrids, the greatest SCA effects were for A.319*RSC15-13 F_{2:3} high, A.319*RSC38-9 BC₁F_{2:3} low, and A.301*RSC19-10 BC₁F_{2:3} low considered desirable by virtue of their positive SCA effects. The three crosses A.Tx3197*RSC117-2 F_{2:3} high, A.338*RSC112-5 F_{2:3} high, and A.301*RSC19-10 BC₁F_{2:3} low had negative SCA effects for the trait (Table 25). Parents with high x low A.319 x RSC15-13 F_{2:3} high produced crosses with positive SCA effects, indicating additive x dominance type of gene action. The findings were in agreement with Patel et al. (1993) and Naik et al. (1994). In the cross, A.301*RSC19-10 BC₁F_{2:3} low, two poor combiners also resulted in large positive SCA effects that might be caused by high complementarity between the parents. This indicated that the parental combinations provided environments for full expression of genes controlling the trait, although the parents themselves would not express any superiority of the trait; accumulation of favorable genes might be the cause of parents with poor GCA effects producing hybrids with greater SCA effects. However, Premalatha et al. (2006) reported that the parental combination of low x low for GCA effects might be suitable for selection in later generations. Greater SCA variance (0.359)

Table 25. Specific Combining Ability (SCA) estimates for panicle length (cm) for each hybrid combination in the combined analysis across four environments with overall rank.

	Lines	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC†	Selections‡								
RSC73-9	F _{2:3} High	0.760	30	-0.122	90	-0.218	100	-0.420	118
RSC73-6	BC ₁ F _{2:3} High	0.534	41	-0.762	136	0.517	44	-0.288	105
RSC73-1	F _{2:3} Low	1.446	8	-1.018	143	0.180	62	-0.607	128
RSC73-5	BC ₁ F _{2:3} Low	-0.524	123	-0.757	135	0.335	52	0.946	22
RSC83-1	F _{2:3} High	-1.476	154	0.208	59	1.444	9	-0.176	93
RSC83-14	BC ₁ F _{2:3} High	-0.383	116	-0.032	78	0.017	74	0.398	47
RSC83-10	F _{2:3} Low	-0.061	80	-0.189	95	0.800	28	-0.550	124
RSC83-1	BC ₁ F _{2:3} Low	1.556	6	-1.076	146	0.206	60	-0.686	132
RSC112-5	F _{2:3} High	1.122	16	0.534	42	0.667	34	-2.324	159
RSC112-19	BC ₁ F _{2:3} High	1.371	11	-1.034	144	-0.589	126	0.251	56
RSC112-8	F _{2:3} Low	-0.185	94	-0.106	88	-0.513	121	0.804	27
RSC112-15	BC ₁ F _{2:3} Low	-0.359	112	-0.345	111	0.953	21	-0.249	103
RSC76-4	F _{2:3} High	-1.247	150	-0.335	110	0.297	53	1.284	14
RSC76-16	BC ₁ F _{2:3} High	-0.961	138	-0.195	98	0.294	54	0.862	25
RSC76-13	F _{2:3} Low	-0.523	122	-0.424	119	0.357	50	0.590	37
RSC76-2	BC ₁ F _{2:3} Low	-1.454	153	0.123	67	0.006	75	1.325	12
RSC38-5	F _{2:3} High	0.402	46	1.628	5	-0.006	76	-2.024	158
RSC38-15	BC ₁ F _{2:3} High	0.082	70	0.035	73	-0.749	133	0.633	35
RSC38-8	F _{2:3} Low	-0.284	104	-0.121	89	-0.091	84	0.496	45
RSC38-9	BC ₁ F _{2:3} Low	0.695	32	2.274	2	-1.008	142	-1.961	156
RSC37-12	F _{2:3} High	1.549	7	-0.975	139	-0.840	137	0.266	55
RSC37-12	BC ₁ F _{2:3} High	-1.383	152	0.926	23	-0.066	81	0.523	43
RSC37-7	F _{2:3} Low	0.128	65	-0.232	101	0.195	61	-0.091	85
RSC37-8	BC ₁ F _{2:3} Low	-1.008	141	1.113	18	0.038	72	-0.143	91
RSC15-13	F _{2:3} High	-1.814	155	2.724	1	-0.600	127	-0.311	107
RSC15-15	BC ₁ F _{2:3} High	-1.186	149	-0.377	115	0.133	64	1.429	10
RSC15-11	F _{2:3} Low	-0.194	96	-0.09	83	0.105	69	0.178	63
RSC15-14	BC ₁ F _{2:3} Low	-0.366	113	-1.997	157	1.074	19	1.289	13
RSC124-9	F _{2:3} High	-0.060	79	0.351	51	0.746	31	-1.037	145
RSC124-16	BC ₁ F _{2:3} High	0.208	58	0.056	71	-0.626	129	0.362	49
RSC124-3	F _{2:3} Low	0.565	39	0.811	26	-0.098	86	-1.278	151
RSC124-4	BC ₁ F _{2:3} Low	-0.020	77	1.114	17	-0.424	120	-0.669	131
RSC117-2	F _{2:3} High	1.756	4	0.128	66	-2.904	160	1.019	20
RSC117-4	BC ₁ F _{2:3} High	0.570	38	-0.371	114	0.554	40	-0.753	134
RSC117-10	F _{2:3} Low	-0.575	125	-0.245	102	-0.321	108	1.141	15
RSC117-3	BC ₁ F _{2:3} Low	-0.194	97	-0.158	92	0.246	57	0.106	68
RSC19-3	F _{2:3} High	-0.212	99	-0.069	82	-0.102	87	0.383	48
RSC19-17	BC ₁ F _{2:3} High	0.908	24	-1.116	147	0.621	36	-0.414	117
RSC19-1	F _{2:3} Low	-1.158	148	0.774	29	-0.300	106	0.684	33
RSC19-10	BC ₁ F _{2:3} Low	1.976	3	-0.653	130	-0.331	109	-0.992	140
SE hybrid	0.829								

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

than GCA variance (0.069) indicated dominance of non-additive gene action in inheritance of the trait (Table 22). Dominance variance was more than the additive variance for the trait, with a ratio (σ^2_D / σ^2_A) greater than unity. The results are in accordance with those obtained by Nayakar et al. (1989), Pillai et al. (1995), El-Mottaleb and Asran (2004), El-Menshawi (2005), and Chaudhary et al. (2006), but opposite the results reported by Tadesse et al. (2008).

For plant exsertion, the range of SCA effects varied from -4.600 (A.338*RSC15-11 F_{2:3} low) to 4.954 (A.338*RSC124-3 F_{2:3} low). Among the hybrids, A.338*RSC124-3 F_{2:3} low, A.301*RSC19-1 F_{2:3} low, A.Tx3197*RSC76-4 F_{2:3} high, and A.301*RSC37-8 BC₁F_{2:3} had positive numbers for plant exsertion, while A.338*RSC15-11 F_{2:3} low, A.Tx3197*RSC117-3 BC₁F_{2:3} low, and A.301*RSC76-2 BC₁F_{2:3} low had negative SCA effects (Table 26). All crosses involving low x high combination of GCA effects tended to produce the most positive SCA effects in the hybrids, indicating non-additive gene action, except for cross A.Tx3197*RSC76-4 F_{2:3} high that produced a high x high combination of GCA effects. The variance of GCA was slightly larger than the estimate of SCA variance, suggesting additive gene action (Table 22). The estimate of additive variance was slightly greater than the estimate for dominant variance. It is reasonable to suggest both additive and non-additive gene action.

For three-panicle weight, the range of SCA effects varied from -47.985 (A.338*RSC112-5 F_{2:3} high) to 50.282 (A.301*RSC112-5 F_{2:3} high) (Table 27). This signified the importance of testers. A.338 is a high general combiner making the cross high x high GCA effects produce the worst combination, whereas low x high GCA effects produced

Table 26. Specific Combining Ability (SCA) estimates for plant exsertion (cm) for each hybrid combination in the combined analysis across four environments with overall rank.

	Lines	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC†	Selections‡								
RSC73-9	F _{2:3} High	-2.208	140	-0.238	89	-0.459	93	2.904	12
RSC73-6	BC ₁ F _{2:3} High	1.113	46	0.807	61	-3.247	154	1.327	35
RSC73-1	F _{2:3} Low	1.244	39	-0.058	82	-2.156	139	0.970	53
RSC73-5	BC ₁ F _{2:3} Low	-0.780	105	0.066	80	0.564	67	0.151	77
RSC83-1	F _{2:3} High	-1.608	123	0.930	54	-0.517	95	1.195	43
RSC83-14	BC ₁ F _{2:3} High	0.922	55	-1.852	134	-2.227	142	3.157	10
RSC83-10	F _{2:3} Low	2.392	16	-2.781	150	-1.610	124	1.999	22
RSC83-1	BC ₁ F _{2:3} Low	1.263	38	-1.796	131	-0.498	94	1.032	49
RSC112-5	F _{2:3} High	-1.934	135	-1.797	132	1.144	45	2.588	15
RSC112-19	BC ₁ F _{2:3} High	-0.757	104	1.578	27	1.392	32	-2.213	141
RSC112-8	F _{2:3} Low	1.363	34	-0.848	109	1.574	28	-2.089	138
RSC112-15	BC ₁ F _{2:3} Low	-1.041	115	0.820	60	-0.561	99	0.783	62
RSC76-4	F _{2:3} High	-2.456	146	-0.088	84	4.233	3	-1.689	127
RSC76-16	BC ₁ F _{2:3} High	-2.987	152	0.824	58	3.728	6	-1.565	122
RSC76-13	F _{2:3} Low	1.219	42	1.045	48	-1.373	119	-0.892	110
RSC76-2	BC ₁ F _{2:3} Low	-4.085	158	3.698	7	0.177	76	0.210	75
RSC38-5	F _{2:3} High	0.970	52	-0.973	111	0.089	79	-0.086	83
RSC38-15	BC ₁ F _{2:3} High	-2.716	149	-0.523	97	3.861	5	-0.622	101
RSC38-8	F _{2:3} Low	3.227	9	-0.398	92	-1.007	112	-1.822	133
RSC38-9	BC ₁ F _{2:3} Low	1.184	44	0.113	78	-1.153	118	-0.144	88
RSC37-12	F _{2:3} High	0.773	63	-1.699	128	0.232	72	0.694	65
RSC37-12	BC ₁ F _{2:3} High	1.390	33	-4.013	157	1.534	29	1.089	47
RSC37-7	F _{2:3} Low	-1.099	117	1.934	23	0.823	59	-1.658	126
RSC37-8	BC ₁ F _{2:3} Low	3.939	4	-1.487	121	0.861	57	-3.313	155
RSC15-13	F _{2:3} High	-0.838	108	1.265	37	2.638	14	-3.065	153
RSC15-15	BC ₁ F _{2:3} High	0.259	71	-2.445	145	2.702	13	-0.517	96
RSC15-11	F _{2:3} Low	1.838	24	1.239	40	1.522	30	-4.600	160
RSC15-14	BC ₁ F _{2:3} Low	-0.143	87	0.722	64	-2.028	137	1.449	31
RSC124-9	F _{2:3} High	1.295	36	0.441	69	-1.963	136	0.227	73
RSC124-16	BC ₁ F _{2:3} High	2.004	21	-0.126	86	-2.436	144	0.558	68
RSC124-3	F _{2:3} Low	-2.886	151	-1.039	114	-1.029	113	4.954	1
RSC124-4	BC ₁ F _{2:3} Low	-3.873	156	0.862	56	-0.096	85	3.108	11
RSC117-2	F _{2:3} High	-0.728	103	-0.275	90	1.015	51	-0.012	81
RSC117-4	BC ₁ F _{2:3} High	-0.574	100	-0.815	107	2.081	18	-0.692	102
RSC117-10	F _{2:3} Low	-1.716	129	2.217	17	1.223	41	-1.724	130
RSC117-3	BC ₁ F _{2:3} Low	1.725	25	3.621	8	-4.565	159	-0.782	106
RSC19-3	F _{2:3} High	1.647	26	-0.529	98	-1.446	120	0.328	70
RSC19-17	BC ₁ F _{2:3} High	-2.675	148	2.025	20	1.031	50	-0.381	91
RSC19-1	F _{2:3} Low	4.696	2	-2.492	147	-2.427	143	0.223	74
RSC19-10	BC ₁ F _{2:3} Low	0.643	66	2.064	19	-1.627	125	-1.079	116
SE hybrid	2.182								

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

Table 27. Specific Combining Ability (SCA) estimates for three-panicle weight (g) for each hybrid combination in the combined analysis across four environments with overall rank.

RSC†	Lines Selections‡	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC73-9	F _{2:3} High	-22.118	147	-3.619	89	-1.516	81	27.253	11
RSC73-6	BC ₁ F _{2:3} High	-4.471	95	-26.710	151	-2.869	84	34.050	7
RSC73-1	F _{2:3} Low	-3.190	87	14.834	31	3.975	65	-15.619	129
RSC73-5	BC ₁ F _{2:3} Low	19.592	24	-17.860	135	-5.769	101	4.037	64
RSC83-1	F _{2:3} High	-19.193	139	-26.169	150	8.959	49	36.403	4
RSC83-14	BC ₁ F _{2:3} High	-19.455	140	7.231	53	26.759	12	-14.535	124
RSC83-10	F _{2:3} Low	5.948	58	10.284	46	-11.838	115	-4.394	92
RSC83-1	BC ₁ F _{2:3} Low	13.423	36	6.971	56	-3.575	88	-16.819	133
RSC112-5	F _{2:3} High	50.282	1	-4.757	97	2.459	70	-47.985	160
RSC112-19	BC ₁ F _{2:3} High	-19.630	141	38.314	3	-3.107	85	-15.576	128
RSC112-8	F _{2:3} Low	23.754	19	-8.297	109	-6.406	104	-9.050	111
RSC112-15	BC ₁ F _{2:3} Low	2.560	69	-27.629	152	7.062	55	18.006	25
RSC76-4	F _{2:3} High	10.826	44	11.099	43	-11.959	116	-9.966	113
RSC76-16	BC ₁ F _{2:3} High	16.217	26	12.078	40	-7.756	108	-20.538	143
RSC76-13	F _{2:3} Low	-4.130	90	23.931	18	-15.203	127	-4.597	96
RSC76-2	BC ₁ F _{2:3} Low	-21.255	146	14.268	32	-18.691	137	25.678	16
RSC38-5	F _{2:3} High	-15.077	125	-6.166	102	12.575	38	8.668	50
RSC38-15	BC ₁ F _{2:3} High	-1.171	79	4.490	62	2.244	73	-5.563	100
RSC38-8	F _{2:3} Low	-22.393	148	-6.707	105	-18.253	136	47.353	2
RSC38-9	BC ₁ F _{2:3} Low	25.779	14	13.553	35	-6.294	103	-33.038	157
RSC37-12	F _{2:3} High	-4.980	98	34.093	6	-12.953	119	-16.16	132
RSC37-12	BC ₁ F _{2:3} High	0.392	76	25.515	17	-12.594	117	-13.313	121
RSC37-7	F _{2:3} Low	-32.574	156	15.499	29	3.941	66	13.134	37
RSC37-8	BC ₁ F _{2:3} Low	-36.096	158	-15.685	130	20.894	22	30.887	8
RSC15-13	F _{2:3} High	8.571	51	13.982	33	-12.69	118	-9.863	112
RSC15-15	BC ₁ F _{2:3} High	-12.971	120	-11.197	114	-1.719	82	25.887	13
RSC15-11	F _{2:3} Low	7.284	52	-18.717	138	9.224	48	2.209	74
RSC15-14	BC ₁ F _{2:3} Low	10.288	45	-8.351	110	-27.659	153	25.722	15
RSC124-9	F _{2:3} High	5.900	59	-21.227	145	9.923	47	5.404	60
RSC124-16	BC ₁ F _{2:3} High	-6.975	106	21.978	20	-15.102	126	0.100	78
RSC124-3	F _{2:3} Low	2.445	71	-4.469	94	-1.403	80	3.428	67
RSC124-4	BC ₁ F _{2:3} Low	27.690	10	-15.769	131	5.155	61	-17.076	134
RSC117-2	F _{2:3} High	1.832	75	6.443	57	-5.166	99	-3.110	86
RSC117-4	BC ₁ F _{2:3} High	-14.083	123	-13.647	122	20.581	23	7.150	54
RSC117-10	F _{2:3} Low	-7.753	107	4.437	63	0.212	77	3.105	68
RSC117-3	BC ₁ F _{2:3} Low	-22.758	149	28.290	9	15.644	27	-21.175	144
RSC19-3	F _{2:3} High	-4.290	91	-32.516	155	15.612	28	21.193	21
RSC19-17	BC ₁ F _{2:3} High	14.851	30	-30.751	154	13.616	34	2.284	72
RSC19-1	F _{2:3} Low	12.426	39	-4.401	93	11.778	42	-19.803	142
RSC19-10	BC ₁ F _{2:3} Low	34.507	5	-2.644	83	11.909	41	-43.772	159
SE hybrid	17.739								

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

the best combination. Among the hybrids, three crosses A.301*RSC112-5 F_{2:3} high, A.338*RSC38-8 BC₁F_{2:3} low, and A.319*RSC112-19 BC₁F_{2:3} high were considered desirable by virtue of their positive SCA effects, and four crosses A.338* RSC112-5 F_{2:3} high, A.338*RSC19-10 BC₁F_{2:3} low, A.301*RSC37-8 BC₁F_{2:3} low, and A.338*RSC38-9 BC₁F_{2:3} low had negative SCA effects. Parents with a low x high (A.301*RSC112-5 F_{2:3} high) GCA effect produced crosses with greatest SCA effects, indicating additive x dominance type of gene interaction. The findings were in agreement with those by Patel et al. (1993) and Naik et al. (1994). In the crosses A.338*RSC38-8 BC₁F_{2:3} low, and A.319*RSC112-19 BC₁F_{2:3} high, two good combiners also resulted in large positive SCA effects that might be caused by high complementarity between the parents. According to Premalatha et al. (2006), the hybrids might be desirable for biparental selection or inter-mating. Results indicated the need to exploit hybrid vigor to enhance levels of yield. For three-panicle weight, the ratio of GCA to SCA variances was less than unity, indicating the predominant role of non-additive gene action. This was in accordance with earlier reports by Badhe and Patel (1997), Siddiqui and Baig (2001), Kulakarni et al. (2006), and Aruna et al. (2010). Influence of non-additive gene action in controlling the trait also was indicated by the degree of dominance (Table 22).

For 1000-kernel weight, SCA effects ranged from -3.061 (A.338*RSC112-19 BCF_{2:3} high) to 3.185 (A.Tx3197*RSC117-2 F_{2:3} high). Of the 160 crosses, three crosses A.Tx3197*RSC117-2 F_{2:3} high, A.319*RSC112-19 BC₁F_{2:3}, and A.338*RSC73-6 BC₁F_{2:3} had large positive SCA effects, and four crosses A.338*RSC112-19 BC₁F_{2:3} high, A.Tx3197*RSC76-13 F_{2:3} low, A.338*RSC112-8 F_{2:3} low, and A.301*RSC37-7

F_{2:3} low had negative SCA effects (Table 28). The hybrid A.Tx3197*RSC117-2 F_{2:3} high expressed the greatest positive SCA effect and also showed positive average heterosis. One of the other two hybrids that had positive SCA effects was a combination of high x low (A.338*RSC73-6 BC₁F_{2:3}) GCA effects of parents, indicating non-additive gene action. The other one that had positive SCA effects was a combination low x low (A.319*RSC112-19 BC₁F_{2:3}), as previously observed; two poor combiners also resulted in large positive SCA effects that might have been caused by great complementarity between the parents. According to Premalatha et al. (2006), the hybrid should be delayed until future generations. This indicated that the parental combinations provided an environment for full expression of genes controlling the trait, although the parents themselves would not express any superiority for the trait. Accumulation of favorable genes might be the cause of parents with poor GCA giving rise to hybrids with greater SCA effects. Results indicated the need to exploit hybrid vigor to enhance greater yield potential. For 1000-kernel weight, the estimate of SCA variance was greater than that of the GCA variance (0.115), with a ratio of 0.258, indicating the importance of non-additive gene action in inheritance of the trait (Table 22). Dominance variance was greater than the additive variance for the trait. The results conformed to earlier reports by Badhe and Patil (1997), El-Menshawhi (2005), Chaudhary et al. (2006), and Premalatha et al. (2006). In opposition to results obtained in the present study, only additive gene action as reported by Mahdy et al. (2011) and both additive and non-additive gene action reported by Salini et al. (2008) influenced 1000-kernel weight.

Table 28. Specific Combining Ability (SCA) estimates for 1000-kernel weight (g) for each hybrid combination in the combined analysis across four environments with overall rank.

RSC†	Lines Selections‡	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC73-9	F _{2:3} High	-1.913	154	-0.634	118	0.639	42	1.908	8
RSC73-6	BC ₁ F _{2:3} High	-1.327	144	-0.951	133	-0.242	94	2.521	3
RSC73-1	F _{2:3} Low	-1.031	136	0.659	41	-0.167	89	0.540	46
RSC73-5	BC ₁ F _{2:3} Low	0.788	37	0.727	38	-0.372	102	-1.144	140
RSC83-1	F _{2:3} High	-1.034	137	0.144	67	0.691	40	0.199	62
RSC83-14	BC ₁ F _{2:3} High	-0.020	78	-0.847	128	1.159	25	-0.293	97
RSC83-10	F _{2:3} Low	0.552	44	-1.504	148	0.810	35	0.142	68
RSC83-1	BC ₁ F _{2:3} Low	-0.021	79	0.104	71	1.018	28	-1.100	139
RSC112-5	F _{2:3} High	-1.815	153	1.376	18	-0.969	135	1.408	17
RSC112-19	BC ₁ F _{2:3} High	0.067	75	2.626	2	0.368	57	-3.061	160
RSC112-8	F _{2:3} Low	0.939	29	1.250	22	-0.067	81	-2.122	158
RSC112-15	BC ₁ F _{2:3} Low	2.378	4	1.131	26	-1.437	146	-2.072	155
RSC76-4	F _{2:3} High	2.022	7	-0.078	83	-1.471	147	-0.473	110
RSC76-16	BC ₁ F _{2:3} High	1.510	13	0.385	55	-1.787	152	-0.107	85
RSC76-13	F _{2:3} Low	0.374	56	2.058	5	-2.301	159	-0.131	87
RSC76-2	BC ₁ F _{2:3} Low	1.842	9	-0.824	127	-0.203	92	-0.815	126
RSC38-5	F _{2:3} High	-0.886	129	-1.358	145	0.190	63	2.054	6
RSC38-15	BC ₁ F _{2:3} High	-0.586	112	-0.115	86	0.542	45	0.159	66
RSC38-8	F _{2:3} Low	-0.360	101	-0.408	103	0.518	48	0.251	59
RSC38-9	BC ₁ F _{2:3} Low	-0.591	113	-0.534	111	0.490	49	0.635	43
RSC37-12	F _{2:3} High	-0.135	88	1.504	14	-0.771	124	-0.598	115
RSC37-12	BC ₁ F _{2:3} High	1.365	19	-0.964	134	0.525	47	-0.927	132
RSC37-7	F _{2:3} Low	-2.115	157	0.186	64	1.754	10	0.175	65
RSC37-8	BC ₁ F _{2:3} Low	0.453	51	-0.438	104	-0.447	106	0.432	53
RSC15-13	F _{2:3} High	-0.718	121	-0.456	108	-0.303	99	1.477	15
RSC15-15	BC ₁ F _{2:3} High	-0.261	95	-0.613	117	0.807	36	0.067	76
RSC15-11	F _{2:3} Low	0.119	69	-0.765	123	-0.262	96	0.907	31
RSC15-14	BC ₁ F _{2:3} Low	-0.656	119	1.164	24	-0.603	116	0.094	72
RSC124-9	F _{2:3} High	1.476	16	0.206	61	-1.645	151	-0.037	80
RSC124-16	BC ₁ F _{2:3} High	-0.807	125	1.562	12	-1.570	150	0.815	34
RSC124-3	F _{2:3} Low	1.619	11	-0.444	105	-0.469	109	-0.706	120
RSC124-4	BC ₁ F _{2:3} Low	1.164	23	-2.102	156	-0.314	100	1.252	21
RSC117-2	F _{2:3} High	-0.592	114	-1.321	143	3.185	1	-1.272	142
RSC117-4	BC ₁ F _{2:3} High	-1.257	141	-0.203	91	1.350	20	0.110	70
RSC117-10	F _{2:3} Low	0.883	32	-0.763	122	0.088	73	-0.208	93
RSC117-3	BC ₁ F _{2:3} Low	0.084	74	-0.086	84	-0.454	107	0.456	50
RSC19-3	F _{2:3} High	-1.050	138	-0.300	98	0.433	52	0.918	30
RSC19-17	BC ₁ F _{2:3} High	0.387	54	0.829	33	0.294	58	-1.510	149
RSC19-1	F _{2:3} Low	0.044	77	-0.908	131	1.059	27	-0.195	90
RSC19-10	BC ₁ F _{2:3} Low	-0.889	130	0.708	39	-0.068	82	0.249	60
SE hybrid	1.048								

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

GCA effects of protein content in grain ranged from -0.887 (A.Tx3197*RSC37-12 BC₁F_{2:3} high) to 0.961 (A.319*RSC37-12 BC₁F_{2:3} high) (Table 29). Of the 160 crosses, A.319*RSC37-12 BC₁F_{2:3} high, A.338*RSC73-9 F_{2:3} high, and A.319*RSC19-17 BC₁F_{2:3} high had positive SCA effects and were identified as good specific combiners for the trait. Of the crosses, A.319 was the only good general combiner with high x high combination of GCA effects. It involved good general combiners as parents and can be used for isolating promising sorghum material in later generations for development of superior varieties (Rao, 1970). The least specific combiners were A.Tx319* RSC37-12 BC₁F_{2:3} high, A.Tx3197*RSC73-9 F_{2:3} high, and A.Tx319*RSC117-3 BC₁F_{2:3} low, all of which had negative SCA effects. For protein content in grain, the estimate of SCA variance was less than that of the GCA variance, indicating dominance of additive gene action (Table 22). Additive variance was greater than the dominance variance for the trait. But, dominance of both additive and non-additive gene action in inheritance of the trait was reported by Govil and Murty (1973). Results of the present study with additive variance being greater than dominance were opposite those of Rani et al. (2015).

Starch content of grain ranged from -0.67 (A.301*RSC112-5 BC₁F_{2:3} high) to 0.668 (A.301*RSC73-1 F_{2:3} low). Of the hybrids, A.301*RSC73-1 F_{2:3} low, A.301*RSC38-9 BC₁F_{2:3} low, A.338*RSC124-9 BC₁F_{2:3}, and A.301*RSC73-9 F_{2:3} high had the greatest positive SCA effects, while A.301*RSC112-5 BC₁F_{2:3} high, A.301*RSC124-16 BC₁F_{2:3} high, and A.338*RSC73-9 F_{2:3} high had negative SCA effects (Table 30). The greatest positive SCA effects were by the cross A.301*RSC73-1 F_{2:3} low, followed by A.301*RSC38-9 BC₁F_{2:3} low. For starch content in grain, the

Table 29. Specific Combining Ability (SCA) estimates for concentration of protein in grain (%) for each hybrid combination in the combined analysis across four environments with overall rank.

	Lines	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC†	Selections‡								
RSC73-9	F _{2:3} High	-0.593	158	0.454	13	-0.612	159	0.751	2
RSC73-6	BC ₁ F _{2:3} High	-0.077	98	-0.036	86	-0.092	102	0.206	35
RSC73-1	F _{2:3} Low	-0.509	156	0.239	28	-0.189	125	0.459	12
RSC73-5	BC ₁ F _{2:3} Low	0.137	45	-0.041	88	-0.041	89	-0.055	91
RSC83-1	F _{2:3} High	0.234	29	-0.172	121	0.331	20	-0.393	151
RSC83-14	BC ₁ F _{2:3} High	-0.016	75	-0.106	108	0.153	44	-0.032	84
RSC83-10	F _{2:3} Low	0.058	55	-0.015	73	0.227	31	-0.269	138
RSC83-1	BC ₁ F _{2:3} Low	-0.233	134	-0.144	113	0.535	7	-0.158	116
RSC112-5	F _{2:3} High	-0.039	87	-0.176	122	-0.006	67	0.221	34
RSC112-19	BC ₁ F _{2:3} High	0.325	21	0.173	42	-0.311	143	-0.187	124
RSC112-8	F _{2:3} Low	0.226	33	-0.024	81	0.086	52	-0.287	141
RSC112-15	BC ₁ F _{2:3} Low	0.555	5	-0.034	85	-0.060	92	-0.462	154
RSC76-4	F _{2:3} High	-0.275	139	-0.163	117	0.539	6	-0.101	107
RSC76-16	BC ₁ F _{2:3} High	-0.501	155	-0.009	69	0.284	25	0.227	32
RSC76-13	F _{2:3} Low	-0.374	149	0.229	30	-0.127	110	0.271	26
RSC76-2	BC ₁ F _{2:3} Low	-0.195	126	-0.208	128	0.419	16	-0.016	76
RSC38-5	F _{2:3} High	-0.152	114	-0.044	90	0.171	43	0.025	62
RSC38-15	BC ₁ F _{2:3} High	0.136	48	-0.169	119	0.202	36	-0.169	120
RSC38-8	F _{2:3} Low	-0.356	146	-0.265	137	0.522	8	0.099	51
RSC38-9	BC ₁ F _{2:3} Low	-0.010	71	0.027	60	-0.336	144	0.319	23
RSC37-12	F _{2:3} High	-0.135	111	0.137	47	-0.183	123	0.181	39
RSC37-12	BC ₁ F _{2:3} High	0.137	46	0.961	1	-0.887	160	-0.212	129
RSC37-7	F _{2:3} Low	-0.075	97	-0.007	68	0.065	53	0.018	63
RSC37-8	BC ₁ F _{2:3} Low	-0.093	103	0.393	17	-0.227	133	-0.074	96
RSC15-13	F _{2:3} High	0.025	61	-0.224	131	0.367	18	-0.168	118
RSC15-15	BC ₁ F _{2:3} High	-0.023	80	-0.439	153	-0.019	77	0.481	9
RSC15-11	F _{2:3} Low	0.419	15	-0.225	132	-0.068	94	-0.125	109
RSC15-14	BC ₁ F _{2:3} Low	0.062	54	-0.277	140	0.180	40	0.035	58
RSC124-9	F _{2:3} High	0.285	24	-0.243	136	0.325	22	-0.366	147
RSC124-16	BC ₁ F _{2:3} High	0.461	11	-0.233	135	-0.072	95	-0.157	115
RSC124-3	F _{2:3} Low	0.004	65	0.100	50	-0.011	72	-0.094	104
RSC124-4	BC ₁ F _{2:3} Low	0.469	10	0.008	64	-0.097	105	-0.380	150
RSC117-2	F _{2:3} High	-0.101	106	-0.009	70	0.195	38	-0.086	100
RSC117-4	BC ₁ F _{2:3} High	0.450	14	-0.308	142	-0.342	145	0.200	37
RSC117-10	F _{2:3} Low	-0.065	93	0.051	56	0.030	59	-0.015	74
RSC117-3	BC ₁ F _{2:3} Low	0.586	4	-0.026	83	-0.562	157	0.001	66
RSC19-3	F _{2:3} High	-0.088	101	0.178	41	-0.217	130	0.127	49
RSC19-17	BC ₁ F _{2:3} High	-0.084	99	0.620	3	-0.398	152	-0.138	112
RSC19-1	F _{2:3} Low	-0.201	127	-0.024	82	0.245	27	-0.020	78
RSC19-10	BC ₁ F _{2:3} Low	-0.372	148	0.051	57	-0.022	79	0.342	19
SE hybrid	0.333								

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

Table 30. Specific Combining Ability (SCA) estimates for concentration for starch in grain (%) for each hybrid combination in the combined analysis across four environments with overall rank.

RSC†	Lines Selections‡	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC73-9	F _{2:3} High	0.541	4	-0.206	123	0.224	31	-0.558	158
RSC73-6	BC ₁ F _{2:3} High	0.226	29	-0.079	107	0.091	63	-0.237	126
RSC73-1	F _{2:3} Low	0.668	1	-0.250	127	0.098	60	-0.516	153
RSC73-5	BC ₁ F _{2:3} Low	0.125	51	-0.319	136	0.402	15	-0.208	124
RSC83-1	F _{2:3} High	-0.361	142	0.253	23	-0.337	138	0.445	11
RSC83-14	BC ₁ F _{2:3} High	-0.096	110	-0.047	96	0.048	72	0.096	61
RSC83-10	F _{2:3} Low	0.004	83	0.091	62	-0.418	150	0.323	20
RSC83-1	BC ₁ F _{2:3} Low	0.104	56	-0.152	114	-0.054	99	0.101	59
RSC112-5	F _{2:3} High	-0.198	120	0.037	76	0.210	34	-0.049	98
RSC112-19	BC ₁ F _{2:3} High	-0.489	151	-0.070	103	0.535	5	0.025	79
RSC112-8	F _{2:3} Low	-0.132	113	-0.075	105	0.047	73	0.160	43
RSC112-15	BC ₁ F _{2:3} Low	-0.670	160	0.101	57	0.227	28	0.342	18
RSC76-4	F _{2:3} High	0.322	21	-0.036	94	-0.294	133	0.008	82
RSC76-16	BC ₁ F _{2:3} High	0.462	10	-0.263	129	0.138	48	-0.337	140
RSC76-13	F _{2:3} Low	0.441	12	-0.203	122	0.060	69	-0.297	135
RSC76-2	BC ₁ F _{2:3} Low	0.115	55	-0.209	125	0.203	37	-0.109	111
RSC38-5	F _{2:3} High	0.155	45	0.195	40	-0.337	139	-0.013	87
RSC38-15	BC ₁ F _{2:3} High	0.066	67	-0.010	86	-0.175	115	0.120	54
RSC38-8	F _{2:3} Low	0.496	6	0.137	49	-0.368	146	-0.265	130
RSC38-9	BC ₁ F _{2:3} Low	0.597	2	-0.268	131	0.087	64	-0.415	149
RSC37-12	F _{2:3} High	-0.014	88	-0.072	104	0.062	68	0.024	81
RSC37-12	BC ₁ F _{2:3} High	-0.365	143	-0.331	137	0.209	35	0.488	7
RSC37-7	F _{2:3} Low	-0.115	112	0.185	41	-0.294	134	0.224	32
RSC37-8	BC ₁ F _{2:3} Low	-0.031	92	-0.368	145	0.245	25	0.153	46
RSC15-13	F _{2:3} High	0.042	74	0.480	9	-0.547	157	0.025	80
RSC15-15	BC ₁ F _{2:3} High	0.050	71	0.347	16	0.144	47	-0.541	156
RSC15-11	F _{2:3} Low	-0.367	144	0.416	13	-0.077	106	0.029	77
RSC15-14	BC ₁ F _{2:3} Low	0.135	50	0.177	42	-0.054	100	-0.258	128
RSC124-9	F _{2:3} High	-0.513	152	0.339	19	-0.400	147	0.574	3
RSC124-16	BC ₁ F _{2:3} High	-0.564	159	0.201	38	0.205	36	0.158	44
RSC124-3	F _{2:3} Low	0.002	84	-0.056	101	0.002	85	0.053	70
RSC124-4	BC ₁ F _{2:3} Low	-0.198	121	-0.093	109	0.041	75	0.250	24
RSC117-2	F _{2:3} High	-0.032	93	0.232	27	-0.180	116	-0.019	90
RSC117-4	BC ₁ F _{2:3} High	-0.402	148	0.483	8	0.101	58	-0.182	117
RSC117-10	F _{2:3} Low	-0.027	91	-0.079	108	0.026	78	0.079	65
RSC117-3	BC ₁ F _{2:3} Low	-0.530	155	0.244	26	0.347	17	-0.062	102
RSC19-3	F _{2:3} High	0.121	53	-0.280	132	0.198	39	-0.039	95
RSC19-17	BC ₁ F _{2:3} High	-0.048	97	-0.345	141	-0.014	89	0.408	14
RSC19-1	F _{2:3} Low	0.226	30	0.077	66	-0.520	154	0.218	33
RSC19-10	BC ₁ F _{2:3} Low	0.254	22	-0.183	118	0.123	52	-0.195	119
SE hybrid	0.321								

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

estimate of SCA variance was less than the GCA variance, indicating dominance of additive gene action (Table 22). All cross combinations involving bad general combiners tended to produce the high positive SCA effects in the hybrids, which might be caused by good complementarity between parents, except in the cross A.301*RSC38-9 BC₁F_{2:3} low that had low x high combination of GCA effects. Results of the present study with additive variance being greater than dominance were opposite those of Rani et al. (2015).

Fiber content of grain ranged from -0.049 (A.Tx3197*RSC112-19 BC₁F_{2:3} high) to 0.067 (A.301*RSC112-19 BC₁F_{2:3} high) (Table 31). The crosses were low x high and high x high, respectively, for the combination of GCA effects. Although a very small difference, SCA variance was a smaller estimate than GCA variance, indicating dominance of additive gene action (Table 22). The action of good general combiners produced the most positive hybrid for fiber content of grain.

Fat content of grain ranged from -0.283 (A.319*RSC117-4 BCF_{2:3} high) to 0.303 (A.338*RSC73-9 F_{2:3} high) (Table 32). The crosses were low x high and high x low, respectively, for combination of GCA effects, indicating non-additive gene action; however, the estimate for GCA variance was slightly larger than the estimate for SCA variance, indicating dominance of additive gene action (Table 22). The predominance of both additive and non-additive gene action was evident.

Genetic Components

Data in Table 22 show the genetic components and contribution of the lines, testers, and their interaction for all studied traits in all environments. The line x tester analysis revealed that the contribution of the lines to the total sum of squares was greater

Table 31. Specific Combining Ability (SCA) estimates for concentration of fiber in grain (%) for each hybrid combination in the combined analysis across four environments with overall rank.

Lines		A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC†	Selections‡								
RSC73-9	F _{2:3} High	-0.025	141	0.027	17	-0.025	143	0.024	22
RSC73-6	BC ₁ F _{2:3} High	-0.023	138	0.005	58	0.013	45	0.006	57
RSC73-1	F _{2:3} Low	-0.036	155	0.01	52	0.002	72	0.024	23
RSC73-5	BC ₁ F _{2:3} Low	-0.034	153	-0.002	81	0.024	21	0.012	48
RSC83-1	F _{2:3} High	-0.015	113	0.005	59	0.027	18	-0.017	124
RSC83-14	BC ₁ F _{2:3} High	0.040	8	-0.017	122	0.015	40	-0.039	157
RSC83-10	F _{2:3} Low	-0.009	100	-0.019	129	0.018	34	0.010	54
RSC83-1	BC ₁ F _{2:3} Low	0.017	35	-0.001	79	-0.015	114	0.000	78
RSC112-5	F _{2:3} High	0.002	68	0.037	9	-0.02	134	-0.019	130
RSC112-19	BC ₁ F _{2:3} High	0.067	1	-0.004	87	-0.049	160	-0.014	112
RSC112-8	F _{2:3} Low	0.015	38	-0.012	107	0.003	66	-0.006	93
RSC112-15	BC ₁ F _{2:3} Low	0.016	36	-0.012	108	-0.003	85	-0.001	80
RSC76-4	F _{2:3} High	-0.018	125	0.002	70	0.015	41	0.001	77
RSC76-16	BC ₁ F _{2:3} High	-0.044	159	0.02	29	-0.013	111	0.037	10
RSC76-13	F _{2:3} Low	-0.024	140	0.019	30	-0.015	115	0.019	32
RSC76-2	BC ₁ F _{2:3} Low	-0.029	146	0.005	60	-0.004	89	0.028	16
RSC38-5	F _{2:3} High	0.002	69	0.001	75	-0.015	116	0.012	49
RSC38-15	BC ₁ F _{2:3} High	0.026	19	-0.017	123	0.003	67	-0.012	109
RSC38-8	F _{2:3} Low	-0.029	147	-0.003	83	0.042	7	-0.010	103
RSC38-9	BC ₁ F _{2:3} Low	-0.020	132	0.004	63	0.022	25	-0.006	94
RSC37-12	F _{2:3} High	-0.012	105	-0.009	101	0.030	13	-0.009	102
RSC37-12	BC ₁ F _{2:3} High	-0.035	154	0.018	33	0.005	61	0.012	50
RSC37-7	F _{2:3} Low	-0.023	139	0.019	31	0.022	26	-0.019	131
RSC37-8	BC ₁ F _{2:3} Low	-0.003	82	0.001	76	-0.011	104	0.013	46
RSC15-13	F _{2:3} High	-0.012	106	-0.018	127	-0.016	120	0.047	6
RSC15-15	BC ₁ F _{2:3} High	0.021	27	0.002	71	-0.005	91	-0.018	128
RSC15-11	F _{2:3} Low	-0.025	142	0.004	64	-0.030	151	0.050	4
RSC15-14	BC ₁ F _{2:3} Low	0.054	3	-0.029	148	-0.003	86	-0.022	136
RSC124-9	F _{2:3} High	0.013	44	0.016	37	-0.031	152	0.002	74
RSC124-16	BC ₁ F _{2:3} High	0.047	5	-0.013	110	-0.029	149	-0.005	92
RSC124-3	F _{2:3} Low	-0.017	121	0.009	55	-0.015	117	0.023	24
RSC124-4	BC ₁ F _{2:3} Low	0.036	11	0.011	51	-0.026	144	-0.022	137
RSC117-2	F _{2:3} High	0.035	12	-0.008	96	0.015	42	-0.042	158
RSC117-4	BC ₁ F _{2:3} High	0.026	20	-0.003	84	-0.015	118	-0.008	98
RSC117-10	F _{2:3} Low	0.020	28	-0.028	145	0.028	14	-0.020	135
RSC117-3	BC ₁ F _{2:3} Low	0.057	2	-0.03	150	0.012	47	-0.038	156
RSC19-3	F _{2:3} High	-0.020	133	0.004	65	0.002	73	0.014	43
RSC19-17	BC ₁ F _{2:3} High	-0.018	126	0.015	39	0.010	53	-0.008	99
RSC19-1	F _{2:3} Low	-0.016	119	-0.008	97	0.028	15	-0.004	90
RSC19-10	BC ₁ F _{2:3} Low	-0.007	95	-0.004	88	0.006	56	0.005	62
SE hybrid	0.033								

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

Table 32. Specific Combining Ability (SCA) estimates for concentration of fat in grain (%) for each hybrid combination in the combined analysis across four environments with overall rank.

Lines		A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC†	Selections‡								
RSC73-9	F _{2:3} High	-0.236	158	0.208	4	-0.275	159	0.303	1
RSC73-6	BC ₁ F _{2:3} High	0.011	81	-0.077	117	0.027	73	0.039	63
RSC73-1	F _{2:3} Low	-0.174	145	0.109	31	-0.229	156	0.294	2
RSC73-5	BC ₁ F _{2:3} Low	0.037	65	-0.043	102	-0.042	100	0.048	56
RSC83-1	F _{2:3} High	-0.052	107	0.117	29	0.036	67	-0.101	125
RSC83-14	BC ₁ F _{2:3} High	-0.088	120	0.039	62	0.082	40	-0.033	98
RSC83-10	F _{2:3} Low	-0.116	129	0.053	54	0.179	9	-0.116	130
RSC83-1	BC ₁ F _{2:3} Low	0.027	70	0.028	69	0.056	53	-0.111	127
RSC112-5	F _{2:3} High	0.130	25	0.125	27	-0.232	157	-0.023	92
RSC112-19	BC ₁ F _{2:3} High	0.158	17	-0.044	105	0.003	84	-0.116	131
RSC112-8	F _{2:3} Low	-0.019	91	0.077	42	-0.145	140	0.087	38
RSC112-15	BC ₁ F _{2:3} Low	0.197	6	-0.024	93	0.025	75	-0.198	150
RSC76-4	F _{2:3} High	0.014	77	0.042	58	0.079	41	-0.135	138
RSC76-16	BC ₁ F _{2:3} High	-0.080	118	0.027	72	-0.126	134	0.179	10
RSC76-13	F _{2:3} Low	-0.128	136	0.095	37	-0.146	141	0.179	11
RSC76-2	BC ₁ F _{2:3} Low	0.027	71	-0.043	103	-0.045	106	0.061	50
RSC38-5	F _{2:3} High	0.014	78	-0.089	122	0.127	26	-0.052	108
RSC38-15	BC ₁ F _{2:3} High	0.003	83	0.07	47	0.040	59	-0.112	128
RSC38-8	F _{2:3} Low	-0.096	124	-0.014	89	0.072	45	0.038	64
RSC38-9	BC ₁ F _{2:3} Low	0.000	85	0.036	66	0.026	74	-0.061	111
RSC37-12	F _{2:3} High	-0.009	86	0.144	20	-0.075	115	-0.060	110
RSC37-12	BC ₁ F _{2:3} High	-0.118	132	0.261	3	-0.204	152	0.061	51
RSC37-7	F _{2:3} Low	0.169	15	-0.127	135	0.157	18	-0.199	151
RSC37-8	BC ₁ F _{2:3} Low	-0.106	126	0.166	16	-0.036	99	-0.025	94
RSC15-13	F _{2:3} High	0.039	61	-0.129	137	0.151	19	-0.061	112
RSC15-15	BC ₁ F _{2:3} High	0.070	46	-0.176	146	0.070	48	0.036	68
RSC15-11	F _{2:3} Low	0.083	39	-0.028	95	0.104	36	-0.159	144
RSC15-14	BC ₁ F _{2:3} Low	0.072	44	-0.21	154	0.170	14	-0.032	97
RSC124-9	F _{2:3} High	0.133	23	-0.182	147	0.196	7	-0.147	142
RSC124-16	BC ₁ F _{2:3} High	0.180	8	-0.206	153	0.067	49	-0.042	101
RSC124-3	F _{2:3} Low	-0.076	116	-0.014	90	-0.081	119	0.171	13
RSC124-4	BC ₁ F _{2:3} Low	0.109	30	-0.125	133	0.202	5	-0.186	148
RSC117-2	F _{2:3} High	-0.075	114	0.109	32	-0.140	139	0.106	33
RSC117-4	BC ₁ F _{2:3} High	0.105	34	-0.283	160	0.040	60	0.139	22
RSC117-10	F _{2:3} Low	0.012	80	0.014	79	-0.088	121	0.061	52
RSC117-3	BC ₁ F _{2:3} Low	0.074	43	-0.194	149	0.133	24	-0.012	88
RSC19-3	F _{2:3} High	-0.090	123	0.173	12	-0.226	155	0.143	21
RSC19-17	BC ₁ F _{2:3} High	-0.060	109	0.104	35	-0.067	113	0.023	76
RSC19-1	F _{2:3} Low	-0.149	143	0.053	55	0.125	28	-0.029	96
RSC19-10	BC ₁ F _{2:3} Low	0.009	82	-0.043	104	-0.010	87	0.044	57
SE hybrid	0.182								

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

than that of the testers for all studied traits except the number of days to anthesis. The maximum contribution of lines (69.5%) was noted for panicle length, followed by 1000-kernel weight (69.3%), while the lowest values were for the number of days to anthesis (12.7%), followed by fiber concentration in grain (44.9%). Testers contributed a maximum for the number of days to anthesis (73.8%) and least for starch concentration in grain (1.6%). For the line x tester interaction, the maximum contribution to the total sum of squares was 49.2% for starch concentration in grain, whereas the minimum contribution was 7.1% for the height to the flag leaf. Variance due to lines was greater than that of testers for all traits except the number of days to anthesis, which is confirmed by the contributions of lines and testers. By removing NIR data because of observed negative SCA variances, the variance due to GCA (σ^2_{gca}) was less than the variance for SCA (σ^2_{sca}) for all traits except plant exsertion ($\sigma^2_{gca}/\sigma^2_{sca} = 1.5122$) and concentration of protein, starch, fiber, and fat in grain, suggesting that a preponderance of non-additive gene action controlled the characters. Dominance variance (σ^2_D) was large while additive variance (σ^2_A) was less in magnitude for all traits except plant exsertion and concentration of protein, starch, fiber, and fat in grain. The results are supported by the ratio of variance of general to specific combining ability ($\sigma^2_{gca}/\sigma^2_{sca}$) that was less than unity and by the degree of dominance (σ^2_D/σ^2_A) that takes values greater than unity for all traits except plant exsertion where σ^2_A was greater than σ^2_D with the degree of dominance being less than unity. Similar results were found by Fellahi et al. (2013) who suggested that with the preponderance of non-additive gene action, in terms of grain yield, plant height, and duration of the vegetative growth, use of sorghum varieties should be postponed until later generations. Mohammed (2009) reported that additive gene action

was important in the expression of days to flower and forage yield while non-additive gene action was important in plant height where $\sigma^2_{\text{gca}}/\sigma^2_{\text{sca}}$ was less than unity. Mahdy et al. (2011) found that both additive and non-additive gene action were important for inheritance of plant height and grain yield, and they found the additive effect controlled days to anthesis. Several researchers indicated the importance of additive and non-additive gene action in inheritance of grain yield and some agronomic traits (Kenga et al., 2004; Abdel-Mottaleb, 2009; Mohammed, 2009; Mahday et al., 2011).

Midparent Heterosis

Exploitation of hybrid vigor is an appropriate alternative for making further breakthroughs in increasing sorghum yield. Greater yield over high-yielding check varieties and wider adaptability have been instrumental in the rapid spread of hybrid sorghum. Estimates of degree of dominance were greater than the degree of additive for grain yield and yield components except plant exsertion and protein, starch, fiber, and fat contents in grain, which indicates dominance is the primary cause of heterosis (Table 33).

The range of MPH for grain yield was -34.22 (A.338*RSC76-2 BC₁F_{2:3} low) to 61.76% (A.301*RSC83-14 BC₁F_{2:3} high) (Table 33). As many as 31 hybrids expressed significant positive MPH ranging from 32.45 (A.301*RSC38-9 BC₁F_{2:3} low) to 61.76% (A.301*RSC83-14 BC₁F_{2:3} high). Of the 160 hybrids, only two expressed significant negative MPH ranging from -34.22 (A.338*RSC76-2 BC₁F_{2:3} low) to -33.62% (A.338*RSC76-16 BC₁F_{2:3} high). The highest significant positive MPH larger than 50% was among hybrids A.301*RSC83-14 BC₁F_{2:3} high, A.301*RSC124-9 F_{2:3} high, A.319*RSC83-1 BC₁F_{2:3} low, and A.301*RSC76-4 F_{2:3} high. Of the 40 lines, three

Table 33. Midparent heterosis estimates for grain yield (Mgha⁻¹) for each hybrid combination in the combined analysis in four environments, Vega, TX (2015 and 2016), Hutchinson, KS (2015), and Dumas, TX (2016) with overall rank.

Lines		A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC†	Selections‡								
RSC73-9	F _{2:3} High	45.09**	9	18.00	68	5.91	107	-12.51	142
RSC73-6	BC ₁ F _{2:3} High	21.78	58	22.46	54	-14.36	148	-15.18	151
RSC73-1	F _{2:3} Low	15.36	78	8.24	100	-1.41	122	-5.48	131
RSC73-5	BC ₁ F _{2:3} Low	34.63*	24	22.05	56	-8.61	136	-14.33	147
RSC83-1	F _{2:3} High	35.81*	20	14.61	80	-6.30	133	15.01	79
RSC83-14	BC ₁ F _{2:3} High	61.76**	1	35.38*	21	18.60	65	31.14	33
RSC83-10	F _{2:3} Low	10.78	93	8.09	102	-3.53	128	13.36	85
RSC83-1	BC ₁ F _{2:3} Low	45.25**	8	53.87**	3	35.92*	19	22.16	55
RSC112-5	F _{2:3} High	29.96	36	10.84	92	22.85	53	11.79	90
RSC112-19	BC ₁ F _{2:3} High	39.38*	15	30.44	34	42.27*	11	13.80	82
RSC112-8	F _{2:3} Low	24.53	47	5.58	108	12.63	88	1.02	120
RSC112-15	BC ₁ F _{2:3} Low	25.54	43	10.74	94	19.94	62	8.27	99
RSC76-4	F _{2:3} High	52.81**	4	2.17	115	1.44	119	-20.08	152
RSC76-16	BC ₁ F _{2:3} High	28.93	37	20.77	60	10.64	95	-33.62*	159
RSC76-13	F _{2:3} Low	21.15	59	7.41	104	-14.18	146	-20.51	153
RSC76-2	BC ₁ F _{2:3} Low	35.22*	22	-10.54	137	14.53	81	-34.22*	160
RSC38-5	F _{2:3} High	25.73	42	33.02*	29	16.79	70	-12.68	144
RSC38-15	BC ₁ F _{2:3} High	27.69	38	15.79	77	-12.60	143	-24.67	156
RSC38-8	F _{2:3} Low	18.42	66	9.38	96	-7.07	134	-12.73	145
RSC38-9	BC ₁ F _{2:3} Low	32.45*	31	23.50	51	8.72	98	-11.26	139
RSC37-12	F _{2:3} High	47.35**	7	30.43	35	2.64	112	-3.99	129
RSC37-12	BC ₁ F _{2:3} High	33.91*	25	13.42	84	9.25	97	-22.55	155
RSC37-7	F _{2:3} Low	19.48	64	18.02	67	-11.86	140	-3.38	127
RSC37-8	BC ₁ F _{2:3} Low	39.55*	14	11.40	91	-0.52	121	2.00	116
RSC15-13	F _{2:3} High	24.24	48	16.78	71	-25.18	157	-2.25	124
RSC15-15	BC ₁ F _{2:3} High	33.56*	27	19.82	63	23.79	50	6.63	105
RSC15-11	F _{2:3} Low	25.48	44	27.33	39	-14.92	149	-3.12	125
RSC15-14	BC ₁ F _{2:3} Low	16.71	73	-2.12	123	2.61	113	5.51	109
RSC124-9	F _{2:3} High	55.88**	2	33.17*	28	42.00*	12	23.30	52
RSC124-16	BC ₁ F _{2:3} High	12.73	87	12.56	89	20.43	61	-3.22	126
RSC124-3	F _{2:3} Low	26.81	40	2.95	111	-12.22	141	-10.84	138
RSC124-4	BC ₁ F _{2:3} Low	38.58*	16	13.58	83	16.97	69	1.94	117
RSC117-2	F _{2:3} High	34.75*	23	24.84	46	6.30	106	31.52	32
RSC117-4	BC ₁ F _{2:3} High	37.38*	18	32.54*	30	23.91	49	49.64**	6
RSC117-10	F _{2:3} Low	49.68**	5	16.76	72	16.52	74	16.28	76
RSC117-3	BC ₁ F _{2:3} Low	38.42*	17	7.74	103	13.08	86	25.78	41
RSC19-3	F _{2:3} High	43.96*	10	8.20	101	-5.18	130	16.28	75
RSC19-17	BC ₁ F _{2:3} High	41.78*	13	5.01	110	-22.36	154	-15.02	150
RSC19-1	F _{2:3} Low	22.05	57	2.49	114	-7.54	135	-5.57	132
RSC19-10	BC ₁ F _{2:3} Low	33.75*	26	25.13	45	-25.94	158	1.64	118
SE hybrid	15.81								

SE: Standard error, * and **: significant at the 0.05 and 0.01 probability levels.

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

expressed significant positive MPH with three of the four testers: RSC83-1 $F_{2:3}$ high and RSC124-9 $F_{2:3}$ high with A.301, A.319, and A.Tx3197 with RSC117-4 $BC_1F_{2:3}$ high with A.301, A.319, and A.338. Significant positive heterosis for grain yield was reported by Hovny et al. (2000), Abo-Elwafa (2005), Kenga et al. (2005), Hovny and El-Dsouky (2007), Abdel-Mottaleb (2009), and Amir and Mohamed (2015). Both midparent and better parent positive heterosis were reported by Premalatha et al. (2006), Sharma and Sharma (2006), and El-Dardeer et.al. (2011).

The range of MPH for the number of days to anthesis varied from -8.97 (A.301*RSC73-5 $BC_1F_{2:3}$ low) to 9.34% (A.319*RSC37-12 $BC_1F_{2:3}$ high), with a standard error equal to 1.574 (Table 34). The MPH in the negative direction was considered to be desirable for the trait. As many as 56 hybrids expressed significant negative MPH ranging from -8.97 (A.301*RSC73-5 $BC_1F_{2:3}$ low) to -3.19% (A.319*RSC38-8 $F_{2:3}$ low). Some of the early flowering hybrids with tester A.301 consisted of lines RSC124-4 ($BC_1F_{2:3}$ low), RSC15-15 ($BC_1F_{2:3}$ high), RSC37-8 ($BC_1F_{2:3}$ low), and RSC112-15 ($BC_1F_{2:3}$ low) with the number of days to anthesis ranging from 66.3 to 67.0 and with significant negative MPH values of -4.95, -3.47, -4.08, and -4.75, respectively. The results of heterosis for earliness were in accordance with the findings of Kenga et al. (2005), Premalatha et al. (2006), Hovny and El-Dsouky (2007), Abdel-Mottaleb (2009), Essa (2009), Mahdy et al. (2011), Abou-Amer and Kewan (2014), Omar et al. (2014), and Amir and Mohamed (2015).

Almost all the hybrids were taller than their respective mid-parents. The average MPH ranged from -10.44 (A.301*RSC124-3 $F_{2:3}$ low) to 88.29% (A.338*RSC73-5

Table 34. Midparent heterosis estimates for days to anthesis for each hybrid combination in the combined analysis in two environments of 2015 and 2016 at Vega, TX, with overall rank.

RSC†	Lines Selections‡	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC73-9	F _{2:3} High	0.12	58	3.97*	14	-3.07	101	4.92**	11
RSC73-6	BC ₁ F _{2:3} High	-1.37	77	0.73	47	-4.02*	119	2.29	25
RSC73-1	F _{2:3} Low	-1.47	78	7.85**	3	5.61**	6	5.25**	8
RSC73-5	BC ₁ F _{2:3} Low	-8.97**	160	-0.46	65	-5.12**	134	-2.25	92
RSC83-1	F _{2:3} High	-2.56	98	0.83	44	2.24	26	-0.76	69
RSC83-14	BC ₁ F _{2:3} High	-2.07	90	1	42	-4.61**	130	1.56	37
RSC83-10	F _{2:3} Low	-5.52**	137	0.67	48	-4.21*	123	-2.44	95
RSC83-1	BC ₁ F _{2:3} Low	-1.62	80	4.99**	10	-2.18	91	2.07	30
RSC112-5	F _{2:3} High	1.69	36	1.31	41	-5.76**	142	-5.71**	140
RSC112-19	BC ₁ F _{2:3} High	0.85	43	-0.87	72	-5.65**	139	-3.08	102
RSC112-8	F _{2:3} Low	-7.3**	154	-3.43*	111	-4.6**	129	-0.15	62
RSC112-15	BC ₁ F _{2:3} Low	-4.75**	132	0.39	54	-3.24*	106	2.1	29
RSC76-4	F _{2:3} High	-3.28*	107	1.75	34	-1.16	74	-2.29	93
RSC76-16	BC ₁ F _{2:3} High	2.39	24	1.92	32	-1.7	82	-1.53	79
RSC76-13	F _{2:3} Low	0.33	55	2.42	23	-0.65	66	2.84	19
RSC76-2	BC ₁ F _{2:3} Low	0.5	52	0.76	46	-3.14	103	0.14	56
RSC38-5	F _{2:3} High	-4.31*	126	2.43	22	-4.03*	120	1.56	38
RSC38-15	BC ₁ F _{2:3} High	-5.8**	143	-1.86	86	-7.45**	155	2.01	31
RSC38-8	F _{2:3} Low	-6.17**	148	-3.19*	105	-8.47**	158	2.64	21
RSC38-9	BC ₁ F _{2:3} Low	-0.33	63	-0.74	68	-3.92*	116	7.54**	4
RSC37-12	F _{2:3} High	-3.32*	109	9.34**	1	-5.85**	144	-5.41**	135
RSC37-12	BC ₁ F _{2:3} High	-2.51	96	2.76	20	-4.68**	131	-0.01	60
RSC37-7	F _{2:3} Low	-6.11**	147	0.51	51	-6.75**	153	-1.09	73
RSC37-8	BC ₁ F _{2:3} Low	-4.08*	121	2.22	27	-5.91**	146	1.46	39
RSC15-13	F _{2:3} High	1.37	40	0.65	49	-3.31*	108	4.78**	12
RSC15-15	BC ₁ F _{2:3} High	-3.47*	112	-3.33*	110	-7.51**	156	0.02	59
RSC15-11	F _{2:3} Low	-0.06	61	8.15**	2	-1.72	83	3.87*	15
RSC15-14	BC ₁ F _{2:3} Low	-2.36	94	-0.82	71	-3.89*	114	0.77	45
RSC124-9	F _{2:3} High	-4.31*	125	-5.73**	141	-6.66**	152	-1.67	81
RSC124-16	BC ₁ F _{2:3} High	2.2	28	-4.39*	127	-6.57**	151	-1.94	87
RSC124-3	F _{2:3} Low	-3.06	100	-2	89	-4.3*	124	0.65	50
RSC124-4	BC ₁ F _{2:3} Low	-4.95**	133	0.48	53	-8.73**	159	-5.44**	136
RSC117-2	F _{2:3} High	-2.55	97	1.75	35	-1.35	76	3.2*	17
RSC117-4	BC ₁ F _{2:3} High	-5.89**	145	-1.78	84	-6.51**	150	-3.17	104
RSC117-10	F _{2:3} Low	-3.97*	117	-1.16	75	-7.95**	157	0.13	57
RSC117-3	BC ₁ F _{2:3} Low	-5.59**	138	-4.01*	118	-6.18**	149	-0.43	64
RSC19-3	F _{2:3} High	-1.81	85	4.76**	13	-0.67	67	2.88	18
RSC19-17	BC ₁ F _{2:3} High	1.77	33	5.44**	7	4.99**	9	3.38*	16
RSC19-1	F _{2:3} Low	-3.66*	113	6.29**	5	-4.19*	122	-0.81	70
RSC19-10	BC ₁ F _{2:3} Low	-4.44*	128	-1.99	88	-3.9*	115	-2.67	99
SE hybrid	1.57								

SE: Standard error, * and **: significant at the 0.05 and 0.01 probability levels.

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

BC₁F_{2:3} low) (Table 35). One hundred twenty-five hybrids had significant positive MPH, with a maximum plant height (255.6 cm) set by the cross A.338*RSC76-16 BC₁F_{2:3} high with all four selections of RSC76 ranking the best of the four tallest hybrids. The minimum plant height (92.6 cm) for all the crosses was set by A.301*RSC117-10 F_{2:3} low with a MPH value of -1.96%. The testers from shortest to tallest were A.301, A.319, A.Tx3197, and A.338, with average total plant heights of 92.0, 104.3, 112.3, and 128.6 cm, respectively, while the lines ranged from 80.4 (RSC83-1 BC₁F_{2:3} low) to 204.9 cm (RSC76-16 BC₁F_{2:3} high). The findings were in agreement with those obtained by Hovny and El-Dsouky (2007), Abdel-Mottaleb (2009), Essa (2009), Mahdy et al. (2011), and Amir and Mohamed (2015).

The average MPH for height to flag leaf varied from -14.09 (A.301*RSC124-3 F_{2:3} low) to 116.34% (A.338*RSC38-5 F_{2:3} high) (Table 36). One-hundred eleven hybrids had significant positive MPH, with the maximum height to the flag leaf (221.1 cm) by the cross A.338*RSC76-4 F_{2:3} high with all four selections of RSC76 being in the top five for height to the flag leaf. The minimum height to the flag leaf (59.7 cm) for all crosses was by A.301*RSC117-10 F_{2:3} low, followed by A.301*RSC38-5 F_{2:3} high (61.4 cm) with MPH values of -2.87 and -2.05%, respectively. With most of the hybrids being taller than their parents, the height to the flag leaf followed a similar pattern, and the findings are in agreement with those of Hovny and El-Dsouky (2007), Abdel-Mottaleb (2009), Essa (2009), Mahdy et al. (2011), and Amir and Mohamed (2015).

Longer panicle, greater panicle weight, and 1000-kernel weight were generally associated with greater yields and were some of the attributes for greater grain yield by

Table 35. Midparent heterosis estimates for total plant height (cm) for each hybrid combination in the combined analysis in three environments, Vega, TX, in 2015 and 2016, and Dumas, TX, in 2016, with overall rank.

RSC†	Lines Selections‡	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC73-9	F _{2:3} High	34.22**	78	58.97**	30	63.06**	22	71.63**	10
RSC73-6	BC ₁ F _{2:3} High	45.56**	56	60.51**	28	61.01**	26	73.26**	7
RSC73-1	F _{2:3} Low	12.73	127	43.29**	60	48.84**	46	59.29**	29
RSC73-5	BC ₁ F _{2:3} Low	38.46**	70	68.45**	12	77.82**	5	88.29**	1
RSC83-1	F _{2:3} High	20.17**	100	45.49**	57	46.07**	55	60.57**	27
RSC83-14	BC ₁ F _{2:3} High	43.21**	61	68.44**	13	67.09**	16	72.58**	8
RSC83-10	F _{2:3} Low	0.85	152	9.12	142	11.58	133	17.97*	106
RSC83-1	BC ₁ F _{2:3} Low	47.48**	50	65.27**	20	81.17**	3	77.05**	6
RSC112-5	F _{2:3} High	8.81	143	12.57	129	20.74**	99	-1.19	156
RSC112-19	BC ₁ F _{2:3} High	6.71	147	14.75*	121	22.95**	92	24.09**	87
RSC112-8	F _{2:3} Low	1.15	151	11.39	135	15.57*	118	15.03*	120
RSC112-15	BC ₁ F _{2:3} Low	-6.42	158	14.71*	123	19.45*	103	23.89**	88
RSC76-4	F _{2:3} High	27.9**	82	55.01**	33	61.82**	24	69.24**	11
RSC76-16	BC ₁ F _{2:3} High	17*	110	38.5**	69	50.99**	41	53.34**	38
RSC76-13	F _{2:3} Low	33.5**	79	48.36**	49	53.42**	37	51.24**	40
RSC76-2	BC ₁ F _{2:3} Low	30.15**	81	57.11**	32	66.8**	17	66.51**	18
RSC38-5	F _{2:3} High	0.12	153	19.43*	104	19.66*	101	85.58**	2
RSC38-15	BC ₁ F _{2:3} High	17.01*	109	47.39**	51	50.52**	43	71.76**	9
RSC38-8	F _{2:3} Low	8.36	144	37.72**	72	40.35**	67	54.34**	34
RSC38-9	BC ₁ F _{2:3} Low	10.68	139	35.47**	74	37.82**	71	54.29**	35
RSC37-12	F _{2:3} High	32.82**	80	79.59**	4	68.12**	15	54.18**	36
RSC37-12	BC ₁ F _{2:3} High	43.08**	62	43.68**	59	65.22**	21	65.36**	19
RSC37-7	F _{2:3} Low	-8.14	159	52.66**	39	49.12**	44	62.62**	23
RSC37-8	BC ₁ F _{2:3} Low	27.12**	83	57.6**	31	61.42**	25	68.22**	14
RSC15-13	F _{2:3} High	26.82**	84	11.88	132	48.78**	47	47.07**	53
RSC15-15	BC ₁ F _{2:3} High	7.61	145	15.66*	116	21.42**	96	25.87**	86
RSC15-11	F _{2:3} Low	23.64**	89	15.85*	115	23.61**	90	23.33**	91
RSC15-14	BC ₁ F _{2:3} Low	22.89**	93	47.36**	52	41.76**	65	47.01**	54
RSC124-9	F _{2:3} High	2.49	150	12.66	128	10.35	141	10.97	137
RSC124-16	BC ₁ F _{2:3} High	-0.9	155	19.6*	102	20.85**	98	34.38**	77
RSC124-3	F _{2:3} Low	-10.44	160	-0.47	154	10.48	140	16.47*	112
RSC124-4	BC ₁ F _{2:3} Low	3.8	149	12.82	126	15.22*	119	48.59**	48
RSC117-2	F _{2:3} High	7.3	146	11.39	136	21.64**	95	26.8**	85
RSC117-4	BC ₁ F _{2:3} High	4.58	148	16.03*	114	17.95*	107	16.46*	113
RSC117-10	F _{2:3} Low	-1.96	157	15.62*	117	14.72*	122	16.91*	111
RSC117-3	BC ₁ F _{2:3} Low	34.49**	76	12.39	130	22.17**	94	21.13**	97
RSC19-3	F _{2:3} High	18.65*	105	35.2**	75	35.59**	73	38.6**	68
RSC19-17	BC ₁ F _{2:3} High	17.52*	108	50.84**	42	45.12**	58	49.06**	45
RSC19-1	F _{2:3} Low	11.56	134	13.89*	125	12.32	131	14.16*	124
RSC19-10	BC ₁ F _{2:3} Low	10.95	138	43.07**	63	40.83**	66	42.13**	64
SE hybrid	6.88								

SE: Standard error, * and **: significant at the 0.05 and 0.01 probability levels.

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

Table 36. Midparent heterosis estimates for height to flag leaf (cm) for each hybrid combination in the combined analysis in three environments, Vega, TX, in 2015 and 2016, and Dumas, TX, in 2016, with overall rank.

RSC†	Lines Selections‡	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC73-9	F _{2:3} High	40.29**	75	68.81**	25	65.65**	33	80.33**	15
RSC73-6	BC ₁ F _{2:3} High	50.61**	64	71.29**	24	67.37**	28	84.56**	9
RSC73-1	F _{2:3} Low	12.68	130	59.43**	46	60.64**	44	73.28**	22
RSC73-5	BC ₁ F _{2:3} Low	43.41**	70	80.66**	14	83.19**	10	101.79**	3
RSC83-1	F _{2:3} High	32.52*	85	58.48**	49	52.57**	62	77.42**	19
RSC83-14	BC ₁ F _{2:3} High	57.61**	52	93.27**	4	82.01**	13	85.34**	8
RSC83-10	F _{2:3} Low	0.21	151	15.82	122	11.24	135	22.74**	102
RSC83-1	BC ₁ F _{2:3} Low	55.46**	56	78.63**	18	87.8**	6	87.67**	7
RSC112-5	F _{2:3} High	6.46	146	11.32	134	14.67	125	-6.63	157
RSC112-19	BC ₁ F _{2:3} High	7.47	142	19.52**	109	24.1**	100	34.0*	84
RSC112-8	F _{2:3} Low	1.01	149	17.14	113	13.14	126	19.9**	107
RSC112-15	BC ₁ F _{2:3} Low	-6.75	158	18.64**	111	18.89**	110	29.18*	91
RSC76-4	F _{2:3} High	35.2**	83	63.75**	37	65.79**	32	79**	17
RSC76-16	BC ₁ F _{2:3} High	24.2**	99	48.22**	66	55.37**	57	65.54**	35
RSC76-13	F _{2:3} Low	35.24**	82	53.72**	59	58.04**	50	58.52**	48
RSC76-2	BC ₁ F _{2:3} Low	38.72**	76	62.2**	40	72.69**	23	74.78**	21
RSC38-5	F _{2:3} High	-2.05	153	21.16**	104	16.24	120	116.34**	1
RSC38-15	BC ₁ F _{2:3} High	25.35**	95	58.64**	47	57.65**	51	92.93**	5
RSC38-8	F _{2:3} Low	6.38	147	47.67**	68	43.91**	69	66.93**	30
RSC38-9	BC ₁ F _{2:3} Low	7.26	144	36.28**	79	42.59**	71	66.96**	29
RSC37-12	F _{2:3} High	38.31**	77	107.92**	2	80.28**	16	62.75**	39
RSC37-12	BC ₁ F _{2:3} High	53.18**	60	54.31**	58	68.02**	27	76.35**	20
RSC37-7	F _{2:3} Low	-9.41	159	68.16**	26	56.66**	54	82.51**	12
RSC37-8	BC ₁ F _{2:3} Low	24.48**	98	65.55**	34	63.01**	38	82.94**	11
RSC15-13	F _{2:3} High	38.16**	78	6.79	145	53.14**	61	61.2**	43
RSC15-15	BC ₁ F _{2:3} High	10.08	138	22.77**	101	16.24	119	31.05*	88
RSC15-11	F _{2:3} Low	29.91*	89	16.72	114	24.71**	97	31.77*	86
RSC15-14	BC ₁ F _{2:3} Low	29.67*	90	62.1**	41	47.77**	67	59.52**	45
RSC124-9	F _{2:3} High	0.26	150	12.99	129	7.29	143	17.39	112
RSC124-16	BC ₁ F _{2:3} High	-6.48	156	20.57**	106	21.24**	103	40.86**	73
RSC124-3	F _{2:3} Low	-14.09	160	-3.39	155	10.29	137	16.72	115
RSC124-4	BC ₁ F _{2:3} Low	13.09	127	16.55	116	20.88**	105	66.7**	31
RSC117-2	F _{2:3} High	10.71	136	14.74	124	24.85**	96	35.28**	81
RSC117-4	BC ₁ F _{2:3} High	-0.69	152	16.17	121	8.03	140	16.38	118
RSC117-10	F _{2:3} Low	-2.87	154	11.33	133	11.52	132	19.54**	108
RSC117-3	BC ₁ F _{2:3} Low	42.27**	72	3.59	148	26.86*	92	25.76*	93
RSC19-3	F _{2:3} High	15.11	123	35.9**	80	31.33*	87	40.43**	74
RSC19-17	BC ₁ F _{2:3} High	25.75*	94	64.66**	36	50.03**	65	61.92**	42
RSC19-1	F _{2:3} Low	7.95	141	16.52	117	9.68	139	12.42	131
RSC19-10	BC ₁ F _{2:3} Low	13.09	128	56.97**	53	52.51**	63	55.78**	55
SE hybrid	8.92								

SE: Standard error, * and **: significant at the 0.05 and 0.01 probability levels.

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

the sorghum in the study. A large panicle in a hybrid is initiated earlier and develops faster than parents (Blum and Pnuel, 1990). For panicle length, MPH varied from -10.43 (A.338*RSC73-1 F_{2:3} low) to 22.48% (A.301*RSC112-5 F_{2:3} high) (Table 37). Fifty-four hybrids expressed significant positive MPH for panicle length while only one hybrid expressed significant negative MPH. Two hybrids expressed significant positive MPH for all four testers RSC83-14 BC₁F_{2:3} high and RSC19-3 F_{2:3} high, while four other hybrids expressed significant positive MPH for three of the four testers. RSC112-5 F_{2:3} high and RSC124-9 F_{2:3} high expressed significant positive MPH with A.301, A.319, and A.Tx3197, while RSC112-19 BC₁F_{2:3} high and RSC117-2 F_{2:3} high expressed significant positive MPH with A.301, A.319, and A.338. Midparent significant positive heterosis was reported by Sharma and Sharma (2006) and Kanbar et al. (2011) for panicle length. This also was in agreement with the findings of Hovny and El-Dsouky (2007), Abdel-Mottaleb (2009), Essa (2009), Mahdy et al. (2011), and Amir and Mohamed (2015), while Premalatha et al. (2006) reported negative heterosis for the trait.

MPH estimates for plant exertion ranged from -62.6 (A.301*RSC38-15 BC₁F_{2:3} high) to 557.8% (A.319*RSC73-9 F_{2:3} high) (Table 38). Thirty hybrids expressed significant positive MPH ranging from 207.4 (A.338*RSC38-9 BC₁F_{2:3} low) to 557.8% (A.319*RSC73-9 F_{2:3} high), while 30 hybrids expressed non-significant negative MPH ranging from -62.6 (A.301*RSC38-15 BC₁F_{2:3} high) to -4.52% (A.319*RSC19-1 F_{2:3} low). One line (RSC37-12 BC₁F_{2:3} high) expressed significant positive MPH for all four testers, while RSC37-8 BC₁F_{2:3} low expressed significant positive estimates for A.301, A.319, A.Tx3197, and RSC73-9 F_{2:3} high, and RSC83-1 F_{2:3} high expressed significant

Table 37. Midparent heterosis estimates for panicle length (cm) for each hybrid combination in the combined analysis in three environments, Vega, TX, in 2015 and 2016, and Dumas, TX, in 2016, with overall rank.

RSC†	Lines Selections‡	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC73-9	F _{2:3} High	1.15	121	2.25	111	-4.14	150	-3.85	149
RSC73-6	BC ₁ F _{2:3} High	5.11	80	2.09	114	3.26	102	-0.78	135
RSC73-1	F _{2:3} Low	0.45	129	-5.01	154	-4.94	153	-10.43*	160
RSC73-5	BC ₁ F _{2:3} Low	5.86	77	7.78	60	3.99	92	8.93*	49
RSC83-1	F _{2:3} High	-1.59	139	9.88*	39	8.49*	52	2	115
RSC83-14	BC ₁ F _{2:3} High	12.77**	20	15.69**	12	10*	38	12.58**	22
RSC83-10	F _{2:3} Low	4.68	88	7.73	61	5.62	79	1.88	116
RSC83-1	BC ₁ F _{2:3} Low	4.17	91	3.36	101	7.03	67	2.76	107
RSC112-5	F _{2:3} High	22.48**	1	16.56**	10	19.25**	2	3.6	99
RSC112-19	BC ₁ F _{2:3} High	17.98**	5	4.71	87	8.48*	53	9.44*	42
RSC112-8	F _{2:3} Low	9.75*	40	6.95	69	2.14	113	7.4	64
RSC112-15	BC ₁ F _{2:3} Low	2.92	105	4.79	84	7.62	63	3.75	95
RSC76-4	F _{2:3} High	-5.1	155	-0.08	131	0.09	130	2.6	108
RSC76-16	BC ₁ F _{2:3} High	3.69	96	8.99*	47	7.28	65	9.33*	43
RSC76-13	F _{2:3} Low	4.77	85	6.58	72	8.35*	54	7.91	57
RSC76-2	BC ₁ F _{2:3} Low	2.94	104	8.96*	48	5.66	78	11.33*	27
RSC38-5	F _{2:3} High	10.62*	32	16.64**	9	4.87	83	1.46	117
RSC38-15	BC ₁ F _{2:3} High	-0.3	133	0.68	125	-8.11	159	-0.39	134
RSC38-8	F _{2:3} Low	-2.11	142	2.15	112	1.37	118	2.55	109
RSC38-9	BC ₁ F _{2:3} Low	10.66*	31	16.45**	11	3.62	97	0.54	127
RSC37-12	F _{2:3} High	8.07	55	-1.89	141	-5.71	156	4.77	86
RSC37-12	BC ₁ F _{2:3} High	-4.84	152	3.83	94	-2.25	143	0.53	128
RSC37-7	F _{2:3} Low	0.99	123	2.53	110	-1.31	136	-3.79	148
RSC37-8	BC ₁ F _{2:3} Low	-4.37	151	9.54*	41	-1.32	137	-2.52	145
RSC15-13	F _{2:3} High	-3.54	147	13.06**	18	-1.72	140	1.13	122
RSC15-15	BC ₁ F _{2:3} High	3.15	103	7.87	58	6.77	71	10.01*	37
RSC15-11	F _{2:3} Low	9.15*	45	11.52*	25	9.32*	44	8.05	56
RSC15-14	BC ₁ F _{2:3} Low	10.22*	35	3.49	100	17.24**	6	5.88	75
RSC124-9	F _{2:3} High	11.91**	24	14.62**	15	12.39**	23	3.6	98
RSC124-16	BC ₁ F _{2:3} High	10.34*	33	7.62	62	5.01	82	9.12*	46
RSC124-3	F _{2:3} Low	11.29*	28	14.14**	16	1.28	119	3.92	93
RSC124-4	BC ₁ F _{2:3} Low	2.85	106	8.92*	50	-1.4	138	-2.44	144
RSC117-2	F _{2:3} High	18.27**	4	11.47*	26	-6.07	158	13.29**	17
RSC117-4	BC ₁ F _{2:3} High	12.91**	19	10.09*	36	12.62**	21	6.77	70
RSC117-10	F _{2:3} Low	6.97	68	10.67*	30	7.05	66	10.32*	34
RSC117-3	BC ₁ F _{2:3} Low	5.87	76	7.79	59	5.02	81	4.5	89
RSC19-3	F _{2:3} High	15.57**	13	19.04**	3	16.86**	8	16.86**	7
RSC19-17	BC ₁ F _{2:3} High	6.41	73	1.24	120	-0.18	132	0.7	124
RSC19-1	F _{2:3} Low	4.25	90	15.03**	14	6.28	74	8.9*	51
RSC19-10	BC ₁ F _{2:3} Low	10.72*	29	0.56	126	-3.34	146	-6.04	157
SE hybrid	4.08								

SE: Standard error, * and **: significant at the 0.05 and 0.01 probability levels.

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

Table 38. Midparent heterosis estimates for plant exsertion (cm) for each hybrid combination in the combined analysis in three environments, Vega, TX, in 2015 and 2016, and Dumas, TX, in 2016, with overall rank.

RSC†	Lines Selections‡	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC73-9	F _{2:3} High	179.16	35	557.79**	1	526.67**	2	210.79*	29
RSC73-6	BC ₁ F _{2:3} High	153.69	41	379.31**	8	182.48	33	212.94*	25
RSC73-1	F _{2:3} Low	80.4	65	314.06**	13	185.96	32	94.06	54
RSC73-5	BC ₁ F _{2:3} Low	93.43	55	230.31*	22	300.37**	14	182.12	34
RSC83-1	F _{2:3} High	77.62	67	241.44*	19	147.82	43	23.1	104
RSC83-14	BC ₁ F _{2:3} High	59.24	75	75.41	68	53.85	80	84.39	63
RSC83-10	F _{2:3} Low	-5.5	133	-48.16	156	23.89	102	16.87	111
RSC83-1	BC ₁ F _{2:3} Low	154.44	39	318.17**	11	342.97**	10	220.83*	24
RSC112-5	F _{2:3} High	-10.38	139	36.84	91	92.11	58	61.82	74
RSC112-19	BC ₁ F _{2:3} High	-15.48	145	1.62	128	43.03	87	-40.33	155
RSC112-8	F _{2:3} Low	-25.74	152	-10.89	140	72.12	70	-6.56	135
RSC112-15	BC ₁ F _{2:3} Low	-19.42	148	22.56	106	54.24	79	25.91	101
RSC76-4	F _{2:3} High	91.19	59	211.31*	28	270.06*	17	92.57	56
RSC76-16	BC ₁ F _{2:3} High	-12.03	142	6.09	121	57.77	77	0.25	129
RSC76-13	F _{2:3} Low	120.78	47	403.06**	6	153.98	40	33.74	95
RSC76-2	BC ₁ F _{2:3} Low	63.9	73	400.39**	7	162.96	37	90.19	60
RSC38-5	F _{2:3} High	-24.94	151	94.16	53	199.48	31	21.11	107
RSC38-15	BC ₁ F _{2:3} High	-50.64	157	283.65*	15	246.91*	18	-53.53	158
RSC38-8	F _{2:3} Low	79.67	66	30.61	97	105.28	50	31.93	96
RSC38-9	BC ₁ F _{2:3} Low	115.02	49	236.68*	20	167.42	36	207.4*	30
RSC37-12	F _{2:3} High	149.89	42	376.27**	9	314.16**	12	144.74	44
RSC37-12	BC ₁ F _{2:3} High	222.45*	23	211.67*	27	487.86**	3	212.91*	26
RSC37-7	F _{2:3} Low	-22.01	150	84.5	62	87.35	61	23.36	103
RSC37-8	BC ₁ F _{2:3} Low	279.3*	16	448.05**	5	486.86**	4	49.52	83
RSC15-13	F _{2:3} High	7.1	119	55.53	78	155.16	38	13.43	115
RSC15-15	BC ₁ F _{2:3} High	0.13	130	-15.07	144	141.9	45	14.54	114
RSC15-11	F _{2:3} Low	2.76	125	38.59	89	17.36	110	-35.31	154
RSC15-14	BC ₁ F _{2:3} Low	-4.94	132	26.01	100	16.56	112	1.79	127
RSC124-9	F _{2:3} High	1.83	126	3.35	124	34.46	92	-6.44	134
RSC124-16	BC ₁ F _{2:3} High	22.64	105	41.91	88	66.96	71	18.92	108
RSC124-3	F _{2:3} Low	-19.08	147	14.67	113	57.98	76	47.05	85
RSC124-4	BC ₁ F _{2:3} Low	-55.45	159	-10.38	138	-11.07	141	12.2	116
RSC117-2	F _{2:3} High	-62.59	160	30.1	98	100.37	52	-17.59	146
RSC117-4	BC ₁ F _{2:3} High	6.27	120	34.15	93	235.33*	21	33.91	94
RSC117-10	F _{2:3} Low	-20.94	149	26.62	99	44.97	86	8.8	118
RSC117-3	BC ₁ F _{2:3} Low	49.96	82	131.48	46	5.37	123	18.34	109
RSC19-3	F _{2:3} High	65.99	72	101.38	51	118.18	48	75.36	69
RSC19-17	BC ₁ F _{2:3} High	-31.64	153	48.52	84	92.4	57	5.54	122
RSC19-1	F _{2:3} Low	82.94	64	-4.52	131	53.31	81	37.23	90
RSC19-10	BC ₁ F _{2:3} Low	-9.56	137	9.68	117	-13.6	143	-7.53	136
SE hybrid	99.05								

SE: Standard error, * and **: significant at the 0.05 and 0.01 probability levels.

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

positive estimates for A.319, A.Tx3197, and A.338. Many lines had a zero value for plant exertion, causing the magnitude of variation expressed by plant exertion MPH.

For all the hybrids, MPH estimates of three-panicle weight were positive values, suggesting greater panicle weights for hybrids over their parents. MPH for three-panicle weight ranged from 3.22 (A.301*RSC112-19 BC₁F_{2:3} high) to 126.75% (A.319*RSC37-12 F_{2:3} high). Of the 160 hybrids, 74 expressed significant positive MPH ranging from 48.42 (A.301*RSC15-15 BC₁F_{2:3} high) to 126.75% (A.319*RSC37-12 F_{2:3} high) (Table 39). Both RSC112-5 F_{2:3} high and RSC83-14 BCF_{2:3} high each appeared twice in the top six significant positive MPH estimates, with A.319, then A.301 and A.Tx3197, respectively. MPH for significant positive heterosis for panicle weight was reported by Sharma and Sharma (2006).

For 1000-kernel weight, MPH estimates ranged from -7.22 (A.Tx3197*RSC124-16 BC₁F_{2:3} high) to 46.6% (A.338*RSC112-5 F_{2:3} high), with RSC112-5 F_{2:3} high reporting the top two most significant positive MPH (Table 40). Of the 160 hybrids, 68 had significantly positive MPH estimates ranging from 12.63 (A.338*RSC83-1 F_{2:3} high) to 46.6% (A.338*RSC112-5 F_{2:3} high). Significance for 100-kernel weight heterosis was reported by Premalatha et al. (2006), and significant 1000-kernel weight MPH was reported by Mahdy et al. (2011).

MPH estimates for protein concentration in grain ranged from -12.03 (A.319*RSC112-19 BC₁F_{2:3} high) to 17.7% (A.Tx3197*RSC15-13 F_{2:3} high) (Table 41). Of the 160 hybrids, 20 had significant positive MPH estimates while six had significant

Table 39. Midparent heterosis estimates for three-panicle weight (g) for each hybrid combination in the combined analysis in three environments, Vega, TX, in 2015 and 2016, and Dumas, TX in 2016, with overall rank.

	Lines	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC†	Selections‡								
RSC73-9	F _{2:3} High	6.52	158	65.85*	37	35.42	111	71.41**	28
RSC73-6	BC ₁ F _{2:3} High	89.2**	10	70.82**	30	67.79*	34	92.05**	7
RSC73-1	F _{2:3} Low	26.63	129	72.2**	26	46.85	75	30.22	124
RSC73-5	BC ₁ F _{2:3} Low	63.15*	43	46.46	76	40.64	95	55.7*	51
RSC83-1	F _{2:3} High	21.23	136	32.46	116	39.75	97	63.07*	44
RSC83-14	BC ₁ F _{2:3} High	44.69	86	110.29**	2	96.06**	5	74.95**	24
RSC83-10	F _{2:3} Low	18.02	141	52.55*	61	14.04	145	19.38	139
RSC83-1	BC ₁ F _{2:3} Low	48.49*	73	84.51**	17	84.68**	16	72.93**	25
RSC112-5	F _{2:3} High	107.47**	4	93.86**	6	71.1**	29	18.55	140
RSC112-19	BC ₁ F _{2:3} High	3.22	160	37.79	103	51.78*	63	23.64	130
RSC112-8	F _{2:3} Low	45.59	83	29.59	125	17.08	142	19.91	137
RSC112-15	BC ₁ F _{2:3} Low	53.74*	55	36.59	107	50.4*	70	69.95**	32
RSC76-4	F _{2:3} High	44.6	87	77.58**	23	49.89*	71	50.95*	67
RSC76-16	BC ₁ F _{2:3} High	51.28*	66	84.17**	19	65.24*	39	45.05	85
RSC76-13	F _{2:3} Low	35.6	110	67.2*	35	46.25	78	46.42	77
RSC76-2	BC ₁ F _{2:3} Low	37.01	106	89.04**	11	81.23**	21	89.98**	8
RSC38-5	F _{2:3} High	23.42	131	53.46*	56	46.2	80	55.84*	50
RSC38-15	BC ₁ F _{2:3} High	53.45*	57	59.78*	46	32.05	119	39.61	98
RSC38-8	F _{2:3} Low	11.14	151	51.7*	64	13.32	146	85.22**	15
RSC38-9	BC ₁ F _{2:3} Low	37.08	105	50.93*	68	32.23	117	19.69	138
RSC37-12	F _{2:3} High	43.89	91	126.75**	1	26.87	128	35.22	113
RSC37-12	BC ₁ F _{2:3} High	58.59*	47	83.71**	20	66.65*	36	65.43*	38
RSC37-7	F _{2:3} Low	10.45	152	80.38**	22	32.06	118	64.32*	42
RSC37-8	BC ₁ F _{2:3} Low	16.4	143	87.03**	13	86.04**	14	84.3**	18
RSC15-13	F _{2:3} High	8.63	154	30.56	122	7.25	157	39.52	100
RSC15-15	BC ₁ F _{2:3} High	48.42*	74	87.47**	12	62.37*	45	64.85*	40
RSC15-11	F _{2:3} Low	28.91	126	44.54	88	37.71	104	51.83*	62
RSC15-14	BC ₁ F _{2:3} Low	39.58	99	28.86	127	15.73	144	53.32*	58
RSC124-9	F _{2:3} High	50.6*	69	52.69*	60	55.44*	52	44.13	89
RSC124-16	BC ₁ F _{2:3} High	31.51	120	45.18	84	22.09	134	36.47	108
RSC124-3	F _{2:3} Low	31.4	121	34.41	115	12.64	148	42.01	93
RSC124-4	BC ₁ F _{2:3} Low	41.33	94	22.77	133	23.02	132	11.56	150
RSC117-2	F _{2:3} High	49.09*	72	70.65**	31	21.37	135	51.35*	65
RSC117-4	BC ₁ F _{2:3} High	40.4	96	44.1	90	57.39*	49	45.9	82
RSC117-10	F _{2:3} Low	38.11	102	46.1	81	36.28	109	43.36	92
RSC117-3	BC ₁ F _{2:3} Low	54.01*	54	108.95**	3	89.82**	9	35.3	112
RSC19-3	F _{2:3} High	30.47	123	9.14	153	38.86	101	58.31*	48
RSC19-17	BC ₁ F _{2:3} High	54.77*	53	35.04	114	46.21	79	52.81*	59
RSC19-1	F _{2:3} Low	11.91	149	7.5	155	7.28	156	6.3	159
RSC19-10	BC ₁ F _{2:3} Low	72.11**	27	64.75*	41	67.87*	33	12.66	147
SE hybrid	23.92								

SE: Standard error, * and **: significant at the 0.05 and 0.01 probability levels.

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

Table 40. Midparent heterosis estimates for 1000-kernel weight (g) for each hybrid combination in the combined analysis in three environments, Vega, TX, in 2015 and 2016, and Dumas, TX, in 2016, with overall rank.

RSC†	Lines Selections‡	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC73-9	F _{2:3} High	10.99	81	24.82**	15	25.3**	14	38.66**	3
RSC73-6	BC ₁ F _{2:3} High	9.11	94	11.91	78	12.92*	65	32.47**	4
RSC73-1	F _{2:3} Low	3.46	129	15.34*	47	9.46	92	20.16**	29
RSC73-5	BC ₁ F _{2:3} Low	20.94**	26	23.33**	18	15.67*	45	17.08*	41
RSC83-1	F _{2:3} High	-1.22	151	6.99	113	7.35	111	9.7	90
RSC83-14	BC ₁ F _{2:3} High	12.04	73	13.44*	60	15.22*	48	20.37**	28
RSC83-10	F _{2:3} Low	22.95**	21	8.56	102	25.38**	12	25.3**	13
RSC83-1	BC ₁ F _{2:3} Low	14.29*	54	11.91	77	15.89*	44	12.63*	68
RSC112-5	F _{2:3} High	19.93**	30	42.79**	2	25.82**	10	46.6**	1
RSC112-19	BC ₁ F _{2:3} High	0.52	146	8.26	105	1.43	141	-6.98	159
RSC112-8	F _{2:3} Low	5.64	118	10.5	83	8.21	107	1.97	139
RSC112-15	BC ₁ F _{2:3} Low	10.36	85	10.84	82	1.67	140	1.22	144
RSC76-4	F _{2:3} High	24.3**	17	21.96**	24	14.84*	51	25.43**	11
RSC76-16	BC ₁ F _{2:3} High	22.77**	23	21.72**	25	14.15*	55	26.33**	8
RSC76-13	F _{2:3} Low	14.44*	52	26.12**	9	8.97	95	23.13**	20
RSC76-2	BC ₁ F _{2:3} Low	19.82**	31	10.39	84	11.93	75	18.23**	36
RSC38-5	F _{2:3} High	3.84	127	3.39	130	8.81	98	19.13**	32
RSC38-15	BC ₁ F _{2:3} High	13.9*	57	18.07**	38	15.49*	46	20.79**	27
RSC38-8	F _{2:3} Low	12.2	71	16.75*	42	16.44*	43	23.19**	19
RSC38-9	BC ₁ F _{2:3} Low	5.34	119	3.85	126	7.17	112	12.5	69
RSC37-12	F _{2:3} High	7.49	110	17.87**	39	1.41	142	8.31	104
RSC37-12	BC ₁ F _{2:3} High	13.26*	61	6.79	114	8.64	100	11.93	76
RSC37-7	F _{2:3} Low	-4.51	158	9.96	89	13.58*	59	13.98*	56
RSC37-8	BC ₁ F _{2:3} Low	2.28	137	-2.18	154	-0.42	150	1.97	138
RSC15-13	F _{2:3} High	6.48	115	11.83	79	8.45	103	22.77**	22
RSC15-15	BC ₁ F _{2:3} High	1.37	143	-0.03	148	3.52	128	8.25	106
RSC15-11	F _{2:3} Low	12.70*	67	9.64	91	10.23	87	24.43**	16
RSC15-14	BC ₁ F _{2:3} Low	10.08	88	18.8**	33	7.96	108	26.61**	7
RSC124-9	F _{2:3} High	8.73	99	6.24	116	-3.49	155	8.92	97
RSC124-16	BC ₁ F _{2:3} High	-3.97	156	6.14	117	-7.22	160	8.92	96
RSC124-3	F _{2:3} Low	17.21*	40	7.69	109	9.35	93	12.02	74
RSC124-4	BC ₁ F _{2:3} Low	12.25	70	0.72	145	4.95	122	18.1**	37
RSC117-2	F _{2:3} High	3.00	132	0.25	147	15.16*	49	4.97	121
RSC117-4	BC ₁ F _{2:3} High	-3.99	157	4.75	123	8.59	101	10.29	86
RSC117-10	F _{2:3} Low	12.04	72	5.16	120	13.73*	58	12.81*	66
RSC117-3	BC ₁ F _{2:3} Low	2.74	135	-0.12	149	2.9	133	12.95*	64
RSC19-3	F _{2:3} High	-1.97	152	2.55	136	3.26	131	13.22*	62
RSC19-17	BC ₁ F _{2:3} High	12.97*	63	18.51**	34	15.16*	50	11.32	80
RSC19-1	F _{2:3} Low	2.77	134	-2.07	153	4.47	125	4.64	124
RSC19-10	BC ₁ F _{2:3} Low	14.32*	53	28.36**	6	18.27**	35	28.68**	5
SE hybrid	6.21								

SE: Standard error, * and **: significant at the 0.05 and 0.01 probability levels.

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

Table 41. Midparent heterosis estimates for protein concentration in grain (%) for each hybrid combination in the combined analysis in two environments, Vega, TX, in 2015, and Dumas, TX, in 2016, with overall rank.

RSC†	Lines Selections‡	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC73-9	F _{2:3} High	1.49	73	6.87	24	3.09	54	4.46	45
RSC73-6	BC ₁ F _{2:3} High	-1.96	110	3.60	49	-3.52	131	-2.30	114
RSC73-1	F _{2:3} Low	-3.73	133	2.57	60	1.32	78	-0.12	97
RSC73-5	BC ₁ F _{2:3} Low	-4.23	134	0.13	94	5.09	38	-3.40	126
RSC83-1	F _{2:3} High	-7.63*	155	0.33	91	2.00	64	0.48	88
RSC83-14	BC ₁ F _{2:3} High	-3.20	123	4.80	40	3.44	51	1.67	67
RSC83-10	F _{2:3} Low	4.48	43	11.34**	4	9.60*	11	8.68*	13
RSC83-1	BC ₁ F _{2:3} Low	-4.52	138	3.66	46	3.38	52	-5.40	146
RSC112-5	F _{2:3} High	-6.32	151	-5.12	143	1.11	81	-3.43	128
RSC112-19	BC ₁ F _{2:3} High	-0.05	96	-12.03**	160	-11.98**	159	-5.92	149
RSC112-8	F _{2:3} Low	-1.04	102	-4.66	139	-4.68	140	-0.31	98
RSC112-15	BC ₁ F _{2:3} Low	-1.95	109	-3.06	120	-2.78	117	0.29	92
RSC76-4	F _{2:3} High	-6.08	150	1.45	74	9.71*	10	-1.71	107
RSC76-16	BC ₁ F _{2:3} High	-7.35	154	4.57	41	2.92	56	-1.60	106
RSC76-13	F _{2:3} Low	0.02	95	3.14	53	7.48	21	8.43*	15
RSC76-2	BC ₁ F _{2:3} Low	-9.74*	158	-3.46	129	6.59	25	-3.47	130
RSC38-5	F _{2:3} High	7.97*	18	8.33*	16	5.66	34	11.03**	6
RSC38-15	BC ₁ F _{2:3} High	-1.38	103	2.33	61	8.48*	14	1.55	72
RSC38-8	F _{2:3} Low	-2.87	118	-4.25	136	7.98*	17	1.64	68
RSC38-9	BC ₁ F _{2:3} Low	0.47	90	6.41	28	3.65	47	2.07	63
RSC37-12	F _{2:3} High	1.10	83	6.43	27	-5.31	145	-1.88	108
RSC37-12	BC ₁ F _{2:3} High	7.09	23	11.19**	5	-0.93	101	2.69	58
RSC37-7	F _{2:3} Low	2.96	55	2.61	59	1.16	80	0.69	86
RSC37-8	BC ₁ F _{2:3} Low	5.68	33	10.15*	8	-4.38	137	4.53	42
RSC15-13	F _{2:3} High	2.26	62	1.11	82	17.70**	1	9.72*	9
RSC15-15	BC ₁ F _{2:3} High	1.62	69	-7.88*	156	2.70	57	-3.36	125
RSC15-11	F _{2:3} Low	5.12	37	-7.31	153	5.34	35	6.26	29
RSC15-14	BC ₁ F _{2:3} Low	1.10	84	3.51	50	7.69*	19	9.01*	12
RSC124-9	F _{2:3} High	1.37	77	-1.54	105	4.47	44	1.57	70
RSC124-16	BC ₁ F _{2:3} High	-9.64*	157	-6.38	152	1.00	85	-5.30	144
RSC124-3	F _{2:3} Low	-4.78	141	-2.66	116	1.56	71	-5.71	147
RSC124-4	BC ₁ F _{2:3} Low	-0.6	100	-5.79	148	-2.63	115	3.65	48
RSC117-2	F _{2:3} High	1.92	65	1.89	66	5.75	32	-1.98	111
RSC117-4	BC ₁ F _{2:3} High	0.48	89	-2.14	113	0.55	87	-1.47	104
RSC117-10	F _{2:3} Low	-3.26	124	5.25	36	-4.24	135	-3.11	122
RSC117-3	BC ₁ F _{2:3} Low	5.86	31	-2.96	119	1.26	79	-5.07	142
RSC19-3	F _{2:3} High	0.24	93	7.47	22	-3.10	121	1.40	76
RSC19-17	BC ₁ F _{2:3} High	6.55	26	14.88**	2	5.90	30	13.23**	3
RSC19-1	F _{2:3} Low	-3.73	132	1.45	75	-2.05	112	-0.51	99
RSC19-10	BC ₁ F _{2:3} Low	-3.43	127	10.6*	7	4.92	39	7.64*	20
SE hybrid	3.72								

SE: Standard error, * and **: significant at the 0.05 and 0.01 probability levels.

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

negative MPH estimates. MPH estimates for starch concentration in grain ranged from -2.22 (A.Tx3197*RSC38-15 BC₁F_{2:3} high) to 1.62% (A.Tx3197*RSC112-19 BC₁F_{2:3} high) (Table 42). Of the 160 hybrids, four had significant positive MPH estimates while 36 had significant negative MPH estimates.

MPH estimates for fiber concentration in grain ranged from -8.496 (A.319*RSC38-15 BC₁F_{2:3} high) to 7.604% (A.301*RSC112-19 BC₁F_{2:3} high) (Table 43). Of the 160 hybrids, nine had significant positive MPH estimates, while 17 had significant negative MPH estimates. MPH estimates for fat concentration in the grain ranged from -20.136 (A.338*RSC76-4 F_{2:3} high) to 30.74% (A.Tx3197*RSC83-14 BC₁F_{2:3} high) (Table 44). Of the 160 hybrids, 25 had significant positive MPH estimates, while two had significant negative MPH estimates.

The degree of MPH varied considerably for all measured traits except starch concentration in grain. The high percentage of average MPH was observed for plant exsertion, because some parents had a zero value, followed by the height to the flag leaf, three-panicle weight, total height, grain yield, 1000-kernel weight, fat concentration in grain, panicle length, protein concentration in grain, number of days to anthesis, and fiber concentration in grain. Negative MPH was observed for the number of days to anthesis, total plant height, and height to the flag leaf for early maturing hybrids with dwarfing genes. MPH estimates for three-panicle weight was reported without any negative heterosis.

Table 42. Midparent heterosis estimates for starch concentration in grain (%) for each hybrid combination in the combined analysis in two environments, Vega, TX, in 2015, and Dumas, TX, in 2016, with overall rank.

RSC†	Lines Selections‡	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC73-9	F _{2:3} High	-0.886	106	-0.995	113	-1.463*	145	-0.871	104
RSC73-6	BC ₁ F _{2:3} High	-1.228*	129	-1.519*	146	-1.658**	155	-0.842	100
RSC73-1	F _{2:3} Low	-0.357	67	-0.952	112	-1.105	122	-0.936	111
RSC73-5	BC ₁ F _{2:3} Low	-0.383	69	-0.637	88	-0.641	90	-0.621	84
RSC83-1	F _{2:3} High	0.533	19	-0.621	83	-1.147*	125	-0.788	97
RSC83-14	BC ₁ F _{2:3} High	-0.809	98	-1.291*	135	-1.543*	150	-1.108	123
RSC83-10	F _{2:3} Low	-0.568	80	-1.457*	143	-0.671	92	-1.401*	140
RSC83-1	BC ₁ F _{2:3} Low	0.126	36	-1.268*	133	-1.531*	148	-0.091	48
RSC112-5	F _{2:3} High	0.042	40	0.002	43	-0.463	76	-0.626	85
RSC112-19	BC ₁ F _{2:3} High	0.147	35	1.52*	2	1.619**	1	0.202	31
RSC112-8	F _{2:3} Low	0.843	9	0.882	8	0.704	12	-0.428	71
RSC112-15	BC ₁ F _{2:3} Low	-0.178	55	0.677	13	0.383	25	-1.024	116
RSC76-4	F _{2:3} High	-0.188	56	-1.339*	138	-1.318*	137	-0.764	95
RSC76-16	BC ₁ F _{2:3} High	-0.013	44	-1.43*	141	-0.379	68	-1.046	118
RSC76-13	F _{2:3} Low	0.27	30	-0.143	53	-0.779	96	-1.26*	132
RSC76-2	BC ₁ F _{2:3} Low	-0.086	47	-1.008	114	-0.923	109	-0.812	99
RSC38-5	F _{2:3} High	-0.85	101	-1.207*	128	-1.763**	158	-1.678**	156
RSC38-15	BC ₁ F _{2:3} High	-1.10	121	-1.021	115	-2.216**	160	-1.535*	149
RSC38-8	F _{2:3} Low	0.005	42	0.009	41	-1.07	119	-0.64	89
RSC38-9	BC ₁ F _{2:3} Low	-0.432	72	-1.304*	136	-1.718**	157	-1.12	124
RSC37-12	F _{2:3} High	-0.858	102	-0.936	110	-0.177	54	-0.14	52
RSC37-12	BC ₁ F _{2:3} High	-1.455*	142	-1.372*	139	-1.196*	127	-0.565	79
RSC37-7	F _{2:3} Low	-0.491	77	-0.663	91	-0.552	78	-0.328	66
RSC37-8	BC ₁ F _{2:3} Low	-1.237*	130	-1.527*	147	-0.635	87	-1.073	120
RSC15-13	F _{2:3} High	-0.213	58	0.45	22	-1.648**	154	-1.156*	126
RSC15-15	BC ₁ F _{2:3} High	0.981	5	1.472*	3	0.185	32	0.812	11
RSC15-11	F _{2:3} Low	0.098	37	1.362*	4	-0.462	75	-0.894	108
RSC15-14	BC ₁ F _{2:3} Low	0.917	7	0.579	16	-0.859	103	-0.889	107
RSC124-9	F _{2:3} High	-0.101	49	0.362	27	-0.016	45	-0.441	73
RSC124-16	BC ₁ F _{2:3} High	0.54	18	0.347	28	-0.27	62	-0.189	57
RSC124-3	F _{2:3} Low	0.53	20	0.375	26	-0.572	81	0.152	34
RSC124-4	BC ₁ F _{2:3} Low	-0.056	46	0.59	15	0.042	39	-0.424	70
RSC117-2	F _{2:3} High	0.082	38	-0.279	63	-0.718	94	-0.235	59
RSC117-4	BC ₁ F _{2:3} High	0.178	33	0.415	23	-0.325	65	-0.268	61
RSC117-10	F _{2:3} Low	0.963	6	-0.267	60	0.826	10	0.272	29
RSC117-3	BC ₁ F _{2:3} Low	-0.311	64	0.566	17	-0.7	93	0.494	21
RSC19-3	F _{2:3} High	-0.611	82	-1.29*	134	-0.878	105	-1.027	117
RSC19-17	BC ₁ F _{2:3} High	-1.807**	159	-1.251*	131	-1.461*	144	-1.553*	151
RSC19-1	F _{2:3} Low	0.397	24	-0.127	50	0.611	14	-0.14	51
RSC19-10	BC ₁ F _{2:3} Low	-0.451	74	-1.615*	152	-0.63	86	-1.647**	153
SE hybrid	0.561								

SE: Standard error, * and **: significant at the 0.05 and 0.01 probability levels.

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

Table 43. Midparent heterosis estimates for fiber concentration in grain (%) for each hybrid combination in the combined analysis in two environments, Vega, TX, in 2015, and Dumas, TX, in 2016, with overall rank.

RSC†	Lines Selections‡	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC73-9	F _{2:3} High	-0.252	77	-3.562	141	2.29	28	-3.454	136
RSC73-6	BC ₁ F _{2:3} High	2.776	20	-2.059	111	3.091	16	1.555	37
RSC73-1	F _{2:3} Low	-0.433	82	-1.018	96	1.704	35	0.846	49
RSC73-5	BC ₁ F _{2:3} Low	0.457	63	-2.109	115	3.798	10	-0.587	86
RSC83-1	F _{2:3} High	-0.055	73	-4.203*	150	-0.302	79	-2.873	126
RSC83-14	BC ₁ F _{2:3} High	1.844	34	-0.222	76	2.727	21	-1.32	99
RSC83-10	F _{2:3} Low	-0.397	80	-3.459	137	-2.458	119	-2.101	114
RSC83-1	BC ₁ F _{2:3} Low	2.938	19	-0.006	71	5.644**	5	1.116	45
RSC112-5	F _{2:3} High	-0.679	89	-5.35*	156	-4.465*	152	0.545	56
RSC112-19	BC ₁ F _{2:3} High	7.604**	1	0.409	64	-1.872	106	-0.968	94
RSC112-8	F _{2:3} Low	3.561	11	-3.036	130	-4.169*	148	0.289	66
RSC112-15	BC ₁ F _{2:3} Low	1.139	42	-1.994	109	-8.288**	159	-4.136*	147
RSC76-4	F _{2:3} High	-1.609	102	-3.378	134	3.472	12	-0.133	75
RSC76-16	BC ₁ F _{2:3} High	-1.422	101	-1.885	107	0.072	68	-2.355	117
RSC76-13	F _{2:3} Low	1.138	43	-5.155*	154	1.118	44	1.22	40
RSC76-2	BC ₁ F _{2:3} Low	1.006	47	-3.769	143	0.521	60	-1.678	103
RSC38-5	F _{2:3} High	2.322	27	-3.315	133	0.609	55	0.521	59
RSC38-15	BC ₁ F _{2:3} High	-0.646	87	-8.496**	160	-0.828	91	-5.204*	155
RSC38-8	F _{2:3} Low	-0.504	83	-2.657	122	1.664	36	-0.421	81
RSC38-9	BC ₁ F _{2:3} Low	3.041	17	1.365	39	5.831**	4	2.697	22
RSC37-12	F _{2:3} High	3.174	14	-4.189*	149	6.336**	3	6.868**	2
RSC37-12	BC ₁ F _{2:3} High	2.975	18	0.462	62	0.000	70	-2.747	123
RSC37-7	F _{2:3} Low	1.181	41	-1.959	108	3.834*	9	-0.797	90
RSC37-8	BC ₁ F _{2:3} Low	4.743*	6	0.088	67	2.194	29	3.157	15
RSC15-13	F _{2:3} High	-0.076	74	-3.236	132	-3.385	135	0.616	54
RSC15-15	BC ₁ F _{2:3} High	-3.713	142	-3.922*	145	-5.013*	153	-2.605	121
RSC15-11	F _{2:3} Low	2.013	33	-1.168	98	0.693	53	4.558*	7
RSC15-14	BC ₁ F _{2:3} Low	-0.549	85	-3.535	140	-2.787	125	0.529	58
RSC124-9	F _{2:3} High	2.191	30	-2.038	110	0.532	57	-2.77	124
RSC124-16	BC ₁ F _{2:3} High	2.673	23	-2.993	129	-6.809**	158	-2.516	120
RSC124-3	F _{2:3} Low	-2.976	128	-3.959*	146	-0.666	88	-0.034	72
RSC124-4	BC ₁ F _{2:3} Low	0.961	48	0.841	51	-0.921	93	1.064	46
RSC117-2	F _{2:3} High	2.181	31	-2.077	112	-1.145	97	-0.296	78
RSC117-4	BC ₁ F _{2:3} High	3.232	13	-4.375*	151	-3.535	139	0.783	52
RSC117-10	F _{2:3} Low	2.497	25	-1.795	105	-0.869	92	0.844	50
RSC117-3	BC ₁ F _{2:3} Low	2.014	32	-6.173**	157	-3.51	138	-3.1	131
RSC19-3	F _{2:3} High	1.511	38	-0.978	95	0.021	69	-1.706	104
RSC19-17	BC ₁ F _{2:3} High	2.603	24	-3.892*	144	4.046*	8	-1.385	100
RSC19-1	F _{2:3} Low	0.389	65	-2.885	127	-2.398	118	-2.34	116
RSC19-10	BC ₁ F _{2:3} Low	-0.526	84	-2.099	113	0.516	61	2.462	26
SE hybrid	1.892								

SE: Standard error, * and **: significant at the 0.05 and 0.01 probability levels.

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

Table 44. Midparent heterosis estimates for fat concentration in grain (%) for each hybrid combination in the combined analysis in two environments, Vega, TX, in 2015, and Dumas, TX, in 2016, with overall rank.

RSC†	Lines Selections‡	A.301	Rank	A.319	Rank	A.Tx3197	Rank	A.338	Rank
RSC73-9	F _{2:3} High	6.424	78	18.845*	15	11.085	54	-1.579	122
RSC73-6	BC ₁ F _{2:3} High	14.866	33	21.655*	8	27.764**	3	8.367	67
RSC73-1	F _{2:3} Low	12.891	42	15.792	30	11.498	50	15.288	31
RSC73-5	BC ₁ F _{2:3} Low	3.908	97	13.303	39	-11.098	152	-0.624	114
RSC83-1	F _{2:3} High	-5.107	135	-0.789	117	19.73*	12	2.874	101
RSC83-14	BC ₁ F _{2:3} High	13.191	41	24.78**	6	30.744**	1	18.466*	17
RSC83-10	F _{2:3} Low	-0.893	118	20.848*	11	-2.66	129	13.348	38
RSC83-1	BC ₁ F _{2:3} Low	-3.916	131	20.982*	10	22.158*	7	0.611	108
RSC112-5	F _{2:3} High	2.551	104	4.564	92	5.244	85	2.581	103
RSC112-19	BC ₁ F _{2:3} High	4.645	90	17.522*	23	4.714	89	10.513	59
RSC112-8	F _{2:3} Low	3.972	96	-9.552	150	-18.515*	159	4.609	91
RSC112-15	BC ₁ F _{2:3} Low	12.551	43	11.333	52	4.891	87	18.29*	19
RSC76-4	F _{2:3} High	-11.905	154	-1.491	121	-9.961	151	-20.136*	160
RSC76-16	BC ₁ F _{2:3} High	-8.988	147	4.829	88	-16.325	158	-2.638	128
RSC76-13	F _{2:3} Low	-12.421	156	-6.745	143	3.331	100	-0.391	113
RSC76-2	BC ₁ F _{2:3} Low	-5.639	139	-7.33	145	-4.077	133	-9.491	149
RSC38-5	F _{2:3} High	-0.751	115	16.97*	25	17.406*	24	12.14	44
RSC38-15	BC ₁ F _{2:3} High	10.478	61	17.79*	21	9.297	64	11.773	48
RSC38-8	F _{2:3} Low	-9.027	148	2.674	102	3.758	98	-2.333	126
RSC38-9	BC ₁ F _{2:3} Low	5.187	86	18.071*	20	8.193	70	8.806	66
RSC37-12	F _{2:3} High	10.601	58	19.492*	14	11.063	56	4.274	95
RSC37-12	BC ₁ F _{2:3} High	11.635	49	14.129	35	16.815	26	5.574	82
RSC37-7	F _{2:3} Low	-4.952	134	4.373	94	3.602	99	-2.562	127
RSC37-8	BC ₁ F _{2:3} Low	8.205	69	11.356	51	21.391*	9	6.772	76
RSC15-13	F _{2:3} High	19.564*	13	13.51	36	25.818**	4	11.835	47
RSC15-15	BC ₁ F _{2:3} High	0.733	107	5.578	81	15.079	32	1.6	106
RSC15-11	F _{2:3} Low	10.876	57	7.416	74	17.606*	22	13.428	37
RSC15-14	BC ₁ F _{2:3} Low	11.937	45	18.796*	16	18.386*	18	6.221	80
RSC124-9	F _{2:3} High	-7.307	144	-2.041	125	-1.896	124	0.452	109
RSC124-16	BC ₁ F _{2:3} High	6.824	75	2.316	105	-1.08	120	-1.015	119
RSC124-3	F _{2:3} Low	-11.429	153	-1.764	123	-6.106	141	-5.302	136
RSC124-4	BC ₁ F _{2:3} Low	-7.803	146	-6.24	142	-5.585	137	-0.153	111
RSC117-2	F _{2:3} High	-0.759	116	9.654	63	15.92	28	6.446	77
RSC117-4	BC ₁ F _{2:3} High	-0.252	112	8.251	68	6.225	79	-0.128	110
RSC117-10	F _{2:3} Low	-12.076	155	10.506	60	-3.527	130	-5.952	140
RSC117-3	BC ₁ F _{2:3} Low	8.174	71	11.886	46	15.9	29	-3.992	132
RSC19-3	F _{2:3} High	8.011	72	29.132**	2	13.294	40	10.177	62
RSC19-17	BC ₁ F _{2:3} High	8.815	65	11.084	55	16.476	27	25.739**	5
RSC19-1	F _{2:3} Low	-5.618	138	11.176	53	-12.95	157	4.478	93
RSC19-10	BC ₁ F _{2:3} Low	5.301	84	14.698	34	7.682	73	5.457	83
SE hybrid	8.346								

SE: Standard error, * and **: significant at the 0.05 and 0.01 probability levels.

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines.

‡Selections are equal to the High and Low percentage of exotic genome recovery and the generation: High F_{2:3}, BC₁F_{2:3}, Low F_{2:3}, and BC₁F_{2:3}.

Correlation Estimates

The objective of any plant breeder is to select for one or more superior characters in a natural or artificially developed population. Because grain yield in sorghum is quantitative, selection only on the basis of the grain yield character is usually not very effective. However, selection based on its component characters could be more efficient and reliable. Knowledge of association between yield and its component traits and among the component parameters themselves can improve efficiency of selection in plant breeding. Correlation coefficient measures the mutual association between a pair of variables independent of other variables to be considered.

In the present investigation, character associations were studied and presented in Tables 45-51 to assess relationships among yield and its components for enhancing the usefulness of selection. Correlations were estimated separately for parents, including lines and testers, hybrids, checks, and for all genotypes combined at three parental locations: Vega, TX, in 2015 and 2016, and Dumas, TX, in 2016. Correlation coefficient estimates for SCA and heterosis for yield and agronomic traits of the 160 sorghum hybrids across environments and in the combined analysis were significant for all variables analyzed (Table 52).

Grain yield was significantly positively correlated with panicle length (0.219), three-panicle weight (0.205), and concentration of fiber and fat (0.235 and 0.163), respectively, in grain in the combined analysis across environments (Table 45). Significant negative correlations were found with the number of days to anthesis (-0.312), total plant height (-0.137), and height to the flag leaf (-0.158). Grain yield

Table 45. Pearson correlation coefficients among grain yield and agronomic traits based on six environments – Taylor and Vega, TX, and Hutchinson, KS, in 2015, and Dumas and Vega, TX, and Hutchinson, KS, in 2016.

Variable	Grain yield (Mgha-1)	Days to anthesis	Total height (cm)	Height to flag leaf (cm)	Panicle length (cm)	Plant exsertion (cm)	3-panicle weight (g)	1000-seed weight (g)	Grain Protein (%)	Grain Starch (%)	Grain Fiber (%)	Grain Fat (%)
Grain yield	1.000	-0.312**	-0.137**	-0.158**	0.219**	-0.032	0.205**	-0.007	0.022	0.050	0.235**	0.163**
Days to anthesis		1.000	0.151**	0.227**	-0.223**	-0.169**	0.015	0.101**	-0.017	-0.036	-0.030	0.202**
Total height			1.000	0.979**	-0.080**	0.354**	0.476**	0.358**	-0.133**	0.024	-0.209*	-0.067*
Height to flag leaf				1.000	-0.197**	0.174**	0.436**	0.379**	-0.122**	0.019	-0.198*	-0.056*
Panicle length					1.000	0.118**	0.361**	-0.262**	0.000	-0.015	0.072*	0.108**
Plant exsertion						1.000	0.125**	0.088**	-0.081**	0.041	-0.130**	-0.125**
3-panicle weight							1.000	0.250**	-0.220**	0.051	0.003	0.038
1000-seed weight								1.000	-0.068*	0.113**	-0.096**	-0.203**
Grain protein									1.000	-0.661**	0.208**	0.540**
Grain starch										1.000	-0.149**	-0.525**
Grain fiber											1.000	0.385**

* and ** significant at the 0.05 and 0.01 probability levels, N is variable.

Table 46. Pearson correlation coefficients among grain yield and agronomic traits for all genotypes (hybrids, parents, and checks) based on data from three all-parent locations of Vega, TX, in 2015 and 2016, and Dumas, TX, in 2016.

Variable	Grain yield (Mgha-1)	DTF	Total height (cm)	Height to flag leaf (cm)	Panicle length (cm)	Plant exsertion (cm)	3-panicle weight (g)	kernel weight (g)	Grain protein (%)	Grain starch (%)	Grain fiber (%)	Grain fat (%)
Grain yield	1.000	-0.007	-0.254**	-0.283**	0.188**	-0.042	0.057	-0.021	-0.078	0.058	0.241**	0.184**
DTF	1.000	1.000	0.149**	0.215**	-0.152**	-0.173**	0.068	0.105*	-0.017	-0.035	-0.031	0.204**
Total height			1.000	0.973**	0.095*	0.449**	0.531**	0.340**	-0.154**	-0.011	-0.185**	-0.124**
Height to flag leaf				1.000	-0.030	0.246**	0.480**	0.356**	-0.136**	-0.019	-0.185**	-0.099*
Panicle length					1.000	0.168**	0.390**	-0.229**	-0.037	-0.083	-0.001	-0.013
Plant exsertion						1.000	0.256**	0.154**	-0.120**	0.066	-0.070	-0.153**
3-panicle weight							1.000	0.297**	-0.280**	0.039	-0.082*	-0.157**
1000-seed weight								1.000	-0.179**	0.293**	-0.002	-0.236**
Grain protein									1.000	-0.646**	0.270**	0.577**
Grain starch										1.000	-0.167**	-0.582**
Grain fiber											1.000	0.422**

* and **: significant at the 0.05 and 0.01 probability levels, N = 626.

Table 47. Pearson correlation coefficients among grain yield and agronomic traits for hybrids based on data from three all-parent locations at Vega, TX, in 2015 and 2016, and Dumas, TX, in 2016.

Variable	Grain											
	Grain yield (Mgha-1)	Days to anthesis	Total height (cm)	Height to flag leaf (cm)	Panicle length (cm)	Plant exsertion (cm)	3-panicle weight (g)	1000- seed weight (g)	Grain protein (%)	Grain starch (%)	Grain fiber (%)	Grain fat (%)
Grain yield	1.000	-0.002	-0.486**	-0.491**	0.080	-0.165**	-0.133**	-0.179**	0.0162	-0.000	0.181**	0.172**
Days to anthesis		1.000	0.230**	0.301**	-0.176**	-0.181**	0.112*	0.103*	-0.036	-0.015	-0.031	0.254**
Total height			1.000	0.973**	0.055	0.412**	0.500**	0.317**	-0.154**	-0.063	-0.301**	-0.199**
Height to flag leaf				1.000	-0.070	0.207**	0.451**	0.338**	-0.138**	-0.072	-0.283**	-0.152**
Panicle length					1.000	0.174**	0.343**	-0.328**	-0.033	-0.040	-0.108*	-0.154**
Plant exsertion						1.000	0.237**	0.140**	-0.105*	0.039	-0.134**	-0.215**
3-panicle weight							1.000	0.223**	-0.277**	0.026	-0.179**	-0.268**
1000-seed weight								1.000	-0.206**	0.317**	-0.117*	-0.331**
Grain protein									1.000	-0.631**	0.316**	0.610**
Grain starch										1.000	-0.204**	-0.550**
Grain fiber											1.000	0.386**

* and **: significant at the 0.05 and 0.01 probability levels, N = 473.

Table 48. Pearson correlation coefficients among grain yield and agronomic traits for parents based on data from three all-parent locations at Vega, TX, in 2015 and 2016, and Dumas, TX, in 2016.

Variable	Height												
	Grain yield (Mgha-1)	Days to anthesis	Total height (cm)	to flag leaf (cm)	Panicle length (cm)	Plant exertion (cm)	panicle weight (g)	3-panicle weight (g)	1000-seed weight (g)	Grain protein (%)	Grain starch (%)	Grain fiber (%)	Grain fat (%)
Grain yield	1.000	0.157	0.081	0.035	0.092	0.151	0.127	0.123	0.123	-0.154	0.215*	0.210*	0.006
Days to anthesis		1.000	0.007	0.020	-0.002	-0.040	0.121	0.246*	0.246*	-0.039	-0.014	0.011	0.085
Total height			1.000	0.960**	0.017	0.540**	0.342**	0.180*	0.180*	-0.094	0.282**	-0.225*	-0.241*
Height to flag leaf				1.000	-0.101	0.303**	0.308**	0.209*	0.209*	-0.095	0.323**	-0.246*	-0.281**
Panicle length					1.000	0.008	0.298**	-0.250*	-0.250*	0.130	-0.347**	0.035	0.263**
Plant exertion						1.000	0.136	0.075	0.075	-0.087	0.115	-0.039	-0.071
3-panicle weight							1.000	0.296**	0.296**	-0.244*	0.151	-0.282**	-0.293**
1000-seed weight								1.000	1.000	-0.131	0.366**	0.014	-0.331**
Grain protein										1.000	-0.730**	0.289**	0.630**
Grain starch											1.000	-0.141	-0.730**
Grain fiber												1.000	0.392**

* and **: significant at the 0.05 and 0.01 probability levels, N = 132.

Table 49. Pearson correlation coefficients among grain yield and agronomic traits for lines based on data from three all-parent locations at Vega, TX, in 2015 and 2016, and Dumas, TX, in 2016.

Variable	Grain yield (Mgha-1)	Days to anthesis	Total height (cm)	Height to flag leaf (cm)	Panicle length (cm)	Plant exsertion (cm)	3-panicle weight (g)	1000-seed weight (g)	Grain protein (%)	Grain starch (%)	Grain fiber (%)	Grain fat (%)
Grain yield	1.000	0.160	0.076	0.034	0.034	0.157	0.093	0.170	-0.076	0.182	0.166	-0.003
Days to anthesis		1.000	-0.034	-0.045	0.040	0.005	0.130	0.269**	-0.091	0.028	0.055	0.032
Total height			1.000	0.962**	0.011	0.568**	0.337**	0.176	-0.117	0.320**	-0.188*	-0.246*
Height to flag leaf				1.000	-0.101	0.342**	0.308**	0.120*	-0.131	0.375**	-0.195*	-0.298**
Panicle length					1.000	-0.015	0.289**	-0.239*	0.182	-0.385**	0.020	0.305**
Plant exsertion						1.000	0.135	0.094	-0.078	0.110	-0.068	-0.056
3-panicle weight							1.000	0.318**	-0.239*	0.155	-0.302**	-0.304**
1000-seed weight								1.000	-0.158	0.396**	0.061	-0.340**
Grain protein									1.000	-0.714**	0.405**	0.659**
Grain starch										1.000	-0.230*	-0.761**
Grain fiber											1.000	0.413**

* and **: significant at the 0.05 and 0.01 probability levels, N = 120.

Table 50. Pearson correlation coefficients among grain yield and agronomic traits for testers based on data from three all-parent locations at Vega, TX, in 2015 and 2016, and Dumas, TX, in 2016.

Variable	Grain yield (Mgha-1)	Days to anthesis	Total height (cm)	Height to flag leaf (cm)	Panicle length (cm)	Plant exsertion (cm)	3-		1000-		Grain protein (%)	Grain starch (%)	Grain fiber (%)	Grain fat (%)
							panicle weight (g)	seed weight (g)						
Grain yield	1.000	0.356	0.793**	0.648*	0.548	0.051	0.393	-0.143	-0.431	-0.045	-0.194	-0.045		
Days to anthesis		1.000	0.468	0.667*	-0.329	-0.520	0.099	-0.065	0.353	-0.401	-0.133	0.836**		
Total height			1.000	0.910**	0.327	-0.041	0.664*	0.035	-0.378	-0.025	-0.639*	0.002		
Height to flag leaf				1.000	0.027	-0.429	0.524	0.227	-0.197	-0.065	-0.700*	0.257		
Panicle length					1.000	0.378	0.397	-0.457	-0.558	-0.007	-0.042	-0.601*		
Plant exsertion						1.000	0.130	-0.367	-0.221	0.131	0.306	-0.485		
3-panicle weight							1.000	-0.055	-0.264	-0.118	-0.514	-0.141		
1000-seed weight								1.000	-0.127	0.293	-0.462	0.043		
Grain protein									1.000	-0.760**	0.214	0.706*		
Grain starch										1.000	-0.014	-0.575		
Grain fiber											1.000	0.106		

* and **: significant at the 0.05 and 0.01 probability levels, N = 12.

Table 51. Pearson correlation coefficients among grain yield and agronomic traits for commercial hybrid checks based on data from three all-parent locations at Vega, TX, in 2015 and 2016, and Dumas, TX, in 2016.

Variable	Grain		Height		1000-		Grain		Grain		Grain	
	yield (Mgha- l)	Days to anthesis	Total height (cm)	leaf length (cm)	flag length (cm)	panicle exsertion (cm)	3-panicle weight (g)	seed weight (g)	protein (%)	starch (%)	fiber (%)	fat (%)
Grain yield	1.000	0.506*	0.481*	0.482*	0.395	-0.012	0.368	-0.046	-0.523*	-0.135	-0.075	0.001
Days to anthesis		1.000	0.267	0.493*	0.110	-0.402	0.271	0.199	0.035	-0.444*	0.423	0.565**
Total height			1.000	0.868**	0.380	0.535*	0.521*	0.399	-0.231	-0.073	-0.111	-0.098
Height to flag leaf				1.000	0.070	0.121	0.328	0.499*	0.025	-0.411	0.076	0.172
Panicle length					1.000	0.194	0.740**	-0.270	-0.416*	0.464*	-0.065	-0.161
Plant exsertion						1.000	0.191	0.168	-0.377	0.363	-0.399	-0.504*
3-panicle weight							1.000	-0.026	-0.217	0.246	0.056	0.085
1000-seed weight								1.000	0.544*	-0.124	0.255	0.303
Grain protein									1.000	-0.196	0.372	0.591**
Grain starch										1.000	-0.085	-0.375
Grain fiber											1.000	0.733**

* and **: significant at the 0.05 and 0.01 probability levels, N = 12.

Table 52. Pearson correlation coefficients between Specific Combining Ability (SCA) and the corresponding Midparent Heterosis (MPH) for yield and agronomic traits for 160 sorghum hybrids in three environments and combined analysis.

Variable	Vega 2015	Vega 2016	Dumas 2016	Combined locations
Grain yield (Mg ha ⁻¹)	0.519***	0.566***	0.706***	0.539***
DTF	0.592***	0.602***	.	0.595***
Total height (cm)	0.410***	0.451***	0.396***	0.409***
Height to flag leaf (cm)	0.421***	0.482***	0.425***	0.430***
Panicle length (cm)	0.679***	0.673***	0.618***	0.647***
Plant exsertion (cm)	0.406***	0.424***	0.304***	0.346***
3-panicle weight (g)	0.682***	0.643***	0.662***	0.580***
1000-kernel weight (g)	0.530***	0.631***	0.608***	0.557***
Protein content in grain (g)	0.726***	.	0.655***	0.665***
Starch content in grain (%)	0.600***	.	0.537***	0.547***
Fiber content in grain (%)	0.615***	.	0.690***	0.621***
Fat content in grain (%)	0.593***	.	0.555***	0.550***

***Probability of correlation different from zero is <0.001.

had the same pattern except for the number of days to anthesis and three-panicle weight when analyzed for the three-parent environments (Table 46). In the analysis for hybrids, significant positive correlations were found for grain yield and the concentrations of fiber and fat (0.181 and 0.172), respectively, in grain, while significant negative correlations were reported for total plant height (-0.486), height to the flag leaf (-0.491), plant exsertion (-0.165), three-panicle weight (-0.133), and 1000-kernel weight (-0.179) (Table 47). In the analysis of parents, concentration of starch and fiber in grain were the only

traits significantly positively correlated with grain yield (Table 48). In the analysis for testers, grain yield was significantly and positively correlated with total plant height (0.793) and height to the flag leaf (0.648) (Table 49). In the analysis for checks, grain yield was significantly and positively correlated with the number of days to anthesis (0.506), total plant height (0.482), and height to the flag leaf (0.482), while significant negative correlations were found for concentration of protein in grain (Table 50).

Almeida Filho et al. (2014) reported a positive association between plant height and grain yield and a negative correlation with the number of days to anthesis. Omar et al. (2014) found that plant height was significantly positively correlated with grain yield, while 1000-kernel weight had significant negative correlation. The negative correlation for grain yield and the number of days to anthesis also was reported by Exeaku and Mohammed (2006). This might have been caused by poor adaptation of very late-flowering materials included in the experiment in variable environments.

The number of days to anthesis was significantly positively correlated with total plant height (0.151), height to the flag leaf (0.227), 1000-kernel weight (0.101), and concentration of fat in grain (0.163), while significant negative correlations were found for panicle length (-0.223), plant exertion (-0.169), and concentration of fat in grain (0.1629) for genotypes in seven environments (Table 45). The number of days to anthesis exhibited the same pattern when analyzed in the three-parent environments (Table 46). In the analysis for hybrids, the number of days to anthesis had the same pattern except was positively significant for three-panicle weight (0.112) (Table 47). In the analysis for parents, the only significant positive correlation with the number of days

to anthesis was 1000-kernel weight (0.246) (Table 48). In the analysis of lines and testers, lines had a positive correlation for the number of days to anthesis with 1000-kernel weight (0.269), while testers had significant positive correlations with height to the flag leaf (0.667) and fat concentration in the grain (0.836) (Tables 49 and 50). Bohra et al. (1985), Jeyaprakash et al. (1997), and Iyanar et al. (2001) reported similar association between grain yield and the number of days to 50% flowering.

Total plant height was significantly positively associated with all traits evaluated except starch concentration in the grain for genotypes in seven environments and across parent environments only (Tables 45 and 46). In the analysis of hybrids and parents, panicle length was not significant (Tables 47 and 48). In the analysis of lines, 1000-kernel weight was not significantly correlated with total plant height (Table 49). In the analysis of testers, the only positive significant correlations with total plant height were height to the flag leaf (0.910) and three-panicle weight (0.664) (Table 50). Plant height had a high positive correlation coefficient with panicle length, three-panicle weight, and grain yield, indicating the possibility of obtaining taller plants with longer and heavier panicles that yielded more grain. This is in agreement with the findings of Gupta and Sidhu (1972). Similar results for the association of sorghum grain yield with plant height were reported by Mallinath et al. (2004), Ezeaku and Mohammed (2006), Mahajan et al. (2011), and El-Naim et al. (2012).

Three-panicle weight was significantly correlated with all traits except the number of days to anthesis and starch concentration in the grain for genotypes in seven environments (Table 45). Plants in environments with only sorghum parents exhibited

the same behavior except for grain yield which could be attributed to the few kernels produced at Vega, TX, in 2016. Replications were combined for the NIR analysis because three-panicle kernel weight was very small.

In the hybrid analysis, significant correlations were found for all traits except starch concentration in the grain, with negative correlations for grain yield (-0.133) and protein, starch, and fat concentration in the grain (-0.277, -0.179, and -0.268, respectively) (Table 47). Results on panicle weight correlated with grain yield conformed with results of Jeyaprakash et al. (1997), Ezeaku and Mohammed (2006), and Deepalakshmi and Ganesamurthy (2007). Panicle weight was significantly positively correlated with plant height, as was reported also by El-Naim et al. (2012).

Thousand-kernel weight was positively correlated with all traits except grain yield for all genotypes in the seven environments and except for fiber concentration in the grain in sorghum in the environment with only parents (Tables 45 and 46). In the hybrid analysis, 1000-kernel weight was significantly correlated with all traits, but with only a 0.05 probability for the number of days to anthesis and fiber concentration in the grain (Table 47). 1000-kernel weight revealed significant negative correlations with grain yield (-0.179), panicle length (-0.327), and protein, fiber, and fat concentration in grain (-0.206, -0.117, and -0.331), respectively) (Table 47). In analysis of hybrids, significant negative correlations were between 1000-kernel weight and grain yield, and 1000-kernel weight and panicle length. Ezeaku and Mohammed (2006) found nonsignificant correlation between 1000-kernel weight and panicle length (0.005), while 1000-kernel weight and grain yield were significantly correlated (0.522). Omar et al. (2014) found

that plant height was significantly positively correlated with grain yield, while 1000-kernel weight was significantly negatively correlated.

Concentration of protein in grain was significantly negatively correlated with concentration of starch in grain across all analyses except commercial hybrid checks, while commercial hybrid checks had significant correlation with 1000-kernel weight (0.544) (Tables 45-51). No significant correlation was found for grain yield and protein across all analyses except commercial hybrid checks which was significantly negative (-0.553) (Table 51). This confirmed earlier reports by El-Hifney et al. (1972), Crook and Casady (1974), El-Gasim (1975), Ross et al. (1981), and Bohra et al. (1985). Rani et al. (2015) reported no correlation between protein and starch concentration in grain.

Means Separation

Yield, with a standard error of 0.4068, was significantly different ($P < 0.0001$) for all the 160 hybrids in the six environments. Yield varied from 3.072 (A.338*RSC76-16 BC₁F_{2:3} high) to 8.21 Mg ha⁻¹ (A.319*RSC83-1 BC₁F_{2:3} low). Lines were significantly different ($P < 0.0001$), with a standard error of 0.2034, with yields ranging from 4.771 (RSC76-13 F_{2:3} low) to 7.048 Mg ha⁻¹ (RSC117-4 BC₁F_{2:3} high). Testers were significantly different ($P < 0.0001$), with a standard error of 0.0646, with yields ranging from 5.429 (A.338) to 6.073 Mg ha⁻¹ (A.301). Line x tester interaction was significant ($P < 0.0001$), with a standard error of 0.4068. Yields were high among testers, with A.319 ranging from 4.68 (RSC76-2 BC₁F_{2:3} low) to 8.21 Mg ha⁻¹ (RSC83-1 BC₁F_{2:3} low), followed by A.338 ranging from 3.072 (RSC76-16 BC₁F_{2:3} high) to 8.06 Mg ha⁻¹ (RSC117-4 BC₁F_{2:3} high), A.301 ranging from 4.78 (RSC38-5 F_{2:3} high) to 7.47 Mg ha⁻¹

(RSC83-14 BC₁F_{2:3} high), and A.Tx3197 ranging from 3.77 (RSC76-13 F_{2:3} low) to 7.003 Mg ha⁻¹ (RSC112-5 F_{2:3} high). Line x environment interaction was significantly different to at least $P < 0.002$, with a standard error of 0.4982, indicating little variability in the standard error among environments. Yield was least at Taylor, TX, in 2015, varying from 1.557 (RSC15-14 BC₁F_{2:3} low) to 3.82 Mg ha⁻¹ (RSC112-5 F_{2:3}), followed by Vega, TX, in 2015, varying from 3.215 (RSC76-16 BC₁F_{2:3} high) to 7.88 Mg ha⁻¹ (RSC117-4 BC₁F_{2:3} high), Vega in 2016, ranging from 4.73 (RSC124-4 BC₁F_{2:3} low) to 7.882 Mg ha⁻¹ (RSC112-19 BC₁F_{2:3} high), Hutchinson, KS, in 2016, ranging from 4.052 (RSC38-5 BC₁F_{2:3} high) to 7.962 Mg ha⁻¹ (RSC124-16 BCF_{2:3} high), Dumas, TX, in 2016, ranging from 5.30 (RSC38-15 F_{2:3} high) to 8.353 Mg ha⁻¹ (RSC83-1 BC₁F_{2:3} low), and the greatest yield at Hutchinson in 2015, ranging from 5.72 (RSC76-16 BCF_{2:3} high) to 10.757 Mg ha⁻¹ (RSC124-9 F_{2:3} high). Tester x environment interaction was significantly different ($P < 0.0001$), with a standard error of 0.1595 without variation among the environments. High grain yields were produced at Hutchinson in 2015, ranging from 7.50, 8.13, 8.42, and 8.60 Mg ha⁻¹ for A.Tx3197 A.338, A.319, and A.301, respectively. A.301 yielded most at Vega in 2015, followed by A.319, A.338, and A.Tx3197, with yields of 6.28, 5.88, 5.03, and 4.91 Mg ha⁻¹, respectively. At Hutchinson in 2016 and Vega in 2016, A.319 yielded most, while at Dumas in 2016, A.301 yielded the most, followed by A.319. At Taylor in 2015, record low yields were obtained from all testers A.338, A.Tx3197, A.301, and A.319, with 2.23, 2.34, 2.64, and 2.79 Mg ha⁻¹, respectively. The lowest yields were produced at Taylor, which could be attributed to low nitrogen in the soil, as reported by local farmers and fellow researchers. Late

planting at Hutchinson in 2015 might have contributed to the low yield of A.338 hybrids (Table 2).

Ten RSC families (RSC15, 19, 37, 38, 73, 76, 83, 112, 117, and 124) from different counties of origin, races, working groups, and their respective selections with percentage of exotic genome recovered in the research are presented in Table 1. Hybrids with RSC15 significantly differed with a greater mean (5.43 Mg ha^{-1}) than that of RSC76 (4.85 Mg ha^{-1}), and significant differences with a lesser mean with RSC37, 83, 112, 117, and 124, with grain yield means of 5.95, 6.22, 6.29, 6.29, and 5.98 Mg ha^{-1} , respectively (Table 53). RSC19 (5.65 Mg ha^{-1}) significantly differed from RSC76, 83, 112, and 117, while RSC37 differed significantly from RSC38 and 76. RSC38 differed significantly from RSC83, 112, 117, and 124, while RSC73 was significantly different from RSC76, 83, 112, and 117. RSC76 differed significantly from RSC83, 112, 117, and 124. The significant differences confirmed the variability already mentioned in the research. Hybrids of RSC83, 112, and 117 yielded most; however, not significantly more than each other (6.21 , 6.29 , and 6.29 Mg ha^{-1} , respectively). The low grain yield was observed for hybrids of RSC76 (4.85 Mg ha^{-1}). Grain yield was significantly different for all 10 RSC families for the RSC * environment interaction ($P < 0.0001$), with a standard error from 0.2762 to 0.2858 based on environment. RSC112 hybrids had the high grain yield at Hutchinson in 2015 and Vega in 2015 and 2016, with 8.88, 7.27, and 6.83 Mg ha^{-1} , respectively. RSC37 hybrids yielded most at Hutchinson in 2016 and Taylor in 2015, with yields of 6.68 and 3.03 Mg ha^{-1} , respectively, while RSC83 hybrids ranked the highest at Dumas in 2016, with a yield of 7.60 Mg ha^{-1} . RSC76 hybrids at Vega in 2015

Table 53. Mean and *P*-values from pairwise *t*-test comparing differences in the RSC families (15, 19, 37, 38, 73, 76, 83, 112, 117, and 124) represented in this research for 160 hybrids across six locations - Taylor and Vega, TX, and Hutchinson, KS, in 2015, and Dumas and Vega, TX, and Hutchinson, KS, in 2016 for grain yield Mgha⁻¹.

RSC†	15	19	37	38	73	76	83	112	117	124
15		0.1605	0.0012	0.1565	0.128	0.0003	<.0001	<.0001	<.0001	0.0006
19	0.1605		0.0645	0.0047	0.9052	<.0001	0.0004	<.0001	<.0001	0.0418
37	0.0012	0.0645		<.0001	0.0836	<.0001	0.0952	0.0318	0.031	0.8496
38	0.1565	0.0047	<.0001		0.0032	0.0291	<.0001	<.0001	<.0001	<.0001
73	0.128	0.9052	0.0836	0.0032		<.0001	0.0007	0.0001	0.0001	0.0553
76	0.0003	<.0001	<.0001	0.0291	<.0001		<.0001	<.0001	<.0001	<.0001
83	<.0001	0.0004	0.0952	<.0001	0.0007	<.0001		0.6269	0.6228	0.1398
112	<.0001	<.0001	0.0318	<.0001	0.0001	<.0001	0.6269		0.9965	0.0506
117	<.0001	<.0001	0.031	<.0001	0.0001	<.0001	0.6228	0.9965		0.0494
124	0.0006	0.0418	0.8496	<.0001	0.0553	<.0001	0.1398	0.0506	0.0494	

RSC†	Grain Yield Mgha ⁻¹
112	6.29
117	6.29
83	6.21
124	5.98
37	5.95
73	5.67
19	5.65
15	5.43
38	5.20
76	4.85

†RSC represents the different Reinstated Sorghum Conversion (RSC) lines and their respective families used in the study.

and at Hutchinson in 2016 and 2015 yielded least, with 3.64, 5.06, and 6.75 Mg ha⁻¹, respectively. RSC38 hybrids yielded least at Dumas in 2016, while RSC15 hybrids were least at Taylor in 2015 and Vega in 2016, with yields of 5.72, and 1.90, and 5.18 Mg ha⁻¹, respectively. Grain yield was significantly different for the RSC * tester interaction (*P* < 0.0001), and standard error ranged from 0.2255 to 0.2282 depending on the environment. The small grain yield for RSC76 with A.338, A.Tx3197, and A.301 was 3.62, 4.39, and

5.31 Mg ha⁻¹, respectively. Small grain yield in A.301 was observed with RSC38 (5.56 Mg ha⁻¹). High grain yield varied by tester and RSC family; A.338 yielded most with RSC117 (6.87 Mg ha⁻¹), followed by A.319 with RSC83 (6.78 Mg ha⁻¹), A.Tx3197 with RSC112 (6.48 Mg ha⁻¹), and A.301 with RSC19 (6.40 Mg ha⁻¹). *P*-values for combined percentage of exotic genome recovery for RSC families are presented in Table 54.

The sorghum selections in this research consisted of the high and low percentage of exotic genome recovery and the generation: BC₁F_{2:3} high, BC₁F_{2:3} low, F_{2:3} high, and F_{2:3} low. The BC₁F_{2:3} high differed significantly from the F_{2:3} low with a *P* < 0.0001, while the significant difference with F_{2:3} high was *P* < 0.03601 (Table 55). The BC₁F_{2:3} low and F_{2:3} high generation differed significantly from the F_{2:3} low (*P* < 0.0001), while the F_{2:3} low significantly differed (*P* < 0.0001) from all three other generation selections. The advantage of the additional backcross with the high percentage of exotic genome

Table 54. *P*-values and means of the percentage of exotic genome recovered value (%) for 160 sorghum hybrids classified by RSC family based on combined analysis across six environments, Vega, TX, in 2015 and 2016, Hutchinson, KS, in 2015 and 2016, Taylor, TX, in 2015, and Dumas, TX, in 2016.

RSC†	Value of exotic genome recovered	RSC†	15	19	37	38	73	76	83	112	117	124
15	0.39875	15		<.0001	0.4284	0.0001	0.0071	<.0001	0.0521	0.0727	0.2862	0.7675
19	0.286	19	<.0001		<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	0.0002	<.0001
37	0.4175	37	0.4284	<.0001		0.002	0.0574	<.0001	0.2498	0.3158	0.0632	0.2768
38	0.49075	38	0.0001	<.0001	0.002		0.2328	0.1311	0.0521	0.0366	<.0001	<.0001
73	0.4625	73	0.0071	<.0001	0.0574	0.2328		0.0069	0.4534	0.3694	0.0002	0.0028
76	0.5265	76	<.0001	<.0001	<.0001	0.1311	0.0069		0.0006	0.0003	<.0001	<.0001
83	0.44475	83	0.0521	<.0001	0.2498	0.0521	0.4534	0.0006		0.8825	0.0026	0.0253
112	0.44125	112	0.0727	<.0001	0.3158	0.0366	0.3694	0.0003	0.8825		0.0043	0.0366
117	0.3735	117	0.2862	0.0002	0.0632	<.0001	0.0002	<.0001	0.0026	0.0043		0.4408
124	0.39175	124	0.7675	<.0001	0.2768	<.0001	0.0028	<.0001	0.0253	0.0366	0.4408	

† RSC represents the different Reinstated Sorghum Conversion (RSC) lines and their respective families used in the study.

Table 55. *P*-values and mean grain yield Mgha⁻¹ for 160 sorghum hybrids classified by generation and the percentage of recovered exotic genome: High BC₁F_{2:3} or F_{2:3} and Low BC₁F_{2:3} or F_{2:3} based on combined analysis across six environments, Vega, TX, in 2015 and 2016, Hutchinson, KS, in 2015 and 2016, Taylor, TX, in 2015, and Dumas, TX, in 2016. *P*-values are presented as BCF_{2:3} High and Low percentage of exotic genome recovered and F_{2:3} High and Low percentage of exotic genome recovered.

Selections [†]	BC ₁ F _{2:3} High	BC ₁ F _{2:3} Low	F _{2:3} High	F _{2:3} Low
BC ₁ F _{2:3} High		0.3076	0.0301	<.0001
BC ₁ F _{2:3} Low	0.3076		0.2519	<.0001
F _{2:3} High	0.0301	0.2519		<.0001
F _{2:3} Low	<.0001	<.0001	<.0001	

Selections [†]	Grain yield (Mgha ⁻¹)
BC ₁ F _{2:3} High	6.02
BC ₁ F _{2:3} Low	5.91
F _{2:3} High	5.78
F _{2:3} Low	5.30

[†]Selections are equal to the high and low percentage of exotic genome recovery and the generation: High F_{2:3}, BCF_{2:3}, Low F_{2:3}, and BCF_{2:3}.

recovery contributed to the significant increase in grain yield over that of the F_{2:3} generation. There was no difference in the high and low percentage of exotic genome recovery in the backcross generation, thus confirming that the additional backcross generation was beneficial to the sorghum breeder. However, for early testing, a combination of resources, including genomic recovery, would benefit a sorghum breeder. The interaction for selection x environment was significant at $P < 0.001$, with a standard error from 0.1893 to 0.1906 based on environment. The BC₁F_{2:3} high yielded most grain, while F_{2:3} low yielded least grain in the environments of Hutchinson in 2015 and 2016, Vega and Dumas in 2016, and Taylor in 2015. Vega in 2015 had the high grain yield BC₁F_{2:3} low, with the low grain yield F_{2:3} high. Vega in 2015 was planted later than

usual because of additional rainfall in May (Table 2). With the later planting date, hybrids required fewer days before anthesis as compared to other environments. The environment plays an integral component when selecting hybrids. *P*-values for combined percentage of exotic genome recovery for selections are presented in Table 56.

Table 56. *P*-values and means of the percentage of exotic genome recovered value (%) for 160 sorghum hybrids classified by RSC selections based on combined analysis across six environments, Vega, TX, in 2015 and 2016, Hutchinson, KS, in 2015 and 2016, Taylor, TX, in 2015, and Dumas, TX, in 2016. *P*-values presented.

Selections†	Value of exotic genome recovered	Selections	BC ₁ F _{2:3} High	BC ₁ F _{2:3} Low	F _{2:3} High	F _{2:3} Low
BC ₁ F _{2:3} High	0.7	BCF _{2:3} High		<.0001	<.0001	<.0001
BC ₁ F _{2:3} Low	0.509	BCF _{2:3} Low	<.0001		<.0001	<.0001
F _{2:3} High	0.3597	F _{2:3} High	<.0001	<.0001		<.0001
F _{2:3} Low	0.1246	F _{2:3} Low	<.0001	<.0001	<.0001	

†Selections are equal to the high and low percentage of exotic genome recovery and the generation: High F_{2:3}, BCF_{2:3}, Low F_{2:3}, and BCF_{2:3}.

Chapter V

Conclusions

In summary, A.301*RSC83-14 BC₁F_{2:3} high had the third-highest mean of overall grain yield for hybrids, yielding slightly more than the commercial check 301/41, with 67.3 days to anthesis and 127.5 cm total plant height. The hybrid had significant MPH with a positive SCA effect and GCA combining effect combination of high x high. The hybrid would have a good combination for earliness, dwarfing genes, and high yield. A.319*RSC83-1 F_{2:3} high had the largest overall mean for grain yield, out-yielding four of the best six commercial check hybrids with a positive MPH and SCA effect. The hybrid flowered in 72.2 days, with a total plant height of 153.0 cm and a high x low GCA effects combination. Further observations included line RSC112-19 BC₁F_{2:3} high, with large overall mean grain yield among all four testers A.301 (7.22), A.319 (7.10), A.Tx3197 (6.67), and A.338 (6.20 Mg ha⁻¹); SCA effects of 0.124, -0.06, 0.212, and -0.276, with GCA combination effects of (high x high), (high x high), (low x high), and (low x high), respectively, with all exhibiting positive MPH, and corresponding number of days to anthesis and plant height of 71.2, 72.2, 70.8, and 75.5 days and 116.1, 131.9, 146.8, and 157.8 cm.

Selection of sorghum parental lines with larger percentage of exotic genome recovery in the F_{2:3} produced greater yielding F₁ hybrids over those with smaller percentage of exotic genome recovery. It is possible to improve grain yield and

important agronomic traits using the high percentage of exotic genome recovery in the F_2 population. This research aimed to develop earlier hybrids through evaluation of F_1 hybrids produced from exotic germplasm within the generation of $F_{2:3}$'s or $BC_1F_{2:3}$'s and to understand the nature of gene action involved in control of grain yield and its components. The significant differences confirmed the variability already mentioned in the research. Hybrids of RSC83, 112, and 117 yielded most, but not significantly different from each other (6.21, 6.29, and 6.29 Mg ha⁻¹, respectively). The low grain yield was by hybrids of RSC76 (4.85 Mg ha⁻¹). Grain yield was significantly different for all 10 RSC families for the RSC * environment interaction ($P < 0.0001$), with a standard error from 0.2762 to 0.2858 based on environment.

After removing the NIR data, combining ability estimates of SCA variance were greater than GCA variance, i.e., the ratio of GCA to SCA variances was less than unity for all traits except plant exsertion, which showed that non-additive gene action was dominant in the inheritance of all traits studied except panicle length. The results were supported by the ratio of variance of general to specific combining ability ($\sigma^2_{gca}/\sigma^2_{sca}$) which was smaller than unity and by the degree of dominance (σ^2_D/σ^2_A) that requires values greater than unity for all traits except plant exsertion. Therefore, superior hybrids can be developed through exploitation of heterosis.

Analysis of GCA revealed that among the parents, lines RSC117-4 $BC_1F_{2:3}$ high, RSC83-1 $BC_1F_{2:3}$ low, RSC83-14 $BC_1F_{2:3}$ high, and RSC112-19 $BC_1F_{2:3}$ high, and the testers A.301 and A.319 were promising general combiners for grain yield and most of the traits. Therefore, the parents can be used in sorghum breeding programs to develop

high-yielding hybrids. The family of RSC15 was a good general combiner for concentration of protein in grain, RSC112 for concentration of starch in grain, and RSC117-4 BC₁F_{2:3} high for fiber concentration in grain.

Based on MPH, specific agronomic traits of interest, and SCA effects of the hybrids, five hybrids were identified as good combinations for grain yield:

A.319*RSC83-1 BC₁F_{2:3} low (high x high), A.338*RSC117-4 BC₁F_{2:3} high (low x high), A.301*RSC83-14 BC₁F_{2:3} high (high x high), A.301*RSC112-19 BC₁F_{2:3} high (high x high), and A.319* RSC112-19 BC₁F_{2:3} high, while considering the combination effect of GCA. The hybrid A.301*RSC83-14 BC₁F_{2:3} high had significant positive MPH for grain yield, total plant height, height to the flag leaf, panicle length, three-panicle weight, and 1000-kernel weight. The hybrids A.319*RSC83-1 BC₁F_{2:3} low, A.338*RSC117-4 BC₁F_{2:3} high, A.301*RSC83-14 BC₁F_{2:3} high, A.319*RSC19-10 BC₁F_{2:3} low, and A.301*RSC112-19 BC₁F_{2:3} high were taller and yielded more than the parents, with significantly greater MPH *per se* parents. Line RSC19-17 BC₁F_{2:3} high when crossed to A.319 and A.338 had significantly positive MPH for protein in the grain but significantly negative MPH for starch in the grain. Crosses with high x low or low x high GCA effects of parents indicated the presence of additive x dominance type of gene interaction. Therefore, the crosses might produce desirable transgressive segregates because of an additive genetic system in one general combiner of the parent and complimentary epistatic effects in the other. Tester A.301 was observed to promote earliness and dwarfing genes while A.319 produced greater three-panicle weight and 1000-kernel weight.

In addition, understanding the nature of association among grain yield, yield components, and agronomic traits was emphasized. Significant positive correlation coefficients were found for grain yield with panicle length, three-panicle weight, and concentration of fiber and fat in grain, while significant negative correlations were observed for the number of days to anthesis, total plant height, and height to the flag leaf. Correlation association between SCA and MPH for yield and agronomic traits of the 160 sorghum hybrids were significant for all variables analyzed.

For early testing, the percentage of exotic genome recovery would be a beneficial option for selecting in the $F_{2:3}$ generation over the $BC_1F_{2:3}$ generation. Selection in the $F_{2:3}$, revealed the family of RSC112, 117, and 73 on the basis of GCA effects within the $F_{2:3}$ generation with ranks of 2, 3, and 4, respectively; however, RSC83 could have been missed based solely on the $F_{2:3}$ generation, while RSC124 could have been selected. RSC83 benefited from the additional backcross generation while RSC124 did not. A plant breeder relies on a combination of resources to aid in selection for early testing. The high-yielding RSC families were 117, 112, and 83. Significant differences were found in selection generation, the $F_{2:3}$ low significantly differed from the other three ($BC_1F_{2:3}$ high, low, and $F_{2:3}$ high), while the value of exotic genome recovery would provide insight into the early selection process because grain yield of the highest $F_{2:3}$ significantly differed from that of the lowest $F_{2:3}$. The backcross generation significantly improved grain yield; however, stricter selection in the $F_{2:3}$, with available resources, could potentially reduce the workload in the future backcross generation. Relying on the GCA effects in the combination of hybrids provides insight as previously discussed;

however, the low x low combination might be suitable for selection in later generations while the high x high combination would provide complimentary gene interaction. The magnitude of the research provides insight into early testing that would provide an opportunity for a sorghum breeder to select the high percentage of genome recovery in the $F_{2:3}$ generation, thus potentially reducing the number of hybrids to be evaluated and also the cost associated with phenotyping a large number of hybrids in the field. From the study, it can be concluded that selection in the $F_{2:3}$ generation, with the aid of selecting the most genome recovery materials over the low genome recovery, would provide insight for greater yielding hybrids with early maturity and dwarfing genes before the backcross generation. Although hybrid-breeding technology has been a great success in increasing yields in many cereal crops including sorghum, the process of developing and evaluating the performance of hybrids is the most expensive and time-consuming activity. Developing sorghum parental inbred lines and evaluating their potential hybrid performance are very expensive and time-consuming.

Future research should include an understanding of the RSC families that were 2 or 3-dwarf and separate the hybrids into these categories for forage or grain harvest, respectively, and harvest accordingly. The amount of total dry matter would provide insight into limitation by mechanical harvesting of grain, thus allowing calculation for harvest index. Population and tiller counts could aid in explaining negative correlations found in lines and hybrids. The Reinstated Sorghum Conversion Program strives to substitute recessive Ma_1 and Dw_2 and Dw_3 , with additional knowledge of Ma_5 and Ma_6 . Additional substitution of these height and maturity genes might provide insight into

earlier populations with variable height. More research into the sorghums in the Reinstated Sorghum Conversion Program is needed to facilitate more diversity by crossing Caudatum and Durra varieties. The understanding of RSC76 being tall, late, and having large kernels gives insight into its race being a Caudatum-Durra type. RSC73 and 83 are Caudatum while RSC112 and 117 are Durra, and with a potential to cross these races might give rise to additional diversity within the R- and B-lines for sorghum breeding programs. Further investigation of the fertility of these exotic lines would provide sorghum breeders with more diversity in elite inbreds. The magnitude of this research would need to be simplified to a smaller scale for constraints in resources.

References

- Abdel-Mottaleb, A.A. 2009. Heterosis and combining ability in grain sorghum (*Sorghum bicolor* L. Moench) under optimum and low level of nitrogen. Ph.D. thesis, Faculty of Agriculture, Assiut University, Egypt.
- Abe, K., Y. Sato, T. Saito, A. Kurihara, and K. Kotobuki. 1995. Narrow-sense heritability of fruit characters in Japanese pear (*Pyrus pyrifolia* Nakai). Japanese J. Breeding 45:1-5.
- Abou-Amer, A. and K. Kewan. 2014. Effect of NP fertilization levels on sorghum (*Sorghum bicolor* L.) yield and fodder quality for animals. Alex. J. Agric. Res. 59:51-59.
- Al-Kaisi, M.M., X. Yin. 2003. Effects of nitrogen rate, irrigation rate, and plant population on corn yield and water use efficiency. Agron. J. 95:1475-1482.
- Allison, J.C.S., and R.N. Curnow. 1966. On the choice of tester parent for the breeding of synthetic varieties of maize (*Zea mays* L.). Crop Sci. 6:541-544.
- Almeida Filho, G.E., F.D. Tardin, M.D. Vilela de Resende, F.F. Silva, I.S.C. Granato, and C.B. de Menezes. 2014. Genetic evaluation of grain sorghum hybrids in Brazilian environments using the REML/BLUP procedure. Sci. Agric. 71(2).
- Amir, A.A., and E.I. Mohamed. 2015. Inheritance of sorghum yield and its components under low nitrogen fertilizer using line by tester analysis. Egypt. J. Plant Breed. 19:1117-1131.
- Amsalu, A.A., and D.R. Bapat. 1990. Heterosis studies in sorghum. J. Maharashtra Agric. Univ. 15:299-302.
- Aruna, C., S. Audilakshmi, and C.D. Reddy. 2010. Sorghum germplasm for yield improvement and evaluation of sorghum [*Sorghum bicolor* (L.) Moench] germplasm lines for their yield components. Indian J. Agric. Sci. 80:409-412.
- Arunkumar, B., B.D. Biradar, and P.M. Salimath. 2004. Genetic variability and character association studies in rabi sorghum. Karnataka J. Agric. Sci. 17:471-475.
- Assefa, Y., and S.A. Staggenborg. 2010. Grain sorghum yield with hybrid advancement and changes in agronomic practices from 1957 through 2008/ Agron. J. 102:703-706.

Assefa, Y., S.A. Staggenborg, and V.P.V. Prasad. 2010. Grain sorghum water requirement and responses to drought stress: a review. Online. Crop Management doi: 10.1094/CM-2010-1109-01-RV.

Assefa, Y., K. Roozeboom, C. Thimpson, A. Schlegel, L. Stone, and J. Lingenfelser. 2013. Corn and grain sorghum comparison all things considered. <https://books.google.com/books?hl=en&lr=&id=N3spAgAAQBAJ&oi=fnd&pg=PP1&dq=%E2%80%9CCorn+and+Grain+Sorghum+Comparison%E2%80%9D+by+Yared+Assefa,+Kraig+Roozeboom,+Curtis+Thompson,+Alan+Schlegel,+Lloyd+Stone+and+Jane+Lingenfelser&ots=I-NXeOdG-n&sig=ITbKoAkfD44DZRymR727NaszW-M> (accessed 13 July 2017).

Audilakshmi, S. and C. Aruna. 2005. Genetic analysis of physical grain quality characters in sorghum. FAO. gris.fao.org/agris-search/search.do?recordID=US201301037413 (accessed 13 June 2017).

Badhe, P.L., and H.S. Patil. 1997. Line x tester analysis in sorghum. *Ann. Agric. Res.* 18:281-284.

Badwal, S.S. 1970. Correlation between grain and fodder yield in jowar. *Madras Agric. J.* 58:531-533.

Bean, B., T. McCollum, M. Rowland, and K. McCuiston. 2003. Seeding and nitrogen rate effect on forage yield, lodging, and nutrient composition of two brown midrib forage sorghums. Pub. AREC 4-04. Texas Cooperative Extension and Texas Agricultural Experiment Station, College Station, TX.

Bean, B., and T. McCollum. 2006. Summary of six years of forage sorghum variety trials. Pub. SCS-2006-04. Texas Cooperative Extension and Texas Agricultural Experiment Station, College Station, TX.

Bean, B.W., T.F. McCollum, B. Villareal, J. Robinson, E. Buttrey, R. VanMeter, and D. Pietsch. 2007. 2007 Texas Panhandle forage sorghum silage trial. Texas Cooperative Extension and Texas Agricultural Experiment Station, Amarillo, TX. <http://animalscience.tamu.edu/images/pdf/eef/beef-2007/pdf>

Beil, G.M., and R.E. Atkins. 1967. Estimates of general and specific combining ability in F1 hybrids for yield and its components in grain sorghum, *Sorghum vulgare* Pers. *Crop Sci.* 7:224-228.

Bennett, J. 2003. Environmental values and water policy. *Geographical Res.* 41:237-250.

Berenji, J. 1988. Evaluation of combining ability and heterosis and analysis of yield components in grain sorghum. *Bilten za Hmelj, Sirak i Lekovito Bilje* 20:56-57.

- Bernardo, R. 1992. Relationship between single-cross performance and molecular marker heterozygosity. *Theor. Appl. Genet.* 83:628-634.
- Bernardo, R. 2012. Breeding for quantitative traits in plants. 2nd ed. Stemma Press, Woodbury, MN.
- Bernardo, R. 2014. Essentials of plant breeding. Stemma Press, Woodbury, MN.
- Biradar, B.D., S.M. Vastrad, R.A. Balikai, and J.M. Nidagundi. 2000. Combining ability studies in *rabi* sorghum. *Karnataka J. Agric. Sci.* 13:721-723.
- Blum, A. 1969. Nature of heterosis in grain production by the sorghum panicle. *Crop Sci.* 10:28-31.
- Blum, A., W.R. Jordan, and G.F. Arkin. 1977. Sorghum root morphogenesis and growth. II. Manifestation of heterosis. *Crop Sci.* 17:153-157.
- Blum, A., and Y. Pnuel. 1990. Physiological attributes associated with drought resistance of wheat cultivars in a Mediterranean environment. 1990. *Aust. J. Agric. Res.* 41:799-810.
- Bohra, P.S.P., P.S. Phul, and A. Rang. 1985. Association analysis for yield and quality traits in sorghum. *Crop Improv.* 12:89-93.
- Bolaños, J., and G.O. Edmeades. 1993. Eight cycles of selection for drought tolerance in lowland tropical maize. II. Responses in reproductive behavior. *Field Crops Res.* 31:253-268.
- Bowling, R.D., M.J. Brewer, D.L. Kerns, J. Gordy, N. Seiter, N.E. Elliott, M.O. Way, T.A. Royer, S. Biles, and E. Maxson. 2016. Sugarcane aphid (Hemiptera: Aphididae): a new pest on sorghum in North America. *J. Integr. Pest Manag.* 7:12.
- Boyer, J.S. 1987. Water and plant productivity. In: W.R. Jordan [ed.] *Water and water policy in world food supplies: Proceedings of the Conference*, Texas A&M University, College Station, TX. p. 233-239.
- Briggs, F.N., and P.F. Knowles. 1967. Introduction to plant breeding. Chapman-Reinhold, Inc.
- Bruce, A.B. 1910. The Mendelian theory of heredity and the augmentation of vigor. *Science* 32:627-628.
- Burns, D., and E. Ciurczak [eds.]. 2007. Handbook of near-infrared analysis. 3rd ed. (Practical Spectroscopy). p. 349-369.

- Butler, T., and B. Bean. 2006. Forage sorghum production guide. www.soilcropandmore.info/...Forage-Sorghum/ForageSorghumProd...
- Can, N.D., S. Nakamura, and T. Yoshida. 1997. Combining ability and genotype x environmental interaction in early maturing grain sorghum for summer seeding. *Japanese J. Crop Sci.* 66:698-705.
- Can, N.D., T.A.D. Haryanto, and T. Yoshida. 1998. Genetic variability and characteristic association analysis in grain sorghum. *J. Faculty of Agric.* 43:25-30.
- Can, N.D., and T. Yoshida. 1999. Genotypic and phenotypic variances and covariances in early maturing grain sorghum in a double cropping. *Pl. Prod. Sci.* 2:67-70.
- Carpenter, S.R., E.H. Stanley, and M.J.V. Zanden. 2011. State of the world's freshwater ecosystems: physical, chemical, and biological changes. *Annu. Rev. Environ. Resour.* 36:75-99.
- Carr, D.E., and M.R. Dudash. 2003. Recent approaches into the genetic basis of inbreeding depression in plants. *Phil. Trans. R. Soc. Lond. B* 358:1071-1084.
- Charlesworth, B., and D. Charlesworth. 1999. The genetic basis of inbreeding depression. *Genet. Res.* 74:329-340.
- Chaudhary, S.B., J.V. Patil, B.B. Thombare, and V.M. Kulkarni. 2006. Selection of parents based on combining ability in sorghum [*Sorghum bicolor* L. Moench]. *Ann. Plant Physiol.* 20:95-97.
- Chaudhuri, U.N., and E.T. Kanemasu. Growth and water use of sorghum (*Sorghum bicolor* (L.) Moench) and pearl millet (*Pennisetum americanum* (L.) Leeke. *Field Crops Res.* 10:113-124.
- Chavan, S.K., R.C. Mahajan, and S.U. Fatak. 2011. Correlation and path analysis studies in sorghum. *Crop Res.* 42:246-250.
- Cockerham, C.C., and Z.-B. Zeng. 1996. Design III with marker loci. *Genet.* 143:1437-1456.
- Colaizzi, P.D., P.H. Gowda, T.H. Marek, and D.O. Porter. 2008. Irrigation in the Texas High Plains: a brief history and potential reductions in demand. *Irrigation Drainage* 58:257-274.
- Collins, G.N. 1921. Dominance and the vigor of first generation hybrids. *Am. Nat.* 55:116-133.

Comstock, R.E., H.F. Robinson, and P.H. Harvey. 1949. A breeding procedure designed to make maximum use of both general and specific combining ability. *Agron. J.* 41:360-367.

Conkwright, J. 2012. The Ogallala Aquifer. High Plains Underground Water Conservation District 1 [online]. http://www.hpwd.com/the_ogallala.asp (accessed 9 Oct. 2012).

Cress, C.E. 1966. Heterosis of the hybrid related to gene frequency differences between two populations. *Genet.* 53:269.

Crook, W.T., and A.J. Casady. 1974. Heritability and interrelationships of grain protein content with other agronomic traits of sorghum. *Crop Sci.* 14:622-624.

Crow, J.F. 1948. Alternative hypotheses of hybrid vigor. *Genet.* 33:477-487.

Crow, J.F. 1952. Dominance and overdominance. In: *Heterosis*. Iowa State College Press, Ames, IA. P.282-297.

Dahlberg, J.A. 2000. Classification and characterization of sorghum. In: C.W. Smith and R.A. Frederiksen [eds.], *Sorghum: origin, history, technology, and production*. John Wiley and Sons, New York. p. 99-130.

Davenport, C.B. 1908. Degeneration, albinism and inbreeding. *Science* 28:454-455.

De Wet, J.M.J., and E.G. Price. 1976. Plant domestication and indigenous African agriculture. In: J.R. Harlan, J.M.J. de Wet, and A. Stemler [eds.], *Origins of African plant domestication*. Mouton, The Hague, Netherlands. p. 453-464.

Degu, E., A. Debello, and K. Belete. 2009. Combining ability study for grain yield and yield related traits of grain sorghum [*Sorghum bicolor* (L.) Moench] in Ethiopia. *Acta Agronomica Hungarica* 57:175-184.

Desai, M.S., and P.T. Shukla. 1995. Genetic analysis of yield characters in grain sorghum. *Gujarat Agric. Univ. Res. J.* 21:83-87.

Dillon, S.L., F.M. Shapter, R.J. Henry, G. Cordeiro, L. Izquierdo, and S. Lee. 2007. Domestication to crop improvement: genetic resources for *Sorghum* and *Saccharum* (Andropogoneae). *Ann. Bot.* 100:975-989.

Doggett, H. 1970. *Sorghum*. Longmans Greens, London.

Doggett, H. 1988. *Sorghum*. 2nd ed. Longmans Greens, London. Published in the United States with Wiley, New York.

- Donald, C.M. 1962. In search of yield. J. Aust. Inst. Agric. Sci. 28:171-178.
- Dudensing, J., J. Johnson, P. Johnson, and C. Villalobos. 2008. Grazing alternatives in the face of declining groundwater: a case from the southern High Plains of Texas. Texas J. Agric. Nat. Res. 21:60-72.
- Duncan, R.R., P.J. Bramel-Cox, and F.R. Miller. 1991. Contributions of introduced sorghum germplasm to hybrid development in the USA. In: Use of plant introductions in cultivar development. Part 1. Special Publ. 17. Crop Science Society of America, Madison, WI. p. 69-102.
- East, E.M. 1908. Inbreeding in corn. Rep. Conn. Agric. Exp. Stn. 1907:419-428.
- El-Dardeer, A.E., M.A. El-Morshidy, A.M. Mahmoud, and H.I. Ali. 2011. Heterosis and combining ability in grain sorghum [*Sorghum bicolor* (L.) Moench] under normal and water stress conditions. 5th Conference of Young Scientists, Faculty of Agriculture. 8 May 2011. Assiut Journal of Agricultural Sciences 442.
- El-Gasim, E.H.A. 1975. Variety and interrelations among characters in indigenous grain sorghums of the Sudan. E. Afr. Agric. For. J. 41:125-133.
- El-Hifney, M.Z., E.S. Kaseem, M. El-Ghavas, and M.K. El-Tohami. 1972. Variability in morphological characters, yield components, quality characters of grain sorghum. Assiut J. Agric. Sci. 3:65-84.
- El-Menshawi, M.M. 2005. Stability and combining ability analysis for grain sorghum hybrids and their parental lines. Bulletin of Faculty of Agriculture, Cairo University 56:271-293.
- El-Mottaleb, A.A.A., and M.R.A. Asran. 2004. Heterosis and combining ability in grain sorghum [*Sorghum bicolor* (L.) Moench]. Assiut J. Agric. Sci. 35:165-183.
- El-Naim, A.M., I.M. Ibrahim, M.E.A. Rahman, and E.A. Ibrahim. 2012. Evaluation of some local sorghum (*Sorghum bicolor* L. Moench) genotypes in rain-fed. Int. J. Plant Res. 2:15-20.
- Essa, H.M.H. 2009. Breeding grain sorghum for drought tolerance. M.Sc. thesis, Faculty of Agriculture, Ain Shams University, Egypt.
- Ezeaku, I.E., and S.G. Mohammed. 2006. Character association and path analysis in grain sorghum. Afr. J. Biotech. 5:1337-1340.

Falconer, D.S., and T.F.C. Mackay. 1996. Introduction to quantitative genetics. 4th ed. Longman, Essex, UK.

FAO (Food and Agriculture Organization of the United Nations). 1995. Sorghum and millets in human nutrition. Food, and Agriculture Organization of the United Nations, Rome. <http://www.fao.org/DOCREP/T0818e/T0818E00.htm#Contents> (accessed 21 July 2012).

FAO (Food and Agriculture Organization of the United Nations). 2002. FAOSTAT Database Results. <http://apps1.fao.org> (accessed 2 May 2012).

FAO (Food and Agriculture Organization of the United Nations). 2007. *Sorghum bicolor* (L.) Moench. <http://www.fao.org/AG/aGp/agpc/data/pf00.19.htm> (27 July 2012).

FAO, FAOSTAT. 2011. Food and Agriculture Organization 2011. <http://faostat.fao.org/site/291/default.aspx>.

FAO (Food and Agriculture Organization of the United Nations). 2011. <https://www.theguardian.com/environment/2011/nov/28/un-farmers-produce-food-population>

FAO (Food and Agriculture Organization of the United Nations). 2016. <http://www.indexmundi.com/agriculture/?commodity=sorghum> (accessed 20 May 2016).

FAS (Foreign Ag Service). USDA. 2012. Global Agricultural Trade System (GATS) (accessed 9 May 2014).

FAS (Foreign Ag Service). USDA. 2015. Global Agricultural Trade System (GATS) (accessed 9 May 2014).

Federer, W.T. 1961. Augmented designs with one-way elimination of heterogeneity. *Biometrics* 17:447-473.

Federer, W.T. 2004. Augmented split block experiment design. <https://ecommons.cornell.edu/bitstream/handle/1813/32252/BU-1650-M.pdf?sequence=1> (accessed 21 June 2016).

Federer, W.T., and D. Raghavarao. 1975. On augmented designs. *Biometrics* 31:29-35.

Fellahi, Zine El-Abidine, A. Hannachi, H. Bouzerzour, and A. Boutekrabt. 2013. Line x tester mating design analysis for grain yield and yield related traits in bread wheat (*Triticum aestivum* L.) (accessed 25 April 2017). <https://www.hindawi.com/journals/ija/2013/201851/>

- Fisher, K.S., and G.L. Wilson. 1975. Studies of grain production in *Sorghum bicolor* (L.) Moench: III. The relative importance of assimilate supply, grain growth capacity and transport system. *Aust. J. Agric. Res.* 26:11-23.
- Fisher, R.A. 1985. Number of kernels in wheat crops and the influence of solar radiation and temperature. *J. Agric. Sci. (Cambridge)* 105:447-461.
- Franca, J.G.E.D.E., D.S. Murthy, D. Nicodemos, and L.R. House. 1986. Genetic analysis of some agronomic traits in grain sorghum [*Sorghum bicolor* (L.) Moench]. *Heterosis Revista Brasileira Genetica* 9:659-678.
- Gampawar, A.S., R.M. Zinjarde, and A.S. Ingole. 2002. Evaluation of sorghum cultivars for forage production under rain-fed conditions. *J. Soils Crops* 12:145-146.
- Ganesh, S., A.K.F. Khan, and N. Senthil. 1996. Heterosis studies for grain yield characters in sweet sorghum. *Madras Agric. J.* 83:655-657.
- Gardner, W.R., and H.R. Gardner. 1983. Principles of water management under drought conditions. *Agric. Water Mgmt.* 7:143-155.
- Giriraj, K., and J.V. Goud. 1982. Genetics of yield and panicle components in grain sorghum. *Crop Improv.* 9:111-114.
- Girma, M., A. Amsalu, and B. Ketema. 2010. Combining ability for yield and its components in Ethiopian sorghum [*Sorghum bicolor* (L.) Moench] landraces. *E. Afr. J. Sci.* 4:34-40.
- Goud, J.V., M.D. Kachapur, and M.J. Vasudeva Rao. 1973. Combining ability in kharif sorghum. *Mysore J. Agric. Sci.* 7:369-376.
- Goud, J.V., and B.M. Asawa. 1978. Yield components in sorghum. *Mysore J. Agric. Sci.* 12:265-268.
- Govil, J.N., and B.R. Murty. 1973. Combining ability for yield and quality characteristics in grain sorghum. *Indian J. Genet. Plant Breeding* 33:239- 251.
- Gowda, P.H., J.L. Chavez, P.D. Colaizzi, S.R. Evett, T. Howell, and J.A. Tolk. 2007. Remote sensing based energy balance algorithms for mapping ET: current status and future challenges. *Transactions ASABE* 50:1639-1644.
- Grains. 2007. Sorghum. <http://www.grains.org/page.wv?sectopm=Barley%2C+Corn%26=Sorghum&name=Sorghum> (accessed 5 May 2012).

- Grains. 2011. Sorghum. USDA Foreign Agricultural Service Circular Series: Grain 2007-2011. USDA, Washington, DC.
- Grains. 2015. Sorghum. USDA Foreign Agricultural Service Circular Series: Grain 2007-2015. USDA, Washington, DC. Sorghum - Production and Exports | U.S. Grains Council.
- Grains. 2016. Sorghum. USDA Foreign Agricultural Service Circular Series: Grain 2009-2016. USDA, Washington, DC. Sorghum - Production and Exports | U.S. Grains Council. Global grain production, 2016 | Statistic (accessed 4 January 2017).
- Green, J.M. 1948a. Relative value of two testers for estimating top cross performance in segregating maize progenies. *Agron. J.* 40:45-57.
- Green, J.M. 1948b. Inheritance of combining ability in maize hybrids. *Agron. J.* 40:58-63.
- Griffing, B. 1956. Concept of general and specific combining ability in relation to diallel crossing systems. *Aust. J. Biol. Sci.* 9:463-493. http://www.publish.csiro.au/?act=view_file&file_id=BI9560463.pdf
- Habyarimana, E., D. Laureti, M. de Ninno, and C. Lorenzoni. 2004. Performances of biomass sorghum [*Sorghum bicolor* (L.) Moench] under different water regimes in Mediterranean region. *Industrial Crops Prod.* 20:23-28.
- Hallauer, A.R., and J.B. Miranda, Fo. 1981. Quantitative genetics in maize breeding. Iowa State Univ. Press, Ames.
- Hammer, G.L., and R.C. Muchow. 1994. Assessing climatic risk to sorghum production in water-limited subtropical environments: I. Development and testing of a simulation model. *Field Crops Res.* 36:221-234.
- Hammer, G.L., and I.J. Broad. 2003. Genotype and environment effects on dynamics of harvest index during grain filling in sorghum. *Agron J.* 95:199-206.
- Hanjra, M.A., and M.E. Qureshi. 2010. Global water crisis and future food security in an era of climate change. *Food Policy* 35:365-377.
- Harlan, J.R., and J.M.J. DeWet. 1972. A simplified classification of cultivated sorghum. *Crop Sci.* 12:172-176.
- Hoshmand, S., and A. Rezai. 1997. Type of gene action and heritability estimates for some agronomic traits in seventeen crosses of grain sorghum [*Sorghum bicolor* (L.) Moench]. *Iranian J. Agric. Sci.* 28:69-79.

House, L.R. 1985. A guide to sorghum breeding. International Crops Research Institute for the Semi-Arid Tropics. ICRISAT, Patancheru, India.

Hovny, M.R.A. 2000. Heterosis and combining ability in grain sorghum [*Sorghum bicolor* (L.) Moench]. Assiut J. Agric. Sci. 31:17-30.

Hovny, M.R.A., B.R. Bakheit, E.A. Hassaballa, and A.A. Amir. 2000. Line x tester analysis for combining ability in grain sorghum [*Sorghum bicolor* (L.) Moench]. Assiut J. Agric. Sci. 31:147-161.

Hovny, M.R.A., M.M. El-Menshaw, and O.O. El-Nagouly. 2001. Combining ability and heterosis in grain sorghum [*Sorghum bicolor* (L.) Moench]. Bulletin of Faculty of Agriculture, Cairo-University 52:47-60.

Hovny, M.R.A., K.M. Mahmoud, M.A. Ali, and H.I. Ali. 2005. The effect of environment on performance, heterosis and combining ability in grain sorghum [*Sorghum bicolor* (L.) Moench]. In: Proceedings of 11th Conference of Agronomy. 15-16 November 2005. Agronomy Department, Faculty of Agriculture, Assiut University. p. 689-699.

Hovny, M. R. A., and M.M. El-Dsouky. 2007. Performance of some grain sorghum lines and their hybrids under optimum and low input nitrogen conditions. Assiut J. Agric. Sci. 38:67-90.

Howell, T.A., B.A. Stewart, and D.R. Nielsen. 1990. Relationship between crop production and transpiration, evapotranspiration, and irrigation. In: Irrigation of Agricultural Crops. ASA, Madison, WI. p. 391-434.

Howell, T.A., A.D. Schneider, and D.A. Dusek. 1991. Corn and sorghum in response to LEPA irrigation. SAAE Paper 91-2520/ ASAE, St. Joseph, MI.

Howell, T.A., J.L. Steiner, S.D. Schneider, and S.R. Evett. 1994. Evapotranspiration of irrigated winter wheat, sorghum, and corn. AGRIS.

Howell, T.A., A.Yazar, A.D. Schneider, D.A. Dusek, and K.S. Copeland. 1995. Yield and water use efficiency of corn in response to LEPA irrigation. Am. Soc. Agric. Engineers 38:1738-1347.

Howell, T.A., J.L. Steiner, A.D. Schneider, S.R. Evett, and J.A. Tolk. 1997. Seasonal and maximum daily evapotranspiration of irrigated winter wheat, sorghum, and corn-Southern High Plains. ASAE 40:623-634.

Howell, T.A., S.R. Evett, J.A. Tolk, and A.D. Schneider. 2004. Evapotranspiration of full, deficit-irrigated, and dryland cotton on the Northern Texas High Plains. *J. Irr. Drainage Engineering* 4:277-285.

Howell, T.A., S.R. Evett, J.A. Tolk, K.S. Copeland, P.S. Colaizzi, and P.H. Gowda. 2008. Evapotranspiration of corn and forage sorghum for silage. In: *Soil and water Management Research News*. USDA-ARS Conservation and Production Research Laboratory, Bushland, TX. Wetting Front Newsletter 10 (June 1).

Hua, J., Y. Xing, W. Wu, C. Xu, X. Sun, S. Yu, and Q. Zhang. 2003. Single-locus heterotic effects and dominance by dominance interactions can adequately explain the genetic basis of heterosis in an elite rice hybrid. *Proc. Natl. Acad. Sci. USA* 100:2574-2579.

Indhubala, M., K. Ganesamurthy, and D. Punitha. 2010. Combining ability studies for quality traits in sweet sorghum [*Sorghum bicolor* (L.) Moench]. *Madras Agric. J.* 97:17-20.

Iyanar, K., A. Gopalan, and P. Ramasamy. 2001a. Combining ability analysis in sorghum [*Sorghum bicolor* (L.) Moench]. *Ann. Agric. Res. new series* 22:341-345.

Iyanar, K., A. Gopalan, and P. Ramasamy. 2001b. Correlation and path analysis in sorghum. *Ann. Agric. Res. new series* 22:495-497.

Jackson, D.M., W.R. Grant, and C.E. Shafer. 1980. U.S. Sorghum Industry. National Economics Division; Economics, Statistics, and Cooperative Services; USDA Ag Eco Report 457. naldc.nal.usda.gov/download/CAT80742394/PDF (accessed 4 Jan. 2017).

Jayasudha, S., and D. Sharma. 2009. Combining ability and gene action analysis for yield and its components in rice (*Oryza sativa* L.). *J. Rice Res.* 2:105-111.

Jenson, M.E., and W.H. Sletten. 1965. Evapotranspiration and soil moisture-fertilizer interrelations with irrigated grain sorghum in the Southern High Plains. U.S.D.A. Conserve. Res. Rept. 5.

Jeyaprakash, P., S. Ganapathy, and M.A. Pillai. 1997. Correlation and path analysis in sorghum [*Sorghum bicolor* (L.) Monech]. *Ann. Agric. Res.* 18:309-312.

Jinks, J.L. 1954. Analysis of continuous variation in diallel cross of *Nicotiana rustica* varieties. *Genet.* 39:767-788.

Jones, D.F. 1917. Dominance of linked factors as a means of accounting for heterosis. *Proc. Nat. Acad. Sci. Wash.* 3:310-312.

- Kambal, A.E., and O.J. Webster. 1965. Manifestation of hybrid vigor in grain sorghum and relations among the components of yield, weight per bushel and height. *Crop Sci.* 6:513-516.
- Kamdi, S. R., M.R. Manjare, and K.V. Sushir. 2009. Combining ability analysis of fodder yield and component characters in sweet sorghum. *Indian J. Agric. Res.* 36:2 38-41.
- Kanawade, D.G., R.B. Deshmukh, N.S. Kute, J.V. Patil, and S.R. Dhonde. 2001. Combining ability studies in sorghum. *Indian J. Agric. Res.* 35:56-59.
- Kanbar, O.Z., A. Kanbar, and S. Shehab. 2011. Combining ability and heterosis for some yield traits in sorghum [*Sorghum bicolor* (L.) Moench] using line x tester design. *J. Plant Production, Mansoura University* 2:1009-1016.
- Karper, R.E., and J.R. Quinby. 1937. Hybrid vigor in sorghum. *J. Hered.* 28:83-91.
- Kempthorne, O. 1957. An introduction to genetic statistics. John Wiley and Sons. New York.
- Kenga, R., S.O. Alabi, and S.C. Gupta. 2004. Combining ability studies in tropical sorghum [*Sorghum bicolor* L. Moench]. *Field Crops Res.* 88:251-260.
- Kenga, R., S.O. Alabi, and S.C. Gupta. 2005. Heterosis and combining ability for grain yield and its components in induced sorghum mutants. *Afr. Crop Sci. J.* 13:143-152.
- Khan, M.Q., S.I. Awan, and M.M. Mughal. 2005. Estimation of genetic parameters in spring wheat genotypes under rain-fed conditions. *Indus J. Biol. Sci.* 2:367-367.
- Kimber, C.T. 2000. Orgins of domesticated sorghum and its diffusion to India and China. In: C.W. Smith and R.A. Frederickson [eds.], *Sorghum: origin, history, technology, and production*. John Wiley and Sons, New York. p. 3-98.
- Kishan, A.G., and S.T. Borikar. 1989. Line x tester analysis involving diverse cytoplasmic systems in sorghum. *Plant Breeding* 102:153-157.
- Klein, R.R., F.R. Miller, P.E. Klein, and J.J. Burke. 2013. Registration of partially converted germplasm from 44 accessions of the USDA-ARS Ethiopian and Sudanese sorghum collections. *J. Plant Reg.* 7:368-372.
- Klein, R.R., F.R. Miller, S. Bean, and P. E. Klein. 2016. Registration of 40 converted germplasm sources from the Reinstated Sorghum Conversion Program. *J. Plant Reg.* pp: 1-5. <https://dl.sciencesocieties.org/publications/jpr/pdfs/10/1/57> (accessed 25 April 2017).

- Knowles, T., P. Nordstrom, and W.B. Klemt. 1984. Evaluation of the ground water resources of the High Plains of Texas. Tex. Dep. Water Resource. Rep. 288.
- Kulakarni, V., P.M. Salimath, and M.S. Patil. 2006. Combining ability analysis in *rabi* sorghum [*Sorghum bicolor* (L.) Moench]. Crop Res. 32:455-458.
- Lamm, F.R., L.R. Stone, and D.M. O'Brien. 2007. Crop production and economics in Northwest Kansas as related to irrigation capacity. Appl. Eng. Agric. 23:737-745.
- Lamm, F.R., R.M. Aiken, and A.A. Abou Kheira. 2009. Corn yield and water use characteristics as affected by tillage, plant density, and irrigation. Appl. Eng. Agric. 52:133-143.
- Lemaire, G., and C.Y. Hebert. 1996. Nitrogen uptake capacities of maize and sorghum crops in different nitrogen and water supply conditions. Agronomie 16:231-246.
- Little, T.M., and F.J. Hills. 1978. Agricultural experimentation. John Wiley and Sons, New York.
- Lokapur, R.G. 1997. Heterosis and combining ability studies in sorghum [*Sorghum bicolor* (L.) Moench]. M.Sc. (Ag.) thesis, University of Agricultural Sciences, Dharwad.
- Lu, H., J. Romero-Severson, and R. Bernardo. 2003. Genetic basis of heterosis explored by simple sequence repeat markers in a random-mated maize population. Theor. Appl. Genet. 107:494-502.
- Luo, L.J., Z.-K. Li, H.W. Mei, Q.Y. Shu, R. Tabien, D.B. Zhong, C.S. Ying, J.W. Stansel, G.S. Khush, and A.H. Paterson. 2001. Overdominant epistatic loci are the primary genetic basis of inbreeding depression and heterosis in rice. II. Grain yield components. Genet. 158:1755-1771.
- Lust, D., L.K. Almas, B.A. Stewart, and A.W. Colette. 2009. Evaluating dryland crop/livestock system alternatives for risk management under declining irrigation in the Texas Panhandle. Selected paper for presentation at Southern Agricultural Economics Association Annual Meeting, Atlanta, GA. 31 January-3 February 2009. <http://ageconsearch.umn.edu/bitstream/46843/2/LustAlmas%20SAEA09.pdf>.
- Madhusudhana, R., and S.S. Patil. 1996. Evaluation of sorghum F₂ population for combining ability. New Botanist 23:5-11.
- Mahajan, R.C., P.B. Wadikar, S.P. Pole, and M.V. Dhuppe. 2011. Variability, correlation and path analysis studies in sorghum. Res. J. Agric. Sci. 2:101-103.

- Mahdy, E.E., M.A. Ali, and A.M. Mahmoud. 2011. The effect of environment on combining ability and heterosis in grain sorghum [*Sorghum bicolor* (L.) Moench]. Asian J. Crop Sci. 3:1-15.
- Makanda, I., P. Tongoona, J. Derera, J. Sibiya, and P. Fato. 2010. Combining ability and cultivar superiority of sorghum germplasm for grain yield across tropical low-and mid-altitude environments. Field Crops Res. 116:75-85.
- Mallinath, V., B.D. Biradar, B.M. Chittapur, P.M. Salimath, N. Yenagi, and S.S. Patil. 2004. Variability and correlation studies in pop sorghum. Karnataka J. Agric. Sci. 17:463-467.
- Malm, N.R. 1968. Exotic germplasm use in grain sorghum improvement. Crop Sci. 8.3:295-298.
- Marsalis, M.A. 2006. Sorghum forage production in New Mexico. Guide A-332. New Mexico State University Cooperative Extension Service, Las Cruces, NM.
- Marsalis, M.A. 2011. Advantages of forage sorghum for silage in limited input systems. In: Proceedings, 2011 Western Alfalfa and Forage Conference, Las Vegas, NM. 11-12 December 2011. UC Cooperative Extension, Plant Sciences Department, University of California, Davis, CA. <http://alfalfa.ucdavis.edu>.
- Marsalis, M.A., R.E. Kirksey, R. Flynn, M.K. O'Neill, L.M. Lauriault, and M. Place. 2008. New Mexico 2007 corn and sorghum performance tests. NM Agric. Exp. Stn., Las Cruces, NM. www.cahe.nmsu.edu/pubs/research/agronomy/perf07.pdf.
- Marsalis, M.A., S. Angadi, F.E. Contreras-Govea, and R.E. Kirksey. 2009. Harvest timing and byproduct addition effects on corn and forage sorghum silage grown under water stress. Res. Bull. BL-799. NM Agric. Exp. Stn., Las Cruces, NM.
- Marsalis, M.A., S.V. Angadi, and F.E. Contreras-Govea. 2010. Dry matter yield and nutritive value of corn, forage sorghum, and BMR forage sorghum at different plant populations and nitrogen rates. Field Crops Res. 116:52-57.
- Martin, J.H. 1930. The comparative drought resistance of sorghums. Agron. J. 22:993-1003.
- Martin, J.H. 1936. Sorghum improvement. In: USDA Yearbook of Agri. 1936.
- Martin, J.H., W.H. Leonard, and D.L. Stamp. 1976. Principles of field crop production Macmillan, New York.

- Mattei, R. 1974. El sistema de apareamiento de las poblaciones de sorgo (*Sorghum bicolor* (L.) Moench), 1: Fertilizacion selectiva. Fitotecnica Latinoamericana.
- Matzinger, D. 1953. Comparison of three types of testers for the evaluation of inbred lines of com. Agron. J. 45:493-495.
- Mejza, I. 1998. Characterization of certain split-block designs with a control. Biometrical J. 40:627-639.
- Menz, M.A, R.R. Klein, N.C. Unruh, W.L. Rooney, P.E. Klein, and J.E. Mullet. 2004. Genetic diversity of public inbreds of sorghum determined by mapped AFLP and SSR markers. Crop Sci. 44:1236-1244.
- Mohammad, D., P.B. Cox, G.L. Posler, M.B. Kirkham, A. Hussain and S. Khan. 1993. Correlation of characters contributing to grain and forage yields and forage quality in sorghum (*Sorghum bicolor*). Indian J. Agric. Sci. 63:92-95.
- Mohammed, M. 2009. Line x tester analysis across locations and years in Sudanese x exotic lines of forage sorghum. J. Plant Breeding Crop Sci. 1:311-319.
- Mohammed, M.I., and N. H. Talib. 2008. Heterosis and combining ability for quality traits in fodder sorghum. Aust. J. Basic Applied Sci. 2:99-104.
- Morishige, D.T., P.E. Klein, J.L. Hilley, S.M.E. Sahraeian, A. Sharma, and J.E. Mullet. 2013. Digital genotyping of sorghum: a diverse plant species with a large repeat-rich genome. BMC Genomics 14:448.
- Murty, B.R., and J.N. Govil. 1967. Description of 70 groups in genus sorghum based on a modified Snowden's classification. Indian J. Genet. 27:75-91.
- Musick, J.T., and D.A. Dusek. 1980. Irrigated corn yield response to water. Transaction ASAE 110:92-98, 103.
- Musick, J.T., and J.D. Walker. 1987. Irrigation practices for reduced water application-Texas High Plains. Applied Eng. in Agri. 3:190-195.
- Musick, J.T., F.B. Pringle, and J.J. Walker. 1988. Sprinkler and furrow irrigation trends: Applied Eng. in Agri. 4:46-52.
- Musick, J.T., F.B. Pringle, W.L. Harman, and B.A. Stewart. 1990. Long-term irrigation trends-Texas High Plains. Applied Eng. in Agric. 6:717-724.
- Naik, V.R., H. Shivanna, M.S. Joshi, and K.G. Parameswarappa. 1994. Heterosis and combining ability analysis in sorghum. J. Maharashtra Agric. Univ. 19:137-138.

- Nandanwankar, K.G. 1990. Heterosis studies for grain yield characters in *rabi* sorghum [*Sorghum bicolor* (L.) Moench]. Indian J. Genet. Plant Breeding 50:83-85.
- NASS. National Agricultural Statistics Service. 2011. https://www.nass.usda.gov/Publications/Todays_Reports/reports/cpvl0212.pdf
- NASS. National Agricultural Statistics Service. 2012. www.nass.usda.gov/Publications/TODAYRPT/cropan15.pdf
- NASS. National Agricultural Statistics Service. 2015. <http://www.usda.gov/nass/PUBS/TODAYRPT/cropan15.pdf>
- NASS. National Agricultural Statistics Service. 2016. <http://www.usda.gov/nass/PUBS/TODAYRPT/cropan16.pdf>
- National Research Council. 1996. Lost crops of Africa. National Academy Press, Washington, DC.
- Nativ, R., and D.A. Smith. 1987. Hydrogeology and geochemistry of the Ogallala Aquifer, southern High Plains. J. Hydrology 91:217-253.
- Nayakar, N.Y., B.T.S. Gowda, and J.V. Goud. 1989. Combining ability analysis for panicle components in *rabi* sorghum [*Sorghum bicolor* (L.) Moench]. Mysore J. Agric. Sci. 23:306-310.
- Nayeem, K.A. 1992. Genetic architecture of protein, lysine and sugar contents in grain sorghum. Ann. Agric. Res. 13:57-60.
- Nayeem, K.A., and D.R. Bapat. 1984. Combining ability in grain sorghum. Indian J. Genet. Plant Breeding 44:353-357.
- Nimbalkar, V.S., D.R. Bapat, and R.C. Patil. 1988. Components of grain yield in sorghum. J. Maharashtra Agric. Univ. 13:206-207.
- Norwood, C.A. 2000. Water use and yield of limited-irrigated and dryland corn. Soil Sci. Soc. Am. J. 64:365-370.
- Ogallala Aquifer. 2004. Annual report: crop irrigation and production.
- Omar, K.A., K.M. Mahmoud, and M.E. Mohamed. 2014. Heterosis in grain sorghum under two levels of nitrogen fertilizer. Middle East J. Agric. Res. 3:176-185.
- Patel, P.I., R.H. Patel, and M.S. Desai. 1990. Heterosis and combining ability of grain sorghum [*Sorghum bicolor* (L.) Moench] at different locations in Gujarat. Indian J. Agric. Sci. 60:382-386.

- Patel, R.H., K.B. Desai, and M.S. Desai. 1991. Note on combining ability analysis for yield and its components in grain sorghum. Gujarat Agric. Univ. Res. J. 17:113-115.
- Pederson, J.F., and W.L. Rooney. 2004. Sorghum. In: L.E. Mosher, B.L. Burson, and L.E. Sollenberger [eds.], Warm-season (C4). ASA/CSSA/SSSA, Madison, WI.
- Pillai, M.A., P. Rangaswamy, N. Nadarjan, C. Vannirajan, and J. Ramalingam. 1995. Combining ability analysis for ear head characters in sorghum. Indian J. Agric. Res. 29:98-102.
- Poehlman, J.M. 1987. Breeding sorghum and millet. In: Breeding field crops. Springer, Netherlands. p. 508-555.
- Prabhakar, E.M., and D.M. Dahadure. 2013 Combining ability of new parental lines for flowering, maturity, and grain yield in *rabi* sorghum. Electronic J. Breeding 4:1214-1218.
- Prakash, R., K. Ganesamurthy, A. Nirmalakumari, and P. Nagarajan. 2010. Combining ability for fodder yield and its components in sorghum (*Sorghum bicolor* L.). Electronic J. Plant Breeding 1:124-128.
- Premalatha, N., N. Kumaravadivel, and P. Veerabadhiran. 2006. Heterosis and combining ability for grain yield and its components in grain sorghum [*Sorghum bicolor* (L.) Moench]. Indian J. Genet. Plant Breeding 66:123-126.
- Prihar, S.S., and B.A. Stewart. 1990. Using the upper-bound slope through origin to estimate genetic harvest index. Agron. J. 82:1160-1165.
- Prihar, S.S., and B.A. Stewart. 1991. Sorghum harvest index in relation to plant size, environment, and cultivar. Agron. J. 83:603-608.
- Quinby, J.R. 1963. Manifestation of hybrid vigor in sorghum. Crop Sci. 3:288-291.
- Quinby, J.R. 1974. Sorghum improvement and the genetics of growth. Texas A&M University Press, College Station, TX.
- Quinby, J.R. 1980. Interaction of genes and cytoplasm in male sterility in sorghum. In: H.D. Loden and D. Wilkinson [ed.] Proc. 35th Ann. Corn and Sorghum Ind. Res. Conf., Chicago, IL. 9-11 December 1980, Am. Seed Trade Assoc., Washington, DC. p. 175-184
- Quinby, J.R., and R.E. Karper. 1954. Inheritance of height in sorghum. Agron. J. 46:211-216.
- Quinby, J.R., and J.H. Martin. 1954. Sorghum improvement. Adv. Agron. 6:305-359.

- Rahimi, R., B. Rabiei, H. Samizadeh, and A. Kafi Ghasemi. 2010. Combining ability and heterosis in rice cultivars. *J. Agric. Sci. Technol.* 12:223-231.
- Rani, C.P., C.Aruna, and F. Jabeen. 2015. Studies on heterosis and combining ability for yield components in grain sorghum [*Sorghum bicolor* (L.) Moench]. *Madras Agric. J.* 102:108-114.
- Rao, M.R.G., S.J. Patil, and R. Parameshwarappa. 1993. Heterosis and heterobeltiosis for development and panicle characters in *rabi* sorghum. *Mysore J. Agric. Sci.* 27:223-228.
- Rao, S.K., A.K. Gupta, S.S. Baghel, and S.P. Singh. 1982. Combining ability analysis of grain quality in sorghum. *Indian J. Agric. Res.* 16:1-9.
- Rashid, M., A.A. Cheema, and M. Ashraf. 2007. Line x tester analysis in basmati rice. *Pakistan J. Bot.* 39:2035-2042.
- Raut, S.K., P.H. Patel, and P.W. Khorgade. 1992. Path analysis of yield components in sorghum. *Agric. Sci. Digest, Karnal* 12:172-174.
- Reddy, J.N., and P. Joshi. 1993. Heterosis, inbreeding depression and combining ability analysis in sorghum [*Sorghum bicolor* (L.) Moench]. *Indian J. Genet.* 53:138-146.
- Riday, H., E.C. Brummer, T.A. Campbell, D. Luth, and P.M. Cazcarro. 2003. Comparisons of genetic and morphological distance with heterosis between *Medicago sativa* subsp. *sativa* and subsp. *falcata*. *Euphytica* 131:37-45.
- Rojas, B.A., and G.F. Sprague. 1952. A comparison of variance components in corn yield trials: III. General and specific combining ability and their interaction with locations and years. *Agron. J.* 44:462-466.
- Rooney, W.L. 2000. Genetics and cytogenetics. In: C.W. Smith and R.A. Frederiksen [eds.], *Sorghum: origin, history, technology, and production*. John Wiley and Sons, New York. p. 261-307.
- Rooney, W.L. 2004. Sorghum improvement-integrating traditional and new technology to produce improved genotype. *Adv. Agron.* 83:37-109.
- Rooney, W.L., and S. Aydin. 1999. The genetic control of a photoperiod sensitive response in sorghum. *Crop Sci.* 39:397-400.
- Rosenow, D.T., and J.A. Dahlberg. 2000. Collection, conversion, and utilization of sorghum. In: C.W. Smith and R.A. Frederiksen [eds.], *Sorghum: origin, history, technology, and production*. John Wiley and Sons, New York. p. 309-328.

- Rosenthal, W.D., R.L. Vanderlip, B.S. Jackson, and G.F. Arkin. 1989. A grain sorghum growth model. TAES. Comput. Software. Documentation Ser. MP-1669. Texas Exp. Station, College Station.
- Ross, W.M., K.D. Kofoed, J.W. Maranville, and R.L. Voigt. 1981. Selection for grain protein and yield in sorghum random mating populations. *Crop Sci.* 21:774-777.
- Saghai-Maroo, M.A., G.P. Yang, Q. Zhang, and K.A. Gravois. 1997. Correlation between molecular marker distance and hybrid performance in U.S. southern long grain rice. *Crop Sci.* 37:145-150.
- Salini, K., V. Satyanarayana Rao, J.V. Ramana, and V. Srinivasa Rao. 2008. Line x tester analysis in dual purpose sorghum [*Sorghum bicolor* (L.) Moench]. *Andhra Agric. J.* 55:166-174.
- Salunke, C.B., and G.N. Deore. 1998. Heterosis in *rabi* sorghum for grain yield and its components. *Madras Agric. J.* 87:143-144.
- Sanchez-Diaz, M.F., and P.J. Kramer. 1971. Behavior of corn and sorghum under water stress and during recovery. *Plant Physiol.* 48:613.
- Sankarapandian, R., D. Krishnadoss, and A.A. Devarathinam. 1996. Genetic parameters, correlations and path analysis among yield and yield characters in grain sorghum. *Madras Agric. J.* 83:625-628.
- Sarker, U., P.S. Biswas, B. Prasad, and M.A. Khaleque Mian. 2002. Heterosis and genetic analysis in rice hybrids. *Pak. J. Biol. Sci.* 5:1-5.
- Schertz, K.F. 1977. Registration of A2Tx2753 and BTx2753 sorghum germplasm (Reg. no. GP30 and 31). *Crop Sci.* 17:983.
- Schertz, K.F., and J.M. Ritchey. 1978. Cytoplasmic-genetic male-sterility systems in *Sorghum*. *Crop Sci.* 18:890-893.
- Schertz, K.F., and D.R. Pring. 1982. Cytoplasmic sterility systems in sorghum. In: J.V. Martin [ed.], *Sorghum in the eighties*. International Crops Research Institute for the Semi-Arid Tropics, Patancheru, India. p. 373-383.
- Shahane, T.G., and D.R. Bapat. 1981. Combining ability and stability studies in sorghum. *Sorghum Newsl.* 24:17.

- Shankaregowda, B.T., M. Rao, and S.W. Mensinkai. 1972. Heterosis and line x tester analysis of combining ability in selected lines of sorghum (*Sorghum vulgare*) 11. Combining ability. Mysore J. Agric. Sci. 6:242-252.
- Sharma, H., and V. Sharma. 2006. Heterosis in sorghum [*Sorghum bicolor* (L.) Monech]. Agric. Sci. Digest 26:245-248.
- Shinde, V.K., and S.M. Sudewad. 1981. Line x tester analysis of combining ability in sorghum (*Sorghum bicolor* (L.) Wench). J. MAU 6:215-218.
- Shull, G.H. 1908. The composition of a field of maize. Rep. Am. Breed. Assoc. 4:296-301.
- Shull, G.H. 1952. Beginnings of the heterosis concept. In: J.W. Gowen [ed.], Heterosis. Iowa State College Press, Ames, Iowa. p. 14-48.
- Siddiqui, M.A., and K.S. Baig. 2001. Combining ability analysis for yield and its components characters in sorghum [*Sorghum bicolor* (L.) Moench]. J. Res. ANGRAU 29:27-34.
- Singhanian D.L. 1980. Heterosis and combining ability studies in grain sorghum. Indian J. Genet. 40:463-471.
- Smith, C.W. 1995. Crop production: evolution, history, and technology. John Wiley and Sons, New York.
- Smith, C.W., and R.A. Frederiksen. 2000. History of cultivar development in the United States: from "Memoirs of A.B. Maunder – Sorghum Breeder. In: C.W. Smith and R.A. Frederiksen [eds.], Sorghum: origin, history, technology, and production. John Wiley and Sons, New York. p. 191-223.
- Snowden, J. D. 1936. The cultivated races of sorghum. Adlard.
- Soltani, A., A.M. Rezai, and M.R.K. Pour. 2001. Genetic variability of some physiological and agronomic traits in grain sorghum (*Sorghum bicolor* L.). J. Sci. Tech. Agric. Nat. Resources 5:127-137.
- Spiegel, B. 2015. Grain sorghum may be more profitable than corn in High Plains. http://www.agriculture.com/crops/other-crops/sorghum/grain-sorghum-may-be-more-profitable-th-cn_153-ar51472 (accessed 28 June 2017).
- Sprague, G.F. 1946. Early testing of inbred lines of corn. J. Am. Soc. Agron. 38:108-117.

Sprague, G. F., and L.A. Tatum. 1942. General versus specific combining ability in single crosses of corn. *J. Am. Soc. Agron.* 34:923-32.

Staggenborg, S. 2013. Corn vs. grain sorghum in water limited scenarios. CropQuest. <https://www.cropquest.com/2013/01/01/corn-vs-grain-sorghum-in-water-limited-scenarios/> (accessed 28 June 2017).

Staggenborg, S. 2015. Grain sorghum may be more profitable than corn in High Plains. http://www.agriculture.com/crops/other-crops/sorghum/grain-sorghum-may-be-more-profitable-th-cn_153-ar51472 (accessed 28 June 2017).

Steel, R.G.D., and J.H. Torrie. 1980. Principles and procedures of statistics. 2nd ed. McGraw-Hill, New York.

Stephens, J.C., and R.F. Holland. 1954. Cytoplasmic male sterility for hybrid sorghum seed production. *Agron. J.* 46:20-23.

Stephens, J.C., F.R. Miller, and D.T. Rosenow. 1967. Conversion of alien sorghums to early combine genotypes. *Crop Sci.* 7:396.

Stewart, B.A., D.A. Dusek, and J.T. Musick. 1981. A management system for the conjunctive use of rainfall and limited irrigation of graded furrows. *Soil Sci. Am. J.* 45:413-419.

Stewart, B.A., J.T. Musick, and D.A. Dusek. 1983. Yield and water use efficiency of grain sorghum in a limited irrigation-dryland farming system. *Agron. J.* 75:629-634.

Stewart, B.A., and E. Burnett. 1987. Water conservation technology in rainfed and dryland agriculture. In: W.R. Jordan [ed.], *Water and water policy in world food supplies*. Texas A&M University Press, College Station. p. 335-359.

Stone, L. 2013. Corn vs. grain sorghum in water limited scenarios. CropQuest. <https://www.cropquest.com/2013/01/01/corn-vs-grain-sorghum-in-water-limited-scenarios/> (accessed 28 June 2017).

Stone, L.R., and A.J. Schlegel. 2006. Crop water use in limited-irrigation environments. <http://irrigationtoolbox.com/ReferenceDocuments/TechnicalPapers/CPIA/2006/Stone06.pdf> (accessed 2 July 2017).

Stout, D.G., and G.M. Simpson. 1978. Drought resistance of *Sorghum bicolor* 1. Drought avoidance mechanisms related to leaf water status. *Can. J. Plant Sci.* 58:213.

- Stubber, C.W., S.E. Lincoln, D.W. Wolff, T. Helentjaris, and E.S. Lander. 1992. Identification of genetic factors contributing to heterosis in a hybrid from two elite maize inbred lines using molecular markers. *Genet.* 132:823-839.
- Subedi, K.D., B.L. Ma, and D.L. Smith. 2006. Response of a leafy and non-leafy maize hybrid to population densities and fertilizer nitrogen levels. *Crop Sci.* 46:1860-1869.
- Subedi, K.D., and B.L. Ma. 2009. Assessment of some major yield-limiting factors on maize production in a humid temperate environment. *Field Crops Res.* 110:21-26.
- Sunku, S.S.K., M.B. Reddy, and P.R.R. Reddy. 2002. Character association and path analysis in grain sorghums (*Sorghum bicolor* L. Moench) *vis-a-vis* the sudan grasses (*Sorghum sudanense*). *Forage Res.* 28:42-45.
- Tadesse, T., T. Tesso, and G. Ejeta. 2008. Combining ability of introduced sorghum parental lines for major morpho-agronomic traits. *J. SAT Agric. Res.* 6:1-7.
- Tariq, A.S., Z. Akram, G. Shabbir, K.S. Khan, and M.S. Iqbal. 2014. Heterosis and combining ability studies for quantitative traits in fodder sorghum (*Sorghum bicolor* L.). *J. Agric. Res.* 52:329-337.
- Taurchi, M., and A.M. Rezai. 1997. Correlation between traits and path analysed for grain yield in sorghum. *Iranian J. Agric. Sci.* 28:73-86.
- Teetes, G.L., K.V. Seshu Reddy, K. Leuschner, and L.R. Hossner. 1983. Sorghum insect identification handbook. Information Bulletin 12. International Crops Research Institute for the Semi-Arid Tropics, Patancheru, India.
- Teetes, G.L., and B.B. Pendleton. 2000. Insect pests of sorghum. In: C.W. Smith and R.A. Frederiksen [eds.], *Sorghum: origin, history, technology, and production*. John Wiley and Sons, New York. p. 443-495.
- Tiwari, D.K., R.S. Gupta, and R. Mishra. 2003. Genotypic and phenotypic correlations in grain sorghum. *Plant Archives* 3:283-286.
- Tolk, J.A., and T.A. Howell. 2008. Field water supply: yield relationships of grain sorghum grown in three USA Great Plains soils. *Agric. Water Management* 95:1303-1313.
- Troyer, A.F., and E.J. Wellin. 2009. Heterosis decreasing in hybrids yield test inbreds. *Crop Sci.* 49:169-1976.

- Undersander, D.J. 1986. Management of sorghum under limited irrigation. *Agron. J.* 78:28-32.
- Unger, P.W. 1972. Dryland winter wheat and grain sorghum cropping systems northern High Plains of Texas. Pamphlet B-1126. Texas A&M Univ., College Station.
- Unger, P.W. 1988. Grain and forage sorghum production with no-tillage on dryland. *Agron. J.* 80:193-197.
- Unger, P.W., and F.B. Pringle. 1981. Pullman soil: distribution importance, variability, and management. Bull. B-1372. Texas Agri. Exp. Station, College Station.
- U.S. Grains. 2016. <http://www.grains.org/buyingselling/sorghum> (accessed 25 February 2016).
- USDA. National Agricultural Statistics Service. 2007. USDA National Agricultural Statistics Service, Washington, DC.
- USDA-ERS. 2011. United States Department of Agriculture: Economic Research Services Food Consumption Database. <http://www.ers/usda.gov/data/foodconsumption> (accessed 4 January 2017).
- Vanderlip, R.L., and H.E. Reeves. 1972. Growth stages of sorghum. *Agron. J.* 64:13-17.
- Veerabathiran, P., S. Palanisamy, and G.A. Palanisamy. 1994. Association analysis in grain sorghum [*Sorghum bicolor* (L.) Monech]. *Madras Agric. J.* 81:532-534.
- Warkad, Y.N., R.T. Tidke, N.M. Maske, A.V. Kharde, and N.R. Potdukhe. 2010. Character association and path analysis in sorghum [*Sorghum bicolor* (L.) Monech]. *Int. J. Agric. Sci.* 6:100-104.
- White, J., K. Bolsen, and B. Kirch. 1988. Relationship between agronomic and silage quality traits of forage sorghum cultivars. *Agric. Ex. Stn. Rep. Prog.* 539, Kansas State University, Manhattan.
- White, J.S., K.K. Bolsen, G. Posler, and J.W. Neill. 1991. Forage sorghum silage dry matter disappearance as influenced by plant part proportion. *Animal Feed Sci. Technol.* 33:313-322.
- Widdicombe, W.D., and K.D. Thelen. 2002. Row width and plant density effect on corn forage hybrids. *Agron. J.* 94:326-330.
- Wilson, N.D., D.E. Weibel, and R.W. McNew. 1978. Diallel analyses of grain yield, percent protein, and protein yield in grain sorghum. *Crop Sci.* 18:491-495.

- Wright, S., J. Edwards, C. Godsey, J.D. Vitale, F.M. Epplin, and R. Taylor. 2010. Conservation tillage in Oklahoma: perceptions and demographics of producers. Oklahoma Cooperative Extension Service and Division of Agricultural Sciences and Natural Resources, Oklahoma State University, Stillwater. E-1019.
- Xiao, J., J. Li, L. Yuan, and S.D. Tanksley. 1995. Dominance is the major genetic basis of heterosis in rice as revealed by QTL analysis using molecular markers. *Genet.* 140:745-754.
- Yadav, R., and S.K. Pahuja. 2007. Combining ability for fodder yield and its components in forage sorghum. *Forage Res.* 32:220-223.
- Yu, S.B., J.X. Li, C.G. Xu, Y.F. Tan, Y.J. Gao, X.H. Li, Q. Zhang, and M.A.S. Maroof. 1997. Importance of epistasis as the genetic basis of heterosis in an elite rice hybrid. *Proc. Natl. Acad. Sci. USA* 94:9226-9231.
- Zhang, Q.F., Y.J. Gao, S.H. Yang, R.A. Ragab, M.A. Saghai, and Z.B. Li. 1994. A diallel analysis of heterosis in elite hybrid rice based on RFLPs and microsatellites. *Theor. Appl. Genet.* 89:185-192.
- Zhang, Q.F., Y.J. Gao, M.A.S. Saghai, S.H. Yang, and J.X. Li. 1995. Molecular divergence and hybrid performance in rice. *Mol. Breed.* 1:133-142.
- Ziyomo, C., and R. Bernardo. 2013. Drought tolerance in maize: indirect selection through secondary traits versus genomewide selection. *Crop Sci.* 53:1269-1275.